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Development and comparison of two high accuracy methods for uranium concentration in nuclear fuel: ID-TIMS and K-edge densitometry

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23 Abstract

This study compares the two analytical methods for uranium concentration determination 24 25 with high accuracy in uranium pellet: K-edge densitometer (KED) and the isotope dilution 26 with Thermal Ionisation Mass Spectrometry measurements (ID-TIMS). Both techniques 27 are compared in terms of time, generated radioactive effluent, simplicity, uncertainty 28 estimation and detection limit. ID-TIMS shows lower detection limit and uncertainties than 29 KED. However, the KED analysis time is shorter and generates less effluent. Both 30 techniques were used for metrological analysis of uranium concentration in nuclear 31 materials. The optimization of sample spike mixture isotope ratio for ID-TIMS to decrease 32 uncertainties is also discussed.

33 Keywords

34 Uranium concentration; TIMS; K-edge densitometer; isotope dilution

36 Introduction

Uranium detailed accountancy is of prime interest in the nuclear industry, at various stages of the nuclear fuel cycle and for safeguards purposes [1–3]. ATALANTE is a nuclear facility of the French Alternative Energies and Atomic Energy Commission (CEA) and dedicated to the research on the nuclear spent fuel reprocessing. Studies such as nuclear fuel dissolution, actinides separation and precipitation are performed, requiring to determine the uranium concentration with high accuracy.

43 Among the specific analytical techniques available in such facility, K-edge densitometry (KED) is an effective technique to determine uranium assay in nuclear fuel dissolution 44 45 solution [4, 5]. KED is an X-ray absorbance spectrometry technique based on the Beer-46 Lambert's law. The uranium concentration in solution is proportional to the X-ray attenuation. High uranium concentrations, typically above 50 g L⁻¹, can be measured with 47 48 high accuracy. For lower concentration, spectral discontinuities make the spectrum process 49 difficult and uncertainties increase significantly. K-edge spectrometry is a nondestructive, 50 non-invasive, relatively fast technique and requires little sample preparation.

51 Isotope Dilution (ID) is a well-known method for quantification based on internal 52 calibration [6, 7]. The ID principle is to mix a known amount of sample containing a known 53 isotope composition with a spike solution containing a known concentration and isotope 54 composition. The spike solution contains the same analyte as the sample with a different isotope composition. The isotope ratio of the sample – spike mixture reflects the sample 55 56 analyte concentration. The ID methodology requires only isotope ratio measurements and 57 knowing masses involved in the sample – spike mixture. As it is based on isotope ratio 58 determination, an accurate technique for isotope ratio measurements must be preferred. 59 Thermal-Ionization Mass Spectrometry (TIMS) is the reference technique for the accurate 60 determination of uranium isotope ratios [8–13]. The ID methodology coupled to TIMS for 61 the isotope ratio measurements (ID-TIMS) is then a powerful combination for the uranium 62 concentration determination.

The present study was performed in the context of the "2017 Nuclear Material Round Robin" organized by the International Atomic Energy Agency (IAEA) which aims at determining the uranium isotope ratio and mass fraction in nuclear materials. The isotope ratios determination was presented in a previous paper [10]. This paper presents a comparison between the ID-TIMS and the K-edge densitometry for the uranium concentration determination with high accuracy in a uranium pellet.

69 **Experimental**

70 Materials, reagents and certified reference materials

All solutions were prepared in polypropylene vials, except for uranium solutions, which were prepared in PFA vials. 1 mol L⁻¹, 3 mol L⁻¹ and 8 mol L⁻¹ nitric acid solutions were prepared by diluting high purity nitric acid (Merck, Suprapur) in deionized water (resistivity: 18 2 M Ω .cm). A high accurated scale (Mettler-Toledo, WXTP 205) was used to prepare all solutions.

The spike solution used in the ID methodology was the IRMM 054 certified reference material (CRM) provided by the Joint Research Centre of the European Commission (EC-JRC). This CRM contains high amounts of the 235 U isotope and is certified for the concentration and the isotope ratios. This solution was diluted gravimetrically in order to obtain a uranium concentration near 10 µg g⁻¹.

81 Sample preparation

The inter-laboratory comparison (ILC) sample is a low enrichment uranium oxide pellet. The K-edge densitometer available in the laboratory and the TIMS require working with solutions. The ILC uranium pellet was dissolved in hot nitric acid (8 mol L⁻¹). This solution (pellet + nitric acid) was heated to 135 °C in a PFA vial until complete dissolution. The dissolution solution had about 250 g L⁻¹ uranium concentration [10]. 87 Instrumental

88 Thermal Ionization Mass Spectrometer

The Thermo Fisher Triton and VG Sector 54 TIMS used for the experiments and the deposit technique were previously described in detail [14]. The measurements were performed using Faraday cups coupled to $10^{11} \Omega$ current amplifiers (hereafter referred to as FC 11). All the isotope ratio measurements were performed with the total evaporation method (TE method) described previously [10]. The ²³⁸U⁺ ion beam target intensity was set at 1 V and 2.2 V for the VG Sector 54 and for the Triton TIMS, respectively.

95 The uranium concentration ([U]_s) was determined using isotope dilution according to Eq.
96 (1):

$$[\mathbf{U}]_{\mathrm{S}} = [\mathbf{U}]_{\mathrm{T}} \cdot \frac{\mathrm{m}_{\mathrm{T}}}{\mathrm{m}_{\mathrm{S}}} \cdot \frac{\mathrm{M}_{\mathrm{S}}}{\mathrm{M}_{\mathrm{T}}} \cdot \frac{(235)_{\mathrm{T}}}{(238)_{\mathrm{S}}} \cdot \frac{\mathrm{R}_{\mathrm{Mix}} - \mathrm{R}_{\mathrm{T}}}{1 - \mathrm{R}_{\mathrm{Mix}} \cdot \mathrm{R}_{\mathrm{S}}}$$
(1)

Where *T* refers to the spike (or tracer), *Mix* refers to the sample - spike mixture and *S* refers to the sample; *m* are the masses involved in the mixture, *M* are the atomic weights, (235) and (238) are the isotope abundances. R_{mix} and R_T correspond to the ²³⁸U/²³⁵U mixture and spike isotope ratio, respectively. R_S corresponds to the ²³⁵U/²³⁸U sample isotope ratio. The isotopic composition of the pellet (needed for the [U]_S calculation by ID-TIMS) was determined with high accuracy measurements in the previous study [10].

103 The uranium mass fraction determination by ID-TIMS was performed using the two TIMS 104 available in the laboratory with 3 different determinations. First, 3 diluted dissolution solutions ([U] $\approx 10 \ \mu g \ g^{-1}$ and hereafter referred to as diluted solution 1, 2 or 3) were 105 106 prepared gravimetrically from the initial dissolution solution ([U]_i ≈ 250 g.L⁻¹). Then, 5 107 (diluted solution 1 – spike) mixtures, 3 (diluted solution 2 – spike) mixtures and 2 (diluted 108 solution 3 – spike) mixtures were prepared gravimetrically. Each mixture was evaporated 109 after preparation, and then dissolved again in 10 μ L of 1 mol L⁻¹nitric acid. 1 μ L (about 400 ng of U) of this final solution was deposited onto a filament for each individual 110

111 measurement. The ${}^{238}U/{}^{235}U$ isotope ratio of each mixture was analyzed using the TE 112 method. The ${}^{238}U/{}^{235}U$ ratios ranged from 1.08 to 1.23. The 5 (diluted solution 1 – spike) 113 mixtures were analyzed using the VG Sector 54 TIMS. The uranium concentration was the 114 average of the 5 individual measurements. The (diluted solution 2 and 3 – spike) mixtures 115 were analyzed using the Triton TIMS and the respective uranium concentrations were 116 determined by computing the average of each respective individual measurements. Finally, 117 the uranium mass fraction of the pellet was the average of the 3 groups of determination.

118 *K-edge densitometry*

119 The nuclearized KED used for the experiments was previously described in details [4]. The 120 X-ray generator and X-ray tube (W anode, MG165) are marketed by Yxlon. The tube 121 configuration used in the laboratory is 150 kV and 10 mA for assay sample measurements, 122 or 5 mA for blank reference solution measurements. The detectors are hyper-pure Ge 123 crystals (Canberra EGX 200-10, active area 200 mm²) cooled with liquid nitrogen. The 124 acquisition electronics are Canberra DSA-1000 coupled with Genie 2000 software. The 125 spectral data were processed using a CEA developed software using standardless 126 algorithms [4]. Measurements are performed without any calibration except for energy. 127 The typical counting time was 1000 s.

This device is located in the back area of the analysis hot cell line of ATALANTE. The sample, packaged in small polypropylene vials, is transferred *via* a pneumatic line from the hot cell to the KED. The minimum volume required to ensure the X-rays are absorbed by the sample in this system is about 1.5 mL.

The dissolution solution was analyzed by KED without any further preparation except vial transfer. The result was obtained from the average of 5 individual measurements of the same vial containing the dissolution solution. Between each individual measurement, the vial was returned to the hot cell line before a new transfer back to the KED in order to take into account the uncertainties due to the vial position in the KED. The concentration determined using the KED system was in g L⁻¹. In order to obtain the uranium mass fraction

in the pellet, the dissolution solution density is necessary and was performed with anAnton-Paar DMA 58 density meter.

140 Results evaluation and uncertainties estimation

- 141 *Results evaluation*
- 142 The bias was calculated using Eq. (2).

Bias (%) =
$$\frac{[U] - \text{cert}}{\text{cert}} \cdot 100$$
 (2)

Where [U] is the uranium concentration determined by ID-TIMS or KED and *cert* is theILC assigned value.

145 The precision was evaluated by computing the Relative Standard Deviation (RSD) of all146 the measurements.

Eq. (3) was used to determine if the analytical method has a statistically significant bias
[15]. If the normalized bias (*NB*) was lower than 2, the method was considered having no
statistically significant bias.

$$NB = \frac{|[U] - cert|}{\sqrt{u^2([U]) + u_{cert}^2}}$$
(3)

150 Where u([U]) is the uranium concentration uncertainty determined by ID-TIMS or KED at 151 k = 1 and u_{cert} is the assigned value uncertainty at k = 1.

152

154 Uranium concentration uncertainty using ID-TIMS

The uranium concentration uncertainty ($u([U]_s)$, at k = 1), determined by ID-TIMS shown in Eq. (4), was estimated by combining the uncertainties from each term of Eq. (1). The terms of Eq. (1) were considered as not correlated.

$$\begin{split} u^{2}([U]_{S}) &= \left(\frac{m_{T}}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \cdot R_{S}}\right)^{2} \cdot u^{2}([U]_{T}) \\ &+ \left([U]_{T} \cdot \frac{1}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \times R_{S}}\right)^{2} \cdot u^{2}(m_{T}) \\ &+ \left(-[U]_{T} \cdot \frac{m_{T}}{m_{S}^{2}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \cdot R_{S}}\right)^{2} \cdot u^{2}(m_{S}) \\ &+ \left([U]_{T} \cdot \frac{m_{T}}{m_{S}} \cdot \frac{1}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \cdot R_{S}}\right)^{2} \cdot u^{2}((235)_{T}) \\ &+ \left(-[U]_{T} \cdot \frac{m_{T}}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \cdot R_{S}}\right)^{2} \cdot u^{2}((235)_{T}) \\ &+ \left([U]_{T} \cdot \frac{m_{T}}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}^{2}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \cdot R_{S}}\right)^{2} \cdot u^{2}((238)_{S}) \\ &+ \left([U]_{T} \cdot \frac{m_{T}}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{R_{Mix} - R_{T}}{1 - R_{Mix} \cdot R_{S}}\right)^{2} \cdot u^{2}(R_{S}) \\ &+ \left(-[U]_{T} \cdot \frac{m_{T}}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{(R_{Mix} - R_{T}) \cdot R_{Mix}}{(1 - R_{Mix} \cdot R_{S})^{2}}\right)^{2} \cdot u^{2}(R_{T}) \\ &+ \left([U]_{T} \cdot \frac{m_{T}}{m_{S}} \cdot \frac{(235)_{T}}{(238)_{S}} \cdot \frac{M_{S}}{M_{T}} \cdot \frac{(1 - R_{Mix} \cdot R_{S})^{2}}{(1 - R_{Mix} \cdot R_{S})^{2}}\right)^{2} \cdot u^{2}(R_{Mix}) + u^{2}(\text{dilution}) \\ &+ u^{2}(\text{precision}) \end{split}$$

In Eq. (4), the molar masses uncertainties were considered negligible and were not taken into account for the final uncertainty computation. The term u(precision) is the method precision, corresponding to the standard deviation of the 3 groups of determination. The term u(dilution) is the sample dilution uncertainty. The concentration ([U]_T), isotope ratio (R_T) and isotope abundance ((235)_T) related to the spike solution and their respective uncertainties were given in the CRM certificate (Table 1). The sample isotope ratio (R_S) and isotope abundance ((238)_S) were estimated in a previous study by the TE method for 165 the ${}^{235}\text{U}/{}^{238}\text{U}$ isotope ratio and the classical method with multi-dynamic sequences for the 166 ${}^{234}\text{U}/{}^{238}\text{U}$ and ${}^{236}\text{U}/{}^{238}\text{U}$ isotope ratios (required to compute the ${}^{238}\text{U}$ isotope abundance) 167 [10].

168 The sample (m_S) and spike (m_T) masses uncertainties involved in the mixture were estimated by taking into account the trueness, drift and resolution of the scale and the 169 170 method precision (Eq. (5)) [16]. The scale trueness and drift were estimated with a type B 171 evaluation and a uniform law (first term in Eq. (5)). This term corresponds to the maximum permissible errors (MPE) fixed by the laboratory divided by square root of 3. The scale 172 173 resolution (q) uncertainty was also estimated using a type B evaluation and a uniform law 174 (second term in Eq. (5)). The method precision, including scale precision and operator 175 effect, corresponds to the experimental standard deviation obtained from a series of n 176 measurements divided by the square root of n and was a type A evaluation (third term in Eq. (5)). The air buoyancy (Archimedes' principle) was taken into account for the masses 177 178 correction and its uncertainty was considered negligible.

$$u^{2}(m) = \left(\frac{MPE}{\sqrt{3}}\right)^{2} + \left(\sqrt{\frac{q^{2}}{12}}\right)^{2} + \left(\frac{s}{\sqrt{n}}\right)^{2}$$
(5)

The mixture isotope ratio uncertainty was estimated at 0.14 % (k = 2), corresponding to the International Target Value (ITV) for a ${}^{235}\text{U}/{}^{238}\text{U}$ isotope ratio of about 1 [17]. A similar uncertainty was previously estimated for a ${}^{235}\text{U}/{}^{238}\text{U}$ isotope ratio close to 1 in these working condition using the TE method with FC 11 [14].

183 Investigation on the optimum isotope ratio of the mixture for the184 ID-TIMS

The ID-TIMS require an optimum mixture isotope ratio (R_{Mix}), which can be determined theoretically [18]. The ID-TIMS parameters other than the mixture isotope ratio ([U]_T, m_T , m_S , (235)_T, (238)_S, R_S or R_T) depend only on the sample and spike characteristics and on

188 the mixture masses. These parameters were assumed constant for the theoretical study 189 hereafter described and equal to the values presented in Table 1.

- 190 Using Eq. (1) and the fixed parameters given in the Table 1, it is possible to compute using
- 191 a spreadsheet the sample concentration for a given mixture isotope ratio. This ${}^{238}U/{}^{235}U$
- mixture isotope ratio is between the sample (0.0581) and spike (50.87) isotope ratios.
- 193 Likewise, the sample concentration uncertainty can be computed using Eq. (4) for each mixture isotope ratio included between the ${}^{238}U/{}^{235}U$ sample and spike isotope ratios. The 194 195 *u*(*precision*) and *u*(*dilution*) terms were assumed to be constant in this theoretical study. 196 The *u*(precision) term was set at 0.06 % (k = 1), which is a value previously obtained in 197 our laboratory for actinide concentration determination by ID-TIMS [14]. The *u(dilution)* term was set at 0.034 %, which is an arbitrary value commonly observed in our laboratory 198 199 for this type of dilution. These 2 set values are not function of the mixture isotope ratio and 200 do not change the optimum determination. By combining Eq. (1) and (4), the concentration 201 relative uncertainty (u([U]/[U], k = 1)), as a function of the mixture isotope ratio, can be 202 drawn from the spreadsheet (Fig. 1). The optimum mixture isotope ratio is obtained when 203 the function reaches its minimum.

In our analysis conditions, the optimum mixture isotope ratio is obtained for a $^{238}U/^{235}U$ 204 205 isotope ratio of 1.7 (Fig. 1). Experimentally, it is possible to adjust the sample and spike 206 solution amount in the mixture to be closer to the optimum mixture isotope ratio. In 207 addition, this theoretical study shows that the sample concentration relative uncertainty is 208 almost stable around the optimum isotope ratio. It shows that the uncertainty increases 209 large when the mixture isotope ratio approaches the spike or sample isotope ratios. The 210 mixture isotope ratios from 1.08 to 1.23 (which are the ratios measured in this study), increase the concentration uncertainty only by 0.09 % to 0.18 % compared to the best 211 212 theoretically achievable uncertainty.

213 Uranium concentration uncertainty using KED analysis

214 The main parameters to take into account for the concentration uncertainty by KED 215 measurement are: the instrument variability (X-ray tube, detector), the vial position, the 216 counting time precision, the vial length, the sample matrix and the theoretical values of 217 mass attenuation coefficients. The uranium concentration relative uncertainty u([U])/[U]218 (k = 1) was estimated using Eq. (6). The first three terms correspond to the KED 219 measurement uncertainty. The fourth term is the density uncertainty of the dissolution 220 solution at k = 1 and also includes the density difference between the density and KED 221 measurements as the two measurements were not performed at the exact same temperature. 222 The first term of Eq. (6) takes into account the measurement precision component. It 223 includes the uncertainty from random effects (instrument variability, vial position and 224 counting time precision) and is given by the RSD of all the measurements. The second and 225 third terms take into account the systematic error components (vial length, sample matrix 226 and theoretical values of mass attenuation coefficients). The measurement trueness 227 $(u^{2}(truness)/(truness)^{2})$ is calculated with Eq. (7). The CRM were chosen so that they 228 respected different criteria: same element, similar analyte concentration and matrix. The 229 EQRAIN solutions, provided by the CEA/CETAMA during ILC were used [4]. They were chosen because: the analyte is the same, the uranium concentrations are close (about 250 230 and 360 g L^{-1} for the EQRAIN solutions and about 280 g L^{-1} for the dissolution sample) 231 232 and the matrix is the same (uranium only in HNO₃ media). Only the EORAIN solutions 233 analyzed using the same configuration and the same detector as the dissolution sample (as 234 the detector was changed in 2016) were used to compute the trueness term.

The term $u^2(ref)/(ref)^2$ corresponds to the CRM (EQRAIN U) concentration relative uncertainty at k = 1.

$$\frac{u^{2}([U])}{([U])^{2}} = \frac{u^{2}(\bar{x})}{\bar{x}^{2}} + \frac{u^{2}(\text{trueness})}{(\text{trueness})^{2}} + \frac{u^{2}(\text{ref})}{(\text{ref})^{2}} + \frac{u^{2}(\text{density})}{(\text{density})^{2}}$$
(6)

$$\frac{u(\text{trueness})}{\text{trueness}} = \frac{\text{Maximum bias on CRM}}{\sqrt{3}}$$
(7)

Results and discussion

238 ID-TIMS

239 The results are presented in Table 2. The RSD of the 3 groups of determinations is equal to 0.12 %. The result obtained for the uranium mass fraction of the pellet is 240 0.8835 ± 0.0028 g g⁻¹ (k = 2). The bias is equal to 0.30 %. The relative uncertainty, 241 computed with Eq. (4), is estimated at 0.32 % and is in compliance with the safeguard ITV 242 243 (0.36%, k=2) [17]. The relative contribution study (expressed in percent) of the main 244 uncertainty sources for the uranium mass fraction is given in Fig. 2. The spike 245 concentration uncertainty (0.04 %) has a very low contribution and is a minor source of 246 uncertainty (4.2 % of the total budget). The main uncertainty sources are the precision 247 (59.4 % of the total budget), the dilution to obtain a sample concentration of about 10 μ g g⁻ ¹ (19.7 % of the total budget) and the ²³⁸U/²³⁵U sample - spike mixture isotope ratio 248 249 determination (14.1 % of the total budget).

250 KED

251 A uranium concentration of 278.4 ± 2.1 g L⁻¹ (k = 2) was found for the dissolution solution. 252 Considering the dissolution solution density and the mass of the initial pellet before dissolution, the uranium mass fraction of the pellet is estimated to 0.8807 ± 0.0067 g g⁻¹ 253 254 (k = 2, Table 2). The RSD of the 5 individual determinations is 0.33 %. The bias is below 255 0.1 %. The uranium mass fraction determined using the KED system is in compliance with 256 the assigned value provided by the IAEA: the normalized bias is equal to 0.09. The relative 257 uncertainty is estimated to 0.76 % and is slightly higher than the ITV (0.56 %, k = 2) [17]. 258 The study of the main uncertainty sources shows the total uncertainty is mainly due to the 259 precision (76% of the total uncertainty). The trueness component (18% of the total 260 uncertainty) and the CRM reference uncertainty component (6 % of the total uncertainty) 261 have a minor contribution to the total uncertainty. The density uncertainty contribution is 262 negligible: below 0.2 % of the total uncertainty.

263 ID-TIMS and KED comparison

The ID-TIMS and the KED perform uranium assay with high accuracy. However, in terms of analytical performance, implementation, analysis time and generated radioactive effluent, they have different assets (Table 3).

267 The KED requires high uranium concentration solution in order to obtain the lowest uncertainties: uranium concentration higher than 50 g L^{-1} is needed to get relative expanded 268 uncertainties (k = 2) lower than 1%. The TIMS analyses require a lower uranium amount 269 270 than the KED: less than 1 µg of uranium is required for TIMS analyses. Solutions containing less than 1 mg L⁻¹ of uranium can be analyzed by ID-TIMS and the ITV can 271 still be achived (0.36 %, k = 2). The ID-TIMS gives directly the uranium mass fraction in 272 273 the nuclear materials, whereas the density determination for the KED system is required 274 beforehand. Both techniques showed no significant bias compared to the assigned value: 275 the normalized biases are below 2. The ID-TIMS uncertainty estimation (0.32 %, k = 2)was in compliance with the ITV and was lower than the KED uncertainty estimation 276 277 (0.76%, k = 2).

278 Both techniques require an identical pellet dissolution step. KED analysis can be performed directly on the dissolution solution (matrix 8 mol L⁻¹ HNO₃). No solution treatment, no 279 dilution and no calibration are required [4]. Cross contamination is then limited. As the 280 281 KED system is a nondestructive technique, the same aliquot can be used to measure uranium with the KED system and be used for others analyses like isotopic, radiometric or 282 283 density measurements (in some situations, taking into account evaporation loss is 284 necessary). High activity solution can be analyzed with the KED system as it is installed 285 in a shielded line. The TIMS can be installed in glove box environment but not in a hot 286 cell: high activity solutions require a dilution step in order to be suitable for working in 287 glove box. The dilution step is also required in order not to consume too much of the 288 solution spike. Indeed, for spiking the dissolution solution directly would have a 289 prohibitive cost as the spike solution quantity would be important. For very irradiating 290 samples such as high active liquid waste, the spiking step must be done in the hot cell 291 before the sample transfers.

292 As the analytical protocol is very different, both techniques present different analysis time. 293 For KED, about 5 minutes are required to prepare the sample in the analytical vial. The 294 analysis time is about 1 h 15 for 5 analysis of the same sample (about 15 min for 1 295 determination). Considering the fast analysis time of the KED system; the dissolution 296 solution density determination is the most critical step to the total analysis time (about 297 5 h 20). The ID-TIMS analysis overall process takes longer than the KED analysis. The 298 dilution step is performed in 1 h. The sample preparation, including sample - spike mixture 299 preparation, sample - spike mixture evaporation, deposit onto the filament, filament 300 loading inside the TIMS source take about 8 h in total. Then, the vacuum must be drawn 301 inside the source (about 4 h). This step is generally not critical in terms of operator working 302 time as the filament loading can be done at the end of a working day and the vacuum inside 303 the source can be obtain overnight. The analysis time of the 10 sample - spike mixtures 304 with the TE method requires about 9 h of work: Faraday cups calibration duration is about 305 10 min per day of use, heating filament about 10 min per analysis, signal optimization 306 about 5 min per analysis, the analysis time duration is about 30 min and the shutdown is 307 about 2 min per analysis. The ID-TIMS methodology requires also the sample isotope 308 determination. In total, we have calculated the analysis time for the sample isotope 309 determination with TIMS at 19 h [10]. The total ID-TIMS analysis time is estimated at 310 41 h, thus about 8 times higher for the ID-TIMS than KED.

311 The KED system does not consume the sample solution as it is a nondestructive technique 312 and generates no effluent as there is no dilution step. However, this technique consumes 313 sample solution indirectly as the density determination requires about 2 mL that cannot be 314 reused due to possible cross-contamination. The effluent generated for cleaning the density 315 meter represents about 10 mL. The total effluent generated was 12 mL, only due to density 316 determination. Moreover, the used KED system requires at least 1.5 mL of solution, which 317 can be a disadvantage when the available sample volume is low. The TIMS analyses need 318 a dilution step as uranium concentration is high (higher than 1 g.L⁻¹). In this study, 600 μ L 319 of the dissolution solution was used, corresponding to $3 \times 200 \ \mu\text{L}$ for the 3 determinations 320 by ID-TIMS. 200 µL of solution (about 300 mg of uranium) is enough to obtain an accurate 321 weight without generating too much nuclear effluent. The generated effluent was estimated 322 at 60 mL for one determination. In this study, the 3 determinations generated 180 mL of

- 323 effluent, only due to the dilution step. The ID-TIMS (0.6 mL) consumes less sample than
- 324 the KED system (2 mL required only for the density determination). However, the KED
- 325 system generates 15 times less volume of radioactive effluent.

326 Conclusions

327 This study presents a comparison between the ID-TIMS and the KED system for uranium 328 concentration determination with high accuracy. The results obtained with both techniques 329 are in compliance with the assigned value. Both techniques can be used for metrological 330 analysis of high uranium concentration while having different assets. The minimal 331 measurable concentration with ID-TIMS is lower than the KED's. The analysis time is 332 about 8 time higher for the ID-TIMS than for the KED. The ID-TIMS generates about 15 333 times more radioactive effluent. However, in our studies, only the estimated uncertainties 334 obtained with ID-TIMS are in compliance with the ITV. The estimated uncertainties 335 obtained with the KED are slightly higher.

336 The main approach to decrease the KED uncertainty, in order to be closer to the ITV, is to 337 improve the measurement precision as it is the main uncertainty source (76 % of the total 338 uncertainty budget). The vial position which may vary slightly from an analysis to another 339 in the KED system and the instrument variability (X-ray tube, detector) are the main 340 parameters influencing the method precision as well as the final uncertainty. An approach 341 to decrease the precision due to the instrument variability would be increasing the 342 measurement time. Even with a measurement time of 1 hour (15 min at present), the KED 343 would be faster than ID-TIMS. The precision due to the instrument variability would be 344 improved. However the precision due to the vial position would be the same whatever the 345 measurement time. The method used in the laboratory is a calibration free method, which 346 slightly increases the estimated uncertainties compared to methods using standard 347 calibration. A calibration would correct all physical phenomena including the ones that are 348 difficult to take into account in theoretical models. However, this correction would only 349 modify the method trueness. Therefore this would not change much the final uncertainty 350 as the trueness has a low contribution part in the final uncertainty (18% of the total

uncertainty). The KED data were also processed using the algorithms with fitting windows as proposed in [5] in order to show the influence on the repeatability. The repeatability were similar for either the algorithms using fitting windows (RSD = 0.47 %) or our inhouse algorithm (RSD = 0.33 %). In addition, the KED system used for this study is designed for dissolution or separation R&D studies that need quick results during the experiments. Our KED system, implemented in hot-cell facility and used as a calibration free method, is fully compatible for this type of application.

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363 Table

Parameters	Value	Uncertainty (k=1)
[U] _T	9.5044	0.0039
m_T	0.246555	0.000059
m_S	0.218465	0.000059
M_S	237.992	-
M_T	235.201	-
(235) _T	93.1760	0.0034
$(238)_{S}$	98.053	0.010
R_S	0.019658	0.000015
R_T	0.058065	0.000017
Parameters	Value	Relative Uncertainty (k=1)
R _{Mix}	Between 0.0586 and 50.87	0.07 %

364Table 1: Parameters used for the theoretical study of our studied case in order to find the365optimum sample – spike mixture isotope ratio in the ID-TIMS

2	6	6
2	U	υ

372	Table 2:	Uranium n	nass fraction	detern	nined b	y ID	-TIMS	and	KED	ob	taine	ed on	th	e
~ - ~		11									11	•		. .

uranium pellet. $U_{ass val.}$ corresponds to the assigned value uncertainty (k = 2) and U 373 corresponds to the total uncertainty (k = 2). NB corresponds to the normalized bias 374

	Uranium mass fraction		
Method	ID-TIMS KED		
Assigned value (g g ⁻¹)	0.88110	0.88110	
Results (g g^{-1})	0.8835	0.8807	
<i>RSD</i> (%)	0.12	0.33	
NB	1.8	0.09	
Bias (%)	0.28	-0.04	
$U_{ass_val.}$ (%, k = 2)	0.05	0.05	
U(%, k = 2)	0.32	0.76	
ITV(%, k = 2)	0.36	0.56	

375

376

 Table 3: Experimental duration and effluents

Technique	ID-TIMS	KED			
	Duration				
Dilution step	1 h	/			
Sample preparation	12 h	5 min			
Analysis	9 h	1 h 15 (5 × 15 min)			
Density	/	4h			
Isotope determination	19 h	/			
Total	41 h	5h20			
	Waste n	nanagement			
Sample solution consumption	600 µL	 2 mL for the density determination 0 mL for the KED analysis 			
Generated effluent	180 mL for the dilution step	10 mL for the density determination			

378 Figure





380	Fig. 1: Schematic of the uranium concentration relative uncertainty $(u([U])_S/[U]_S, \%,$
381	k = 1) as a function of the mixture isotope ratio in order to find the optimum mixture
382	isotope ratio in our studied case. The $[U]_s$ and $u([U]_s)$ terms were calculated using Eq.
383	(1) and Eq. (4), respectively, considering all the isotope dilution terms constants (Table 1)
384	except the mixture isotope ratio value

385



386

Fig. 2: Relative contribution (%) of the main uncertainty sources for the uranium mass
 fraction determined by ID-TIMS

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