



HAL
open science

Interaction of the chaperones alpha B-crystallin and CHIP with fibrillar alpha-synuclein: Effects on internalization by cells and identification of interacting interfaces

Maya Bendifallah, Virginie Redeker, Elodie Monsellier, Luc Bousset, Tracy Bellande, Ronald Melki

► To cite this version:

Maya Bendifallah, Virginie Redeker, Elodie Monsellier, Luc Bousset, Tracy Bellande, et al.. Interaction of the chaperones alpha B-crystallin and CHIP with fibrillar alpha-synuclein: Effects on internalization by cells and identification of interacting interfaces. *Biochemical and Biophysical Research Communications*, 2020, 10.1016/j.bbrc.2020.04.091 . cea-02614589

HAL Id: cea-02614589

<https://cea.hal.science/cea-02614589>

Submitted on 21 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Contents lists available at ScienceDirect

Biochemical and Biophysical Research Communications

journal homepage: www.elsevier.com/locate/ybbrc

Interaction of the chaperones alpha B-crystallin and CHIP with fibrillar alpha-synuclein: Effects on internalization by cells and identification of interacting interfaces

Maya Bendifallah, Virginie Redeker, Elodie Monsellier, Luc Bousset, Tracy Bellande, Ronald Melki*

CEA, Institut François Jacob (MIRcen) and CNRS, Laboratory of Neurodegenerative Diseases (U9199), 18 Route du Panorama, 92265, Fontenay-aux-Roses, France

ARTICLE INFO

Article history:

Received 29 March 2020

Accepted 17 April 2020

Available online xxx

Keywords:

Molecular chaperones

Protein-protein interactions

Protein-protein interfaces

Cross linking, mass spectrometry

ABSTRACT

The spread of fibrillar alpha-synuclein from affected to naïve neuronal cells is thought to contribute to the progression of synucleinopathies. The binding of fibrillar alpha-synuclein to the plasma membrane is key in this process. We and others previously showed that coating fibrillar alpha-synuclein by the molecular chaperone Hsc70 affects fibrils properties. Here we assessed the effect of the two molecular chaperones alpha B-crystallin and CHIP on alpha-synuclein fibrils uptake by Neuro-2a cells. We demonstrate that both chaperones diminish fibrils take up by cells. We identify through a cross-linking and mass spectrometry strategy the interaction interfaces between alpha-synuclein fibrils and alpha B-crystallin or CHIP. Our results open the way for designing chaperone-derived polypeptide binders that interfere with the propagation of pathogenic alpha-synuclein assemblies.

© 2020 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

Parkinson's disease and other synucleinopathies are associated with the aggregation of the protein alpha-synuclein (α Syn) into high molecular weight fibrillar aggregates [1,2]. Aggregated α Syn traffic between neuronal cells, seed the aggregation of endogenous monomeric α Syn and amplify [3–5]. This prion-like phenomenon is presumed to contribute to the stereotypic progression of synucleinopathies [5,6]. α Syn aggregates propagate through exosomes [7,8], tunneling nanotubes [9,10] and following their release to the extracellular space through export by living cells or liberation by dying cells [11–13]. The latter aggregates bind to the cell membrane and are internalized, primarily through endocytosis [14,15]. They eventually compromise the endo-lysosomal integrity [16] and reach the cytosol where they amplify by recruiting endogenous monomeric α Syn [17,18]. The resulting aggregates find next their way into naive, unaffected cells, perpetuating the cycle [17,19]. Fibrillar α Syn assemblies' formation and trafficking represent targets for therapeutic interventions [20]. Those assemblies bind to

the cell membranes through numerous protein [21–25], and lipid [26,27] membrane partners, many of which are essential to cell physiology.

α Syn aggregates in the crowded cellular environment in the presence of a vast number of proteins, including molecular chaperones. Molecular chaperones, the first line of defense in protein homeostasis, interact with aggregating proteins [28,29]. Extensive research has reported that various chaperones, such as Hsp90, Hsp70, Hsp27 and alpha B-crystallin (α Bc), interfere with the assembly of monomeric α Syn into fibrils [30,31,32,33,34,35]. Those molecular chaperones may bind aggregated α Syn *in vivo* as they have been shown to be present within α Syn-rich deposits within cells. This justifies characterizing thoroughly the interactions between molecular chaperones and α Syn [36,37,38,39,40,41], demonstrating chaperone binding to α Syn fibrils *in vitro*.

We previously demonstrated that Hsc70 binds fibrillar α Syn with high affinity [33,40]. Hsc70-coated α Syn fibrils are less toxic to cells compared to naked fibrils [33]. We further identified intermolecular surfaces of the interaction between Hsc70 and α Syn [39,40]. Here, we focused on human sHSP chaperone α Bc and human C-terminus Hsc70-interacting protein (CHIP). α Bc is one of the 2 subunits of the chaperone α -crystallin, a prominent component of the human eye lens that prevents aggregation of lens proteins,

* Corresponding author.

E-mail address: ronald.melki@cnrs.fr (R. Melki).

including itself. It consists of a mixture of oligomers ranging in size from 10 to 40 monomers [42]. α Bc colocalizes with Lewy bodies in Parkinson's disease patients' neurons [43]. *In vitro*, α Bc slows down the aggregation of many different proteins [30,44,34] and binds to preformed α Syn fibrils [37,41]. CHIP is a human dimeric chaperone of 34.8 kDa per monomer with E3 ubiquitin ligase activity [45]. It colocalizes with α Syn in Lewy bodies in Dementia with Lewy Body patient brains [46]. CHIP also targets α Syn oligomers for degradation and has a rescue effect in α Syn-induced-toxicity in different cellular models [46,47].

Here we assessed the uptake of α Syn fibrils assembled in the presence of α Bc and CHIP by Neuro-2a cells. We demonstrate that both chaperones diminish α Syn fibrils take up by cells. Using an optimized cross-linking and mass spectrometry strategy, we identified the interaction interfaces between α Syn fibrils and α Bc or CHIP. The two chaperones bind different α Syn polypeptides exposed at the surface of fibrils: a flexible segment located just before the fibrillar core for α Bc and at a solvent accessible surface of the fibrillar core itself for CHIP. Our results may allow for the development of peptide ligands that can interfere with α Syn fibril prion-like propagation and subsequent disease progression.

2. Materials and methods

2.1. Cloning

A synthetic gene encoding α Bc was purchased from GeneArt (Thermo Fisher Scientific). The vector allowing the bacterial expression of α Bc was obtained by subcloning the corresponding synthetic gene into a pET14b vector (Novagen) at the NcoI-XhoI restriction sites. The pET151/D-TOPO vector (Invitrogen) allowing the bacterial expression of His₆, N-terminal-tagged CHIP with a V5 epitope was a gift from Jeffrey L. Brodsky. The His₆ tag is followed by a tobacco etch virus (TEV) protease cleavage site. Constructs were verified by DNA sequencing (Eurofins Genomics).

2.2. Expression & purification of recombinant proteins

Recombinant human wild-type α -Synuclein (α Syn) was expressed and purified as described [48].

Recombinant human wild-type α Bc was expressed at 37 °C in *E.coli* strain BL21(DE3) (Stratagene) upon induction with IPTG (0.5 mM) for 3 h. Cells were harvested and resuspended in buffer A (50 mM Tris-HCl pH 8.5, 1 mM EDTA). After sonication and centrifugation, lysate supernatants were subjected to PEI precipitation (0.12% v/v). The supernatants were filtered and loaded onto a Q-sepharose FF column (GE Healthcare Lifesciences) equilibrated with buffer A, and eluted with buffer B (50 mM Tris-HCl pH 8.5, 1 mM EDTA, 100 mM NaCl). Fractions containing α Bc were pooled and concentrated with a 3 kDa cutoff (Millipore), then loaded onto a size exclusion chromatography column (Superose 6 10/300 GL, GE Healthcare Lifesciences). α Bc eluted with an apparent molecular weight corresponding to a multimer of 20–35 monomers. Gel filtration standards from Sigma-Aldrich (MWGF1000) were used for apparent molecular weight determination.

Recombinant His₆, N-terminal-tagged CHIP was expressed at 30 °C in *E.coli* strain BL21(DE3). Protein expression was induced by IPTG (0.5 mM) for 2 h. The cells were harvested and resuspended in buffer C (50 mM NaPO₄ pH 8.0, 300 mM NaCl, 3 mM β -mercaptoethanol [β ME]). After sonication and centrifugation, lysate supernatants were filtered and loaded onto a 5 mL Talon metal affinity resin column (Clontech), equilibrated in buffer C. The protein was eluted in buffer C supplemented with 200 mM imidazole, then dialyzed in PBS. The His₆-tag was cleaved by addition of His₆-tagged tobacco etch virus (His-TEV) protease, produced using the

plasmid pRK793 (Addgene), at a 1:20 His-TEV:chaperone molar ratio. 100% cleavage, as assessed by SDS-PAGE, was achieved upon incubating the mixtures for 1 h at 37 °C. The untagged proteins were purified by collecting the flow through of a 5 mL Talon metal affinity resin column.

After purification, all proteins were immediately filtered through sterile 0.22 μ m filters, aliquoted and stored at –80 °C. The purity of all proteins was confirmed by SDS-PAGE and MALDI mass spectrometry (not shown). Protein concentrations were determined spectrophotometrically using the following extinction coefficients at 280 nm ($M^{-1} cm^{-1}$): 5960 for α Syn; 13,980 for α Bc and 29,380 for CHIP.

2.3. Assembly of α Syn into fibrils and labeling

For kinetics and internalization experiments, monomeric α Syn was labeled by addition the aminoreactive fluorescent dye Alexa Fluor 488 (Alexa488, Invitrogen) using a protein:dye molar ratio of 10:1 based on initial monomer concentration. Labelling was performed following the manufacturer's recommendation. The reaction was quenched with Tris-HCl pH 7.5 (40 mM final concentration).

α Syn (100 μ M monomer concentration) was incubated with increasing concentrations of α Bc or CHIP (0–100 μ M) in PBS at 37 °C under continuous shaking in an Eppendorf Thermomixer set at 600 rpm for 2 weeks. Aliquots were withdrawn at different time intervals from the assembly reactions and mixed with a primuline solution (10 μ M). Primuline fluorescence was recorded with a Cary Eclipse spectrofluorimeter (Varian Medical Systems Inc) using excitation and emission wavelengths set at 400 and 480 nm, respectively. The nature of the fibrils obtained at the end of the aggregation reaction was assessed by transmission electron microscopy (TEM) as described below. The proportion of α Syn assembled into fibrils was assessed through a partition assay by centrifugation. To this aim, aliquots were withdrawn, subjected to centrifugation in an eppendorf 5415R centrifuge at 20,000 g and 20 °C, and the amount of protein in the supernatant and pellet fraction determined by SDS-PAGE analysis.

2.4. Transmission electron microscopy (TEM)

Fibrils were imaged by TEM in a Jeol 1400 transmission electron microscope (Jeol SAS) after adsorption onto carbon-coated 200 mesh grids and negative staining with 1% uranyl acetate. The images were recorded with a Gatan Orius CCD camera (Gatan Inc.).

2.5. Cell culture and internalization

Murine neuroblastoma Neuro-2a cells (ATCC) were cultured at 37 °C in humidified air with 5% CO₂ in Dulbecco's modified Eagle's medium (DMEM, Sigma-Aldrich) containing 10% fetal bovine serum (Gibco, Thermo Fisher Scientific), 2 mM glutamine, 100 units.ml⁻¹ penicillin (PAA Laboratories) and 100 μ g ml⁻¹ streptomycin (PAA Laboratories). All materials used for cell culture were from PAA Laboratories.

The internalization of Alexa488-labeled α Syn (1 μ M equivalent monomer concentration) assembled in the presence or absence of α Bc or CHIP was assessed. All fibrils were sonicated for 20 min in a VialTweeter powered by an ultrasonic processor UIS250v (250 W, 2.4 kHz; Hielscher Ultrasonic) set at 75% amplitude, 0.5 s pulses before dilution in DMEM. Neuro-2a cells cultured on 96-well plates were then incubated with these mixtures distributed in 5 independent wells. After 2 h, the medium was removed and Hoescht (Sigma-Aldrich), diluted at 0.2 μ g/mL in serum-free, phenol red-free DMEM, was added for 30 min. The cells were washed twice

and 0.1% Trypan Blue (Sigma-Aldrich) was added to quench the fluorescence of Alexa488-labeled α Syn fibrils that were plasma membrane-bound. For each well, Alexa488 and Hoechst fluorescence values were recorded on a Clariostar microplate reader (BMG LABTECH). For each condition, the Alexa488 fluorescence value was considered and averaged over the 5 wells only if the Hoescht value was not significantly different from the one of untreated cells. Three independent experiments were conducted.

2.6. Chemical cross-linking and western blotting

Cross-linking experiments of chaperones with fibrillar α Syn were performed in PBS using α Syn alone (100 μ M), chaperones alone (either 50 μ M for α Bc, or 33 μ M for CHIP), or α Syn (100 μ M) assembled into fibrils in the presence of chaperones (either 50 μ M of α Bc, corresponding to an α Syn: α Bc molar ratio of 2:1; or 33 μ M of CHIP, corresponding to an α Syn:CHIP molar ratio of 3:1) as described above. After two weeks of assembly, fibrillar α Syn samples were centrifuged at 100,000 g in a TL100 tabletop ultracentrifuge (Hitachi) for 30 min at 20 °C. The pellets were allowed to swell in PBS overnight before gentle resuspension. The final concentrations of α Syn fibrillar material and chaperones in the pellets were assessed by a partition assay by ultracentrifugation as described above.

The samples were exposed to various crosslinkers: glutaraldehyde (GA, ~5 Å spacer arm for monomeric glutaraldehyde, Sigma-Aldrich); bis(sulfosuccinimidyl) glutarate (BS2G, 7.7 Å spacer arm, Pierce); bis(sulfosuccinimidyl) suberate (BS³, 11.4 Å spacer arm, Pierce); 3,3'-dithiobis(sulfosuccinimidyl)propionate (DTSSP, 12 Å spacer arm, Pierce); or N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDC) with N-hydroxysulfosuccinimide (Sulfo-NHS) (0 Å spacer arm, Thermo Fisher Scientific) at a 1:1.25 EDC:Sulfo-NHS molar ratio. For GA, BS2G, BS³ and DTSSP, reactions were quenched with the addition of Tris-HCl pH 7.5 (50 mM final concentration). For EDC/Sulfo-NHS, reactions were quenched with the addition of β ME (20 mM final concentration) and Tris-HCl pH 8.2 (50 mM final concentration).

A specific α Syn- α Bc cross-linked complex was obtained with GA exposition for 30 min at RT using a total protein:GA molar ratio of 1:1, corresponding to the following GA concentrations: 32 μ M for α Bc alone; 88 μ M for α Syn fibrils alone; and 100 μ M for α Syn fibrils formed in the presence of α Bc. Total protein:GA molar ratios of 1:2 and 1:5, corresponding to 77/82/100 μ M and 192/408/500 μ M GA, led to the formation of higher order intermolecular cross-links among chaperones and fibrils alone.

A specific α Syn-CHIP cross-linked complex was obtained with EDC/Sulfo-NHS exposition for 30 min at RT using a total protein:EDC:Sulfo-NHS molar ratio of 1:10:12.5, corresponding to the following EDC/Sulfo-NHS concentrations: 320/400 μ M for CHIP alone; 823/1029 μ M for α Syn fibrils alone; 1178/1473 μ M for α Syn fibrils formed in the presence of CHIP. Total protein:EDC:Sulfo-NHS molar ratio of 1:20:25 or at fixed concentrations of 2945/3683 μ M corresponding to total protein:EDC:Sulfo-NHS molar ratios of 1:91:115, 1:36:45 and 1:25:31, respectively, led to the formation of higher order intermolecular cross-links among chaperones and fibrils alone.

In order to solubilize protein samples before SDS-PAGE analysis, all samples were first vacuum-dried then dissociated by addition of 10–20 μ l of pure hexafluoroisopropanol (HFIP, Sigma-Aldrich). After incubation for 1 h in HFIP at RT, samples were evaporated before resuspension in denaturing Laemmli buffer, with the exception of DTSSP cross-linked samples which were resuspended in the same buffer without β ME. Samples were heated for 5 min at 80 °C and loaded onto 7.5% tris-tricine gels.

For Western blot analysis, proteins separated by SDS-PAGE were transferred to nitrocellulose membranes (GE Healthcare). The membranes were first analyzed using an anti- α Syn antibody, stripped, and then revealed with an anti- α Bc or anti-CHIP antibody. Briefly, the membranes were blocked in 3% skim milk in TBS-Tween 0.5%, probed with a primary then secondary antibody and developed with the enzyme-coupled luminescence technique (ECL, Thermo Fisher Scientific) using a ChemiDoc™ MP Imaging System (BioRad). The primary and secondary antibodies combinations used were the following: SYN1 antibody (1/5000, 610,787 BD Biosciences) followed by anti-mouse-HRP (1/5000, GTX 213111-01 Genetex) for α Syn detection, and rabbit polyclonal antibodies (1/5000) we raised against α Bc or CHIP followed by anti-rabbit-HRP (1/5000, A120–101P Bethyl). For polyclonal antibody immunopurification, 1–10 mg of purified antigen protein was separated into a 15% SDS-PAGE gel and further transferred to a nitrocellulose membrane (GE Healthcare). The antigen-containing membrane was pre-incubated with 100 mM glycine pH 2.5 for 10 min before being blocked with 5% skim milk in TBS-Tween 0.05% and further incubated overnight in the anti- α Bc or anti-CHIP antibody-containing serum. Polyclonal antibodies against α Bc or CHIP were eluted with 2 mL 100 mM glycine pH 2.5 for 2 min. After immediate neutralization with addition of Tris-HCl pH 8.5, the immunopurified antibodies were dialyzed into PBS pH 7.4, 30% glycerol. Stripping of the membrane between two immunodetections was achieved with 100 mM β ME, 62.5 mM Tris-HCl pH 6.8, 2% SDS for 1 h at 50 °C.

2.7. Peptide preparation and NanoLC-MS/MS identification of cross-linked peptides

Bands of interest were excised and cut in pieces of about 2 mm². Each band was subjected to washing and in-gel tryptic digestion as described [49], without reduction and alkylation. Tryptic digestion was performed overnight at 37 °C using Trypsin Gold (Promega) at a concentration of 10 ng/ μ l in 25 mM ammonium bicarbonate. Tryptic peptides were extracted using two volumes of 60% acetonitrile (ACN), 0.1% formic acid for 1 h. The supernatant was recovered and a second extraction was performed by addition of one volume of 100% ACN for 30 min. The two supernatants of extracted peptides were pooled and vacuum dried. Tryptic peptides were resuspended in 0.1% formic acid (v/v) prior to nanoLC-MS/MS analysis.

Tryptic peptides were analyzed with a Data Dependent acquisition method by selecting the 20 most intense precursors for CID fragmentation using a Triple-TOF 4600 mass spectrometer (ABSciex) coupled to a nanoRSLC ultra performance liquid chromatography (UPLC) system (Dionex, Thermo Fisher Scientific) as previously described [50] with the following exceptions. The analytical column was an Acclaim PepMap RSLC C18 (75 μ m i.d. \times 50 cm, 2 μ m, 100 Å, Thermo Fisher Scientific). Peptides were loaded at 6 μ l/min and separated with a gradient of 5–35% 0.1% formic acid in 100% ACN.

2.8. NanoLC-MS/MS data analysis

NanoLC-MS/MS raw data were converted into mgf data files using the MS Data Converter software (Peakview, ABSciex). Sequence coverage of the proteins was determined through database searching using the Mascot search engine (Matrix Science; version 2.6) with the SwissProt mammalian database (SwissProt_2018_10 database release) together with a decoy database search. Only peptides with a mascot score above 20 were considered. Inter-molecular cross-linked peptides between α Syn and

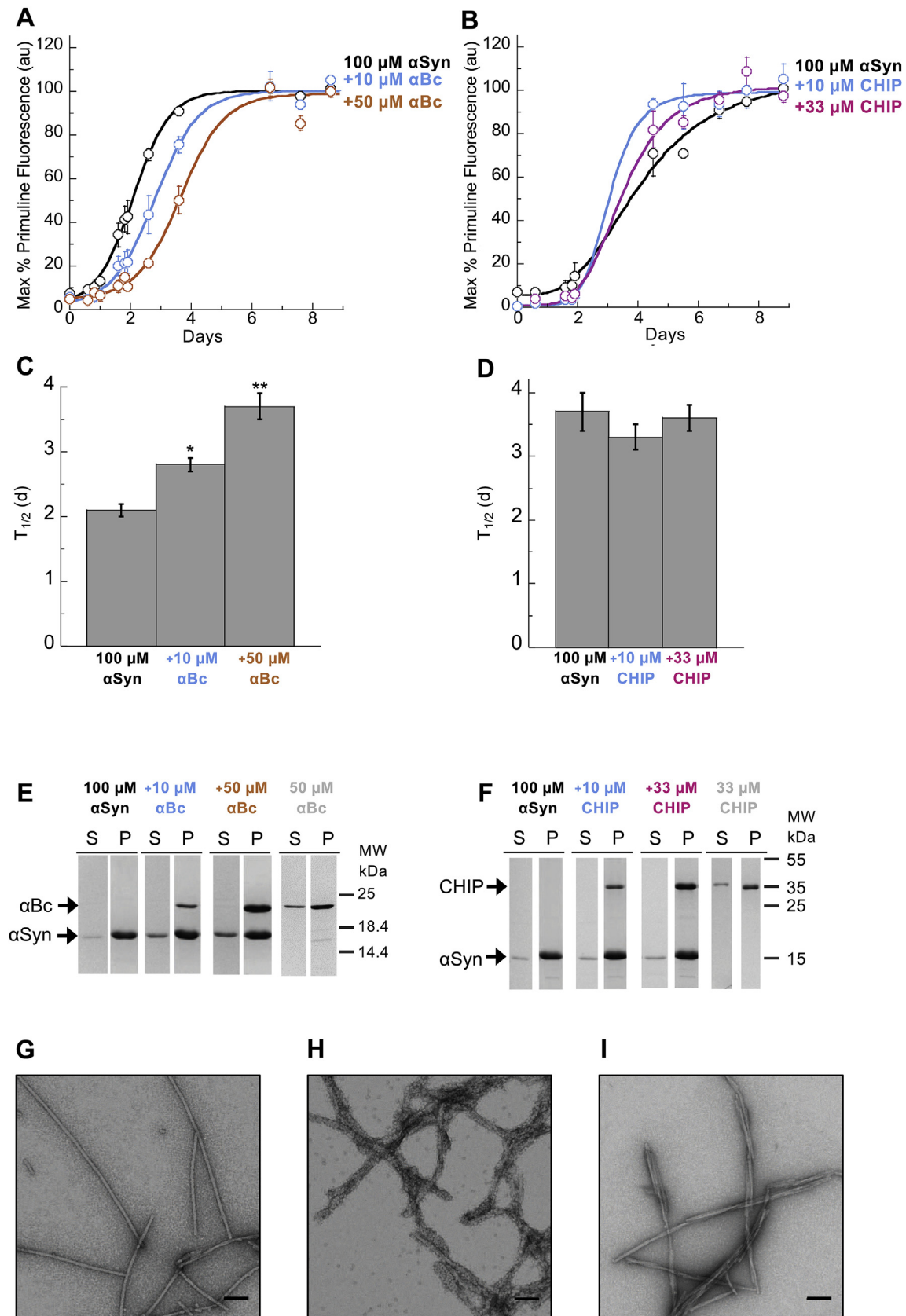


Fig. 1. α Syn fibrillation in presence of chaperones. (A–B) Time-course fibrillation of α Syn-Alexa488 (100 μ M) in the presence of increasing concentrations of α Bc (A) or CHIP (B) was followed by primuline fluorescence. Chaperone concentrations of 0 μ M, 10 μ M, 20 μ M, 33 μ M, 50 μ M are represented as α Syn:chaperone molar ratios of 1:0 (black), 10:1 (blue), 3:1 (pink) or 2:1 (orange) respectively. Means and their associated standard error values were calculated from three independent experiments. Lines through the data points are the best sigmoidal fit. (C–D) Half-time ($t_{1/2}$) of α Syn-Alexa488 aggregation in the presence of increasing concentrations of α Bc (C) or CHIP (D). For each independent experiment, the $t_{1/2}$

chaperones were searched using the STAVROX software (version 3.6.6.6), [51]. For GA, which exists as a mixture of monomers (free aldehyde or cyclic hemiacetal forms) or different polymers [52], we set the mass modification to 64.031 or 82.042 to search for inter-protein peptides cross-linked by a free aldehyde or a cyclic hemiacetal GA, respectively, on a list of potentially reactive amino acids for sites 1 and 2 as reported in the literature, or as indicated by their physico-chemical properties (Lysine, Serine, Threonine, Tyrosine, Arginine, Tryptophan, Phenylalanine, Proline, and N-terminus). One interprotein cross-link by a cyclic hemiacetal monomer of GA was identified and confirmed. For EDC, mass modification was set to 18.01 Da for interprotein cross-linked peptides modified on Lysine, Serine, Threonine, Tyrosine, and N-terminus for site 1 and Aspartate, Glutamate and C-terminus for site 2. For both Mascot and Stavrox searches, the parent and fragment ion mass tolerances were 25 ppm and 0.05 Da, respectively, oxidation of methionine was set as a variable modification, and trypsin cleavage was semi-specific. Candidate cross-linked peptides were validated after confirmation of their absence in the control samples and manual analysis of their MS/MS spectra.

3. Results

Extensive studies have reported that chaperones interact with amyloid-forming proteins and are thus good candidates to prevent either the aggregation reaction or the interaction of the resulting amyloids with binding partners within the cytosol or at the plasma membrane. Previous work in our laboratory showed that the molecular chaperone Hsc70 reduces α Syn assembly and affects pre-formed fibrils binding and subsequent toxicity [33].

We wanted to assess if the ability to bind preformed α Syn fibrils is a generic property of molecular chaperones, in the perspective of expanding our arsenal of chaperones able to modify α Syn fibril surfaces and interfere with their cell-to-cell propagation. We selected two different molecular chaperones that we considered promising based on previous data demonstrating their association or colocalization with aggregated α Syn, either *in vitro* or in pathological lesions or inclusions observed in the brains of patients with synucleinopathies [43,37,34,46,47,41]: the human sHSP chaperone α B-crystallin (α Bc) and human C-terminus Hsc70-interacting protein (CHIP).

Effect of α Bc and CHIP on α Syn fibrils take-up by Neuro-2a cells.

We first assessed the effect of α Bc and CHIP on the assembly of α Syn into fibrils at different α Syn:molecular chaperone molar ratios (Fig. 1). We used Alexa488-labeled monomeric α Syn for these assembly reactions, in order to obtain labeled α Syn fibrils in complex with unlabeled chaperones and assess the uptake of the resulting fibrils by Neuro-2a cells. The kinetics of aggregation were followed by primuline fluorescence. Notably, the assembly kinetics of unlabeled and Alexa488-labeled α Syn into fibrils, as well as the fibrils abilities to seed, are comparable [53].

Fibrillation kinetics of α Syn in presence of increasing concentrations of α Bc show that α Bc significantly slows down α Syn aggregation, starting from an α Syn: α Bc molar ratio of 10:1 (Fig. 1A, C). At a 2:1 α Syn: α Bc molar ratio, α Bc did not significantly modify neither the amount (Fig. 1E) nor the morphology (Fig. 1H) of fibrillar α Syn. While CHIP had no significant effect neither on α Syn

kinetics of aggregation (Fig. 1B,D) nor on the amount (Fig. 1F) of aggregated α Syn, it had an effect on the morphology of fibrils. They appeared more twisted than those formed without molecular chaperones (Fig. 1I). Increases of α Bc or CHIP concentrations induced an increase of the amount of chaperones bound to α Syn fibrils, as identified through a sedimentation assay (Fig. 1E and F).

We next assessed the internalization of α Syn fibrils formed in the presence of α Bc or CHIP by Neuro-2a cells (Fig. 2A and B, respectively). Neuro-2a cells, in 96-well plates, were exposed for 2 h to Alexa488-labeled α Syn fibrils (1 μ M) pre-incubated or not with α Bc or CHIP (0, 2 and 10 μ M), prior to Trypan blue addition and quantification of Alexa488 fluorescence in a plate-reader. α Syn fibril fluorescence quenching by Trypan blue allowed us to determine accurately the amount of internalized fibrils in a dose-dependent manner. We used Hoechst staining to ascertain that the number of cells remained constant (see Material & Methods). As a positive control for this experiment, we examined the effect of heparin, a ligand of cell membrane heparan sulfate proteoglycans that have been implicated in α Syn internalization [24]. When preformed α Syn-Alexa488 fibrils were pre-incubated with 0.2 μ g/mL heparin, we observed an 83% reduction of their internalization (Fig. S1), in agreement with previous reports [24]. α Syn-Alexa488 fibrils assembled in presence of increasing amounts of α Bc or CHIP take-up by Neuro-2a cells decreased in a chaperone concentration-dependent manner (Fig. 2). We conclude from these experiments that α Bc or CHIP affect fibrillar α Syn uptake by Neuro-2a cells, most probably by binding the fibrils and changing their surface properties.

Identification of interaction surfaces between fibrillar α Syn, α Bc and CHIP through chemical cross-linking.

We next mapped the surface interfaces between α Bc or CHIP and fibrillar α Syn to further characterize the interactions. We used chemical cross-linking and mass spectrometric identification of cross-linked peptides to this aim [54,39,55,40].

α Syn fibrils (100 μ M) formed in presence or absence of α Bc (50 μ M) or CHIP (33 μ M) were exposed to different cross-linkers at different total protein to crosslinker molar ratios. α Syn fibrils, α Bc and CHIP were exposed individually to the cross-linkers in control reactions. The cross-linkers we used had distinct structural-chemical characteristics. Three homobifunctional water soluble NHS-ester cross-linkers specific to nucleophilic groups (predominantly primary amines e.g. proteins N-terminus and Lysine residues, but also Tyrosine, Serine and Threonine) with varying spacer arm lengths: 7.7, 11.4 and 12 Å for BS2G, BS³ and DTSSP, respectively, were used. We further used the heterobifunctional zero length carbodiimide crosslinker EDC that crosslinks primary amines to carboxyl groups (e.g. Glu, Asp and proteins C-terminus) in the presence of Sulfo-NHS. Finally, we also used GA, known for its efficiency, but also its limited specificity and molecular heterogeneity with a ~5 Å spacer arm for the monomer [52]. The cross-linking reactions products were analyzed by SDS-PAGE to identify fibrillar α Syn- α Bc or fibrillar α Syn-CHIP complexes, observed neither in the fibrillar α Syn nor in the chaperone control samples subjected to identical cross-linking conditions. The total protein to cross-linker ratio was optimized. A 1:1 and 1:10:12.5 total protein to cross-linker ratios were selected for fibrillar α Syn- α Bc cross-linking with GA and for fibrillar α Syn-CHIP cross-linking with EDC/Sulfo-NHS, respectively. Finally, as the reaction involved cross-

parameter was extracted from the best fit to a sigmoid function. The means are their associated standard error values were calculated from three independent experiments. Statistical significance was determined through an unpaired student's t-test with equal variance. Annotations indicating level of significance are as follows: *p < 0.05; **p < 0.01. Legends state α Syn:chaperone molar ratios. (E–F) Supernatant and pellet samples at the end of the aggregation reactions in absence or presence of α Bc (E) or CHIP (F) resolved by SDS-PAGE and coomassie blue staining. Legends state α Syn and chaperone concentrations. (G–I) TEM micrographs of α Syn-Alexa488 fibrils formed alone (G), fibrils formed in presence of α Bc (2:1 α Syn: α Bc molar ratio, H), and fibrils formed in presence of CHIP (3:1 α Syn:CHIP molar ratio, I). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

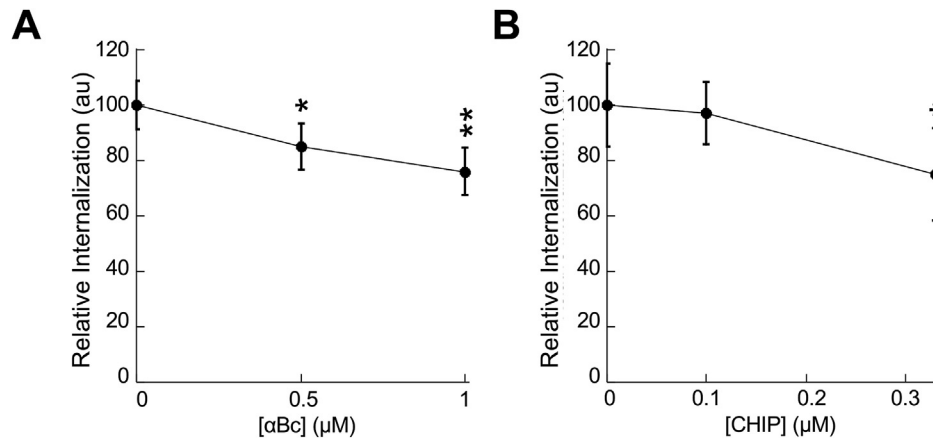


Fig. 2. Effect of α Bc and CHIP on α Syn fibril uptake by Neuro-2a cells. α Syn-Alexa488 fibrils were assembled at 100 μ M in the absence or in the presence of 50 or 100 μ M α Bc or 10 or 33 μ M CHIP. After two weeks of fibril formation, the mixture was diluted 100x in DMEM and incubated for 30 min at 37 °C. Neuro-2a cells in 96-well plates were exposed for 2 h to α Syn-Alexa488 fibrils assembled in the presence of α Bc (A) or CHIP (B). Trypan blue was added after extensive washing to quench the fluorescence of plasma membrane-bound α Syn-Alexa488 fibrils. The amount of internalized α Syn-Alexa488 was measured on a fluorescence plate reader. Means and their associated standard deviation values are calculated over 5 wells. In each case we performed three independent experiments; one representative experiment is shown. The results and the associated significances are reported to the internalization in the absence of chaperone. Statistical significance was determined through an unpaired student's t-test with equal variance. Annotations indicating level of significance are as follows: *p < 0.05; **p < 0.01; ***p < 0.001.

linking fibrillar α Syn to the molecular chaperones, the cross-linked high molecular weight complexes were dissociated to allow their analysis by SDS-PAGE.

Specific cross-linked complexes were observed with GA and fibrillar α Syn with α Bc (Fig. 3A) upon staining the polyacrylamide gel with Coomassie blue and on western blots using antibodies directed against α Syn and α Bc. The exposure of α Syn fibrils alone to GA yielded protein bands representing monomers, dimers, trimers and tetramers, while that of α Bc alone did not affect the intensity of the dimeric form of the protein. Higher concentrations of GA led to the formation of cross-linked higher molecular weight α Syn and α Bc species (Fig. S2A). When α Syn- α Bc complex was cross-linked with GA, a unique and specific protein band appears at around 40 kDa (arrow). Western blot analyses with antibodies directed against α Syn and α Bc confirms that this specific band contains both proteins and thus corresponds to a covalent α Syn- α Bc complex. Its apparent molecular mass of 40 kDa suggests it is formed of one α Syn and one α Bc monomer. The band of interest corresponding to the α Syn- α Bc complex was thus cut off the gel and subjected to trypsin digestion. The resulting peptides were analyzed by nano-liquid chromatography coupled to tandem mass spectrometry (nanoLC-MS/MS). For the α Syn- α Bc complex, using database searching as described in Materials & Methods, we obtained a sequence coverage of 100% for α Syn and 97% for α Bc. Using the Stavrox software [51], we identified one α Syn- α Bc inter-protein cross-linked peptide that was absent from control bands (monomeric or dimeric α Syn or α Bc with or without GA cross-linking) and was validated manually. This peptide was identified as a quadruple peptide ion with a mass-to-charge of 622.847, consistent with an inter-peptide cross-link that covalently binds two peptide sequences and thus is highly charged (Fig. 3B). Its fragmentation spectrum, presented in Fig. 3B, identified the cross-linked peptides and sites. The y and b fragments ions series reported in Fig. 3C identified the α Syn [33–43] and α Bc [153–163] sequences of the cross-linked peptide with a cross-link between Lys-34 from α Syn and Thr-162 from α Bc. The mass deviation of -3.02 ppm measured between the experimental and the theoretical mass of this cross-linked peptide further validated the identification. Interestingly the α Bc peptide [153–163] in which Thr-162 is cross-linked is located in the flexible, unstructured C-terminus of the chaperone [44], and the α Syn peptide [33–43] in which Lys-34 is cross-linked

to α Bc is exposed to the solvent in the fibrillar structures of α Syn (PBD code 6rt0 and 6rtb) we recently solved (Fig. 3E), [56].

Fibrillar- α Syn-CHIP cross-links obtained using EDC/Sulfo-NHS were resolved by SDS-PAGE and detected both by coomassie blue staining and Western blot using antibodies against α Syn and CHIP (Fig. 4A). A dimeric form of CHIP was observed upon treatment of the protein with EDC/Sulfo-NHS, confirming its dimeric state. Higher concentrations of EDC/Sulfo-NHS led to the formation of cross-linked higher molecular weight α Syn and CHIP (Fig. S2B). When α Syn-CHIP complex was cross-linked with EDC, a unique and specific protein band appears at an apparent molecular mass around 55 kDa (arrow). Western blot analyses with antibodies directed against α Syn and CHIP confirm that this specific additional band contains both proteins and thus corresponds to a covalent α Syn-CHIP complex. Its apparent molecular mass of about 55 kDa suggests it is constituted by one α Syn and one CHIP monomer. The band of interest corresponding to the α Syn-CHIP covalent complex was cut off the gel and in-gel digested using trypsin and the resulting peptides were further analyzed by nanoLC-MS/MS as described above. By database search as described in Material and Methods, we obtained a sequence coverage of 100% for α Syn and 85% for CHIP. Using the Stavrox software [51], we identified one α Syn-CHIP inter-protein cross-linked peptide that was absent from control bands (monomeric or dimeric α Syn or CHIP with or without EDC cross-linking) and was validated manually. This peptide was identified as a quadruple charged ion with a mass-to-charge of 616.136 (Fig. 4B). Its fragmentation spectrum presented in Fig. 4B unambiguously identified the cross-linked peptides and sites. The y and β fragments ions, reported in Fig. 4C, identified the α Syn [61–80] and CHIP [254–262] sequences of the cross-linked peptide with a cross-link between Glu-61 from α Syn and Lys-254 from CHIP. Finally, the mass deviation of 4.17 ppm measured between the experimental and the theoretical mass of this cross-linked peptide further validated the identification. Interestingly, CHIP peptide [254–262], in which Lys-254 (in purple, space fill, Fig. 4D) has been cross-linked to fibrillar α Syn, is located within the CHIP U-box domain [45] while α Syn peptide [61–80], in which Glu61 (red sphere, Fig. 4E) was cross-linked to CHIP, is located in the non-amyloid component (NAC) region and is accessible to the solvent in the fibrillar structures of α Syn (PBD code 6rt0 and 6rtb) we recently solved [56].

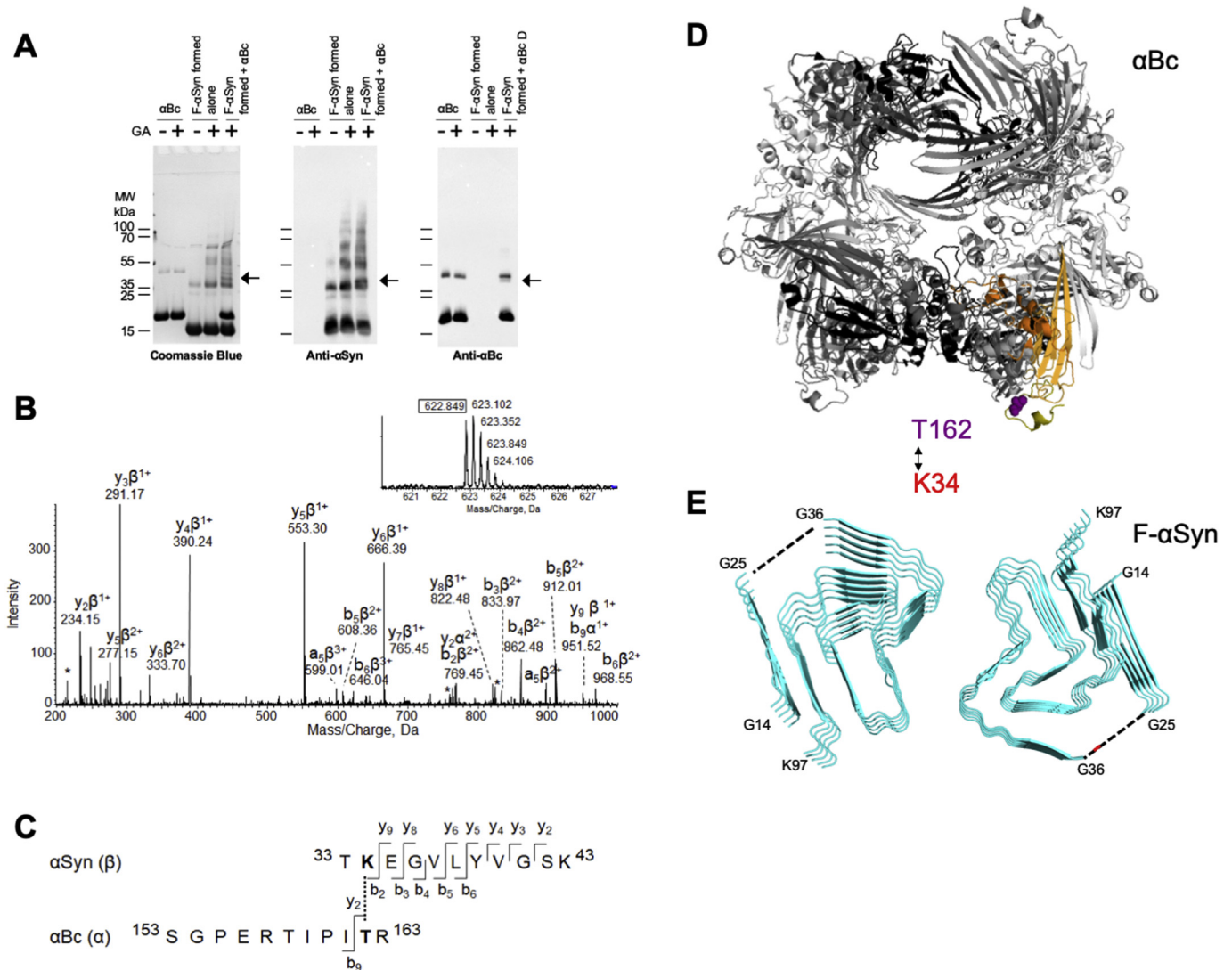


Fig. 3. Cross-linking of α Bc to α Syn fibrils by glutaraldehyde. (A) Cross-linking reaction protein products were separated on a 7.5% tris-tricine SDS-PAGE and detected by coomassie blue staining and immunodetection by Western blot using detections with either SYN1 anti- α Syn or in-house-immunopurified rabbit polyclonal anti- α Bc antibodies. α Bc alone, α Syn fibrils (F- α Syn) alone or α Syn fibrils formed in the presence of α Bc (2:1 α Syn: α Bc molar ratio) were exposed to the glutaraldehyde (GA) cross-linker at the same total protein:GA molar ratio (1:1). The arrows show the band of the α Syn- α Bc complex of interest. (B–C) NanoLC-MS/MS analysis and data analysis using the Stavrox software identified the peptide [153–163] from α Bc cross-linked with peptide [33–43] from α Syn fibrils. In B, the fragmentation spectrum of the quadruple-charged cross-linked precursor ion at the monoisotopic m/z 622.847 (on the top) is annotated with the identified fragments and their charge state. Loss of water (asterisks) and internal fragments (i) are indicated. In C, the identified fragments are indicated on the cross-linked sequences. The α and β sequences correspond to the α Bc [153–163] and α Syn [33–43] peptides, respectively. The identified cross-link involves residues Thr-162 and Lys-34 from α Bc and α Syn, respectively. (D) Location of the cross-linked threonine in a model structure of an α Bc 24-mer (PDB accession number 2ygd, 9.4 Å resolution). Thr-162 (T162) is represented by a purple sphere. (E) Location of the cross-linked lysine in α Syn fibrils (PDB accession number 6rt0). Lys-34 (K34) is in a flexible portion of the sequence, not resolved by cryo-EM, and sketched by a dotted black line; K34 itself is represented by a red line. Figures D–E were generated with Pymol (<http://www.pymol.org>). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

4. Discussion

The aggregation of distinct proteins is hallmark of neurodegenerative disease [29]. Molecular chaperones interfere with this process and/or are a component of the resulting aggregates [29,34,43,44,46,57,58]. α Syn fibrillar aggregates are capable of propagating between neuronal cells and consequently contribute to disease progression in synucleinopathies [3,4]. Therefore, identifying the contribution of molecular chaperones within this process and determining whether they bind α Syn fibrils and affect their propagation is of particular interest. It has been previously shown that Hsc70 not only reduces aggregation of α Syn [33] into fibrils, but also binds the fibrils and affects their toxicity [33]. Furthermore, we identified inter-molecular interaction surfaces

between Hsc70 and α Syn using covalent cross-linking and mass spectrometry [39,40].

The binding of molecular chaperones to preformed α Syn fibrils have been assessed on rare occasions [36,37,38,41]. The molecular chaperone α Bc, we used here, was shown for instance to bind to preformed α Syn fibrils [37,41]. We thus went further and assessed the take-up by cultured cells of fibrillar α Syn assembled in the presence of molecular chaperones. We show that the molecular chaperones α Bc and CHIP bind efficiently to α Syn fibrils generated in their presence and affect their take-up by Neuro-2a cells. We further identified the inter-molecular interaction interfaces between fibrillar α Syn and α Bc or CHIP.

α Bc binds to fibrillar α Syn through its C-terminal unstructured domain. Our cross-linking studies show that α Bc interacts with

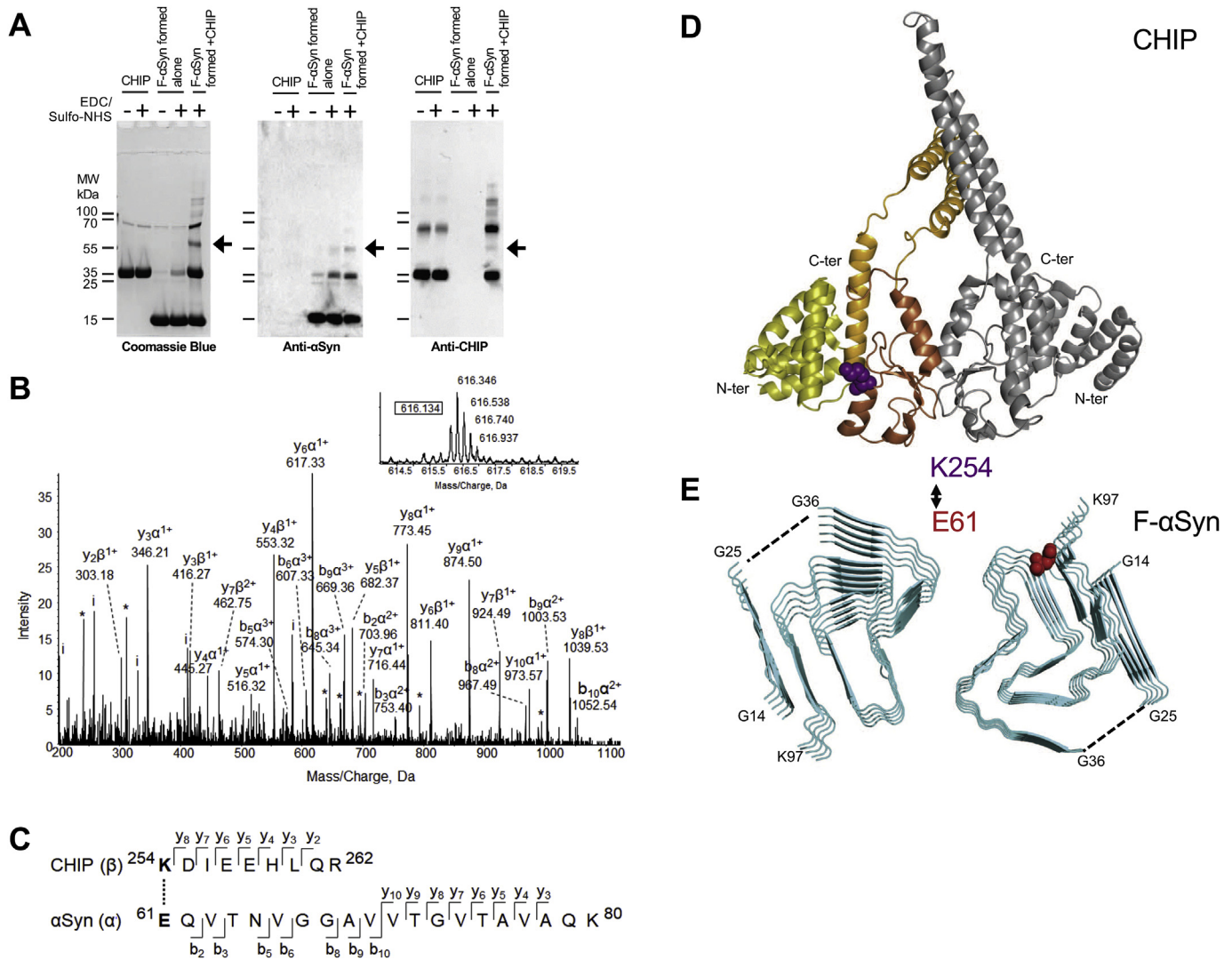


Fig. 4. Cross-linking of CHIP to α Syn fibrils by EDC/SulfoNHS-ester. (A) Cross-linking reaction protein products were separated on a 7.5% tris-tricine SDS-PAGE and detected by coomassie blue staining and immunodetection by Western blot using detections with either SYN1 anti- α Syn or in-house-immunopurified rabbit polyclonal anti-CHIP antibodies. CHIP alone, α Syn fibrils alone or α Syn fibrils formed in the presence of CHIP (3:1 α Syn:CHIP molar ratio) were exposed with EDC/SulfoNHS-ester cross-linker at same total protein:EDC:Sulfo-NHS molar ratio (1:10:12.5). The arrows show the band of the α Syn-CHIP complex of interest. (B–C) NanoLC-MS/MS analysis and data analysis using the Stavrox software identified the peptide [254–262] from CHIP cross-linked with peptide [61–80] from α Syn fibrils. In B, the fragmentation spectrum of the quintuple-charged cross-linked precursor ion at the monoisotopic m/z 616.136 (on the top) is annotated with the identified fragments and their charge state. Loss of water (asterisks) and internal fragments (i) are indicated. In C, the identified fragments are indicated on the cross-linked sequences. The α and β sequences correspond to the α Syn [61–80] and CHIP [254–262] peptides, respectively. The cross-link involves residues Lys-254 from CHIP and fibrillary α Syn, respectively. (D) Location of the cross-linked lysine in a CHIP dimer (Protein Data Bank accession number 2C2L, of murine CHIP, sharing 98% sequence identity with human CHIP). Lys-254 (K254) is represented by a purple sphere. The helical hairpin domains are in light orange, the tetratricopeptide repeat (TPR) domains are in yellow; the U-box domains are in bright orange. (E) Location of the cross-linked glutamate in α Syn fibrils (PDB accession number 6rt0). Glu-61 (E61) is represented by a red sphere. Figures D–E were generated with PYMOL (<http://www.pymol.org>). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

α Syn fibrils. A unique intermolecular peptide was identified, where Thr-162 from α Bc and Lys-34 from α Syn fibrils were cross-linked by GA. Lys-34 from α Syn is located within a flexible loop exposed to the solvent and α Syn fibril binders in the α Syn fibrillar structures (PDB code 6rt0 and 6rtb) we recently solved (Fig. 3E), [56]. Thr-162 from α Bc lies in the unstructured C-terminal domain of α Bc (Fig. 3D). α Bc has been shown to interact with monomeric α Syn through a different region, namely a conserved β 4/ β 8 surface lying within its core [59]. Indeed, α Bc-core prevents the aggregation of α Syn [37,60]. Another heat shock protein, Hsp27, prevents the aggregation of α Syn and binds α Syn fibrils [37,61]. Interestingly, and in contrast with the whole protein, truncated Hsp27 containing only the conserved β 4/ β 8 α -crystallin core domain loses its ability

to bind α Syn fibrils [61]. Furthermore, dimeric, untruncated mutant Hsp27 binds α Syn fibrils, suggesting that residues outside of the core region of the conserved α -crystallin domain play a role in α Syn fibril binding [61].

CHIP binds to fibrillar α Syn through its C-terminal U-box domain. As for α Bc, we mapped the interfaces between α Syn fibrils and the molecular chaperone CHIP. One inter-molecular peptide was identified, where Lys-254 from CHIP and Glu-61 from α Syn fibrils were cross-linked by EDC. Glu-61 from α Syn is located at one tip of the α Syn non-amyloid component (NAC) region and is exposed to the solvent (Fig. 4E), [56]. Lys-254 from CHIP is located in the C-terminal U-box domain of the chaperone (Fig. 4D), responsible for CHIP binding to ubiquitin-conjugating enzymes [45]. This result is

coherent with previously published data. Indeed, a CHIP variant devoid of the Hsp70-binding N-terminal tetratricopeptide repeat domain (TPR, in yellow in Fig. 4D) but in which the U-box domain was still present, was able to immunoprecipitate α Syn [47].

It is worth noting that the interaction areas we identified within the α Bc- α Syn fibril or CHIP- α Syn fibril complexes are not exhaustive as the cross-linking and mass spectrometry strategy we implemented to fibrillar assemblies remains challenging. Nonetheless, identifying polypeptides originating from molecular chaperones come into close proximity with polypeptide stretches exposed at α Syn fibril surfaces may pave the way for the development of finely targeted ligands that can interfere with α Syn fibril prion-like propagation and subsequent disease progression. Notably, peptides are tunable. Their affinity for their protein targets can be optimized and their pharmacokinetics properties can be adjusted [62].

Author contributions

Conceived the project and designed experiments: EM, VR and RM; performed experiments and analyzed data: MB, VR, LB and TB; provided resources, funding and equipment: RM; wrote the manuscript: MB, and RM.

Acknowledgements

We thank David Cornu for expert assistance in MS and Jeffrey L. Brodsky for the CHIP plasmid. This work was supported by the Centre National de la Recherche Scientifique, the Institut National de la Santé et de la Recherche Médicale, the Région Ile de France through DIM Cerveau et Pensée and the EC Joint Programme on Neurodegenerative Diseases and Agence Nationale pour la Recherche (TransPathND, ANR-17-JPCD-0002-02 and Protest-70, ANR-17-JPCD-0005-01). This work also received support from the EU/EFPIA/Innovative Medicines Initiative 2 Joint Undertaking (IMPRiND grant No. 116060 and PD-MitoQUANT grant No. 821522). This work benefited from the electron microscopy facility Imagerie-Gif and the proteomic facility SICaPS.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.bbrc.2020.04.091>.

References

- [1] Spillantini MG, Crowther AR, Jakes R, et al., Filamentous α -synuclein inclusions link multiple system atrophy with Parkinson's disease and dementia with Lewy bodies. *Neurosci. Lett.* 251 (1998a) 205–208.
- [2] Spillantini MG, Crowther AR, Jakes R, et al., Alpha-Synuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with Lewy bodies. *Proc. Natl. Acad. Sci.* 95 (1998b) 6469–6473.
- [3] P. Brundin, R. Melki, R. Kopito, Prion-like transmission of protein aggregates in neurodegenerative diseases, *Nat. Rev. Mol. Cell Biol.* 11 (2010) 301–307.
- [4] A.A. Davis, C.E.G. Leyns, D.M. Holtzman, Intercellular spread of protein aggregates in neurodegenerative disease, *Annu. Rev. Cell Dev. Biol.* 34 (2018) 545–568.
- [5] P. Brundin, R. Melki, Prying into the Prion Hypothesis for Parkinson's Disease, *J. Neurosci.* 37 (2017) 9808–9818.
- [6] H. Braak, K. Del Tredici, U. Rüb, et al., Staging of brain pathology related to sporadic Parkinson's disease, *Neurobiol. Aging* 24 (2003) 197–211.
- [7] E. Emmanouilidou, K. Melachroinou, T. Roumeliotis, et al., Cell-Produced α -Synuclein Is Secreted in a Calcium-Dependent Manner by Exosomes and Impacts Neuronal Survival, *J. Neurosci.* 30 (2010) 6838–6851.
- [8] J. Ngolab, I. Trinh, E. Rockenstein, et al., Brain-derived exosomes from dementia with Lewy bodies propagate α -synuclein pathology, *Acta Neuropathol. Commun.* 5 (2017) 46.
- [9] S. Abouint, L. Bousset, F. Loria, et al., Tunneling nanotubes spread fibrillar α -synuclein by intercellular trafficking of lysosomes, *EMBO J* 35 (2016) 2120–2138.
- [10] J. Rostami, S. Holmqvist, V. Lindstrom, et al., Human Astrocytes Transfer Aggregated Alpha-Synuclein via Tunneling Nanotubes, *J. Neurosci.* 37 (2017) 11835–11853.
- [11] M. Brahic, L. Bousset, G. Bieri, et al., Axonal transport and secretion of fibrillar forms of α -synuclein, A β 42 peptide and HTTExon 1, *Acta Neuropathol* 131 (2016) 539–548.
- [12] P. Desplats, H.-J. Lee, E.-J. Bae, et al., Inclusion formation and neuronal cell death through neuron-to-neuron transmission of α -synuclein, *Proc. Natl. Acad. Sci.* 106 (2009) 13010–13015.
- [13] H.-J. Lee, S. Patel, S.-J. Lee, Intravesicular Localization and Exocytosis of α -Synuclein and its Aggregates, *J. Neurosci.* 25 (2005) 6016–6024.
- [14] V. Grozdanov, K.M. Danzer, Release and uptake of pathologic alpha-synuclein, *Cell Tissue Res.* 373 (2018) 175–182.
- [15] L. Rodriguez, M.M. Marano, A. Tandon, Import and export of misfolded α -synuclein, *Front. Neurosci.* 12 (2018) 1–9.
- [16] W.P. Flavin, L. Bousset, Z.C. Green, et al., Endocytic vesicle rupture is a conserved mechanism of cellular invasion by amyloid proteins, *Acta Neuropathol.* 134 (2017) 629–653.
- [17] M. Jucker, L.C. Walker, Self-propagation of pathogenic protein aggregates in neurodegenerative diseases, *Nature* 501 (2013) 45–51.
- [18] W. Peelaerts, L. Bousset, V. Baekelandt, et al., α -Synuclein strains and seeding in Parkinson's disease, incidental Lewy body disease, dementia with Lewy bodies and multiple system atrophy: similarities and differences, *Cell Tissue Res.* 373 (2018) 195–212.
- [19] T. Tyson, J.A. Steiner, N. Brundin, Sorting out release, uptake and processing of alpha-synuclein during prion-like spread of pathology, *J. Neurochem.* 139 (2016) 275–289.
- [20] S. Pemberton, R. Melki, The interaction of Hsc70 protein with fibrillar alpha-synuclein and its therapeutic potential in Parkinson's disease, *Commun. Integr. Biol.* 5 (2012) 94–95.
- [21] X. Mao, M.T. Ou, S.S. Karuppagounder, et al., Pathological α -synuclein transmission initiated by binding lymphocyte-activation gene 3, *Science* 353 (2016) aah3374–aah3374.
- [22] A.N. Shrivastava, V. Redeker, N. Fritz, et al., α -synuclein assemblies sequester neuronal α 3-Na⁺/K⁺-ATPase and impair Na⁺ gradient, *EMBO J.* 34 (2015) 2408–2423.
- [23] A.N. Shrivastava, A. Triller, R. Melki, Cell biology and dynamics of Neuronal Na⁺/K⁺-ATPase in health and diseases, *Neuropharmacology* 169 (2020) 107461, 0–1.
- [24] B.E. Stopschinski, B.B. Holmes, G.M. Miller, et al., Specific glycosaminoglycan chain length and sulfation patterns are required for cell uptake of tau versus α -synuclein and β -amyloid aggregates, *J. Biol. Chem.* 293 (2018) 10826–10840.
- [25] A.N. Shrivastava, A. Aperia, R. Melki, Triller A Physico-Pathologic Mechanisms Involved in Neurodegeneration: Misfolded Protein-Plasma Membrane Interactions, *Neuron* 95 (2017) 33–50.
- [26] M. Grey, S. Linse, H. Nilsson, et al., Membrane interaction of α -synuclein in different aggregation states, *J. Parkinsons Dis.* 1 (2011) 359–371.
- [27] R.S. Trevino, J.E. Lauckner, Y. Sourigues, et al., Fibrillar structure and charge determine the interaction of polyglutamine protein aggregates with the cell surface, *J. Biol. Chem.* 287 (2012) 29722–29728.
- [28] E.P. De Mattos, A. Wentink, C. Nussbaum-Krammer, C. Hansen, S. Bergink, R. Melki, H.H. Kampinga, Protein quality control pathways at the crossroad of synucleinopathies, *J. Parkinsons Dis.* 10 (2020) 369–382, <https://doi.org/10.3233/JPD-191790>.
- [29] A. Wentink, C. Nussbaum-Krammer, B. Bukau, Modulation of amyloid states by molecular chaperones, *Cold Spring Harb. Perspect. Biol.* 11 (2019) a033969.
- [30] I.B. Bruinsma, K.A. Bruggink, K. Kinast, et al., Inhibition of α -synuclein aggregation by small heat shock proteins, *Protein Struct. Funct. Bioinf.* 79 (2011) 2956–2967.
- [31] M.M. Dedmon, J. Christodoulou, M.R. Wilson, et al., Heat shock protein 70 inhibits α -synuclein fibril formation via preferential binding to prefibrillar species, *J. Biol. Chem.* 280 (2005) 14733–14740.
- [32] S.F. Falsone, A.J. Kungl, A. Rek, et al., The molecular chaperone Hsp90 modulates intermediate steps of amyloid assembly of the Parkinson-related protein α -synuclein, *J. Biol. Chem.* 284 (2009) 31190–31199.
- [33] S. Pemberton, K. Madiona, L. Pieri, et al., Hsc70 protein interaction with soluble and fibrillar α -synuclein, *J. Biol. Chem.* 286 (2011) 34690–34699.
- [34] A. Rekas, C.G. Adda, J. Andrew Aquilina, et al., Interaction of the molecular chaperone α B-crystallin with α -Synuclein: effects on amyloid fibril formation and chaperone activity, *J. Mol. Biol.* 340 (2004) 1167–1183.
- [35] C. Roodveldt, C.W. Bertoncini, A. Andersson, et al., Chaperone proteostasis in Parkinson's disease: stabilization of the hsp70/ α -synuclein complex by hip, *EMBO J.* 28 (2009) 3758–3770.
- [36] F.A. Aprile, P. Arosio, G. Fusco, et al., Inhibition of α -synuclein fibril elongation by Hsp70 is governed by a kinetic binding competition between α -synuclein species, *Biochemistry* 56 (2017) 1177–1180.
- [37] D. Cox, E. Selig, M.D.W. Griffin, et al., Small Heat-shock Proteins Prevent α -synuclein aggregation via transient interactions and their efficacy is affected by the rate of aggregation, *J. Biol. Chem.* 291 (2016) 22618–22629.
- [38] X. Gao, M. Carroni, C. Nussbaum-Krammer, et al., Human Hsp70 disaggregate reverses Parkinson's-linked α -synuclein amyloid fibrils, *Mol. Cell.* 59 (2015) 781–793.
- [39] C. Nury, V. Redeker, S. Dautrey, et al., A novel bio-orthogonal cross-linker for improved protein/protein interaction analysis, *Anal. Chem.* 87 (2015)

- 1853–1860.
- [40] V. Redeker, S. Pemberton, W. Bienvenut, et al., Identification of protein interfaces between α -synuclein, the principal component of Lewy bodies in Parkinson disease, and the molecular chaperones human Hsc70 and the yeast Ssa1p, *J. Biol. Chem.* 287 (2012) 32630–32639.
- [41] C.A. Waudby, T.P.J. Knowles, G.L. Devlin, et al., The interaction of α B-crystallin with mature α -synuclein amyloid fibrils inhibits their elongation, *Biophys. J.* 98 (2010) 843–851.
- [42] JA Aquilina, JLP Benesch, OA Bateman, et al., Polydispersity of a mammalian chaperone: Mass spectrometry reveals the population of oligomers in B-crystallin, *Proc. Natl. Acad. Sci.* 100 (2003) 10611–10616.
- [43] H. Braak, K. Del Tredici, D. Sandmann-Keil, et al., Nerve cells expressing heat-shock proteins in Parkinson's disease, *Acta Neuropathol.* 102 (2001) 449–454.
- [44] A. Mainz, J. Peschek, M. Stavropoulou, et al., The chaperone α B-crystallin uses different interfaces to capture an amorphous and an amyloid client, *Nat. Struct. Mol. Biol.* 22 (2015) 898–905.
- [45] M. Zhang, M. Windheim, S.M. Roe, et al., Chaperoned ubiquitylation—crystal structures of the CHIP U box E3 ubiquitin ligase and a CHIP-Ubc13-Uev1a complex, *Mol. Cell.* 20 (2005) 525–538.
- [46] Y. Shin, J. Klucken, C. Patterson, et al., The Co-chaperone carboxyl terminus of Hsp70-interacting protein (CHIP) mediates α -synuclein degradation decisions between proteasomal and lysosomal pathways, *J. Biol. Chem.* 280 (2005) 23727–23734.
- [47] J.E. Tetzlaff, P. Putcha, T.F. Outeiro, et al., CHIP targets toxic α -synuclein oligomers for degradation, *J. Biol. Chem.* 283 (2008) 17962–17968.
- [48] M. Ghee, R. Melki, N. Michot, et al., PA700, the regulatory complex of the 26S proteasome, interferes with α -synuclein assembly, *FEBS J.* 272 (2005) 4023–4033.
- [49] A. Shevchenko, M. Wilm, O. Vorm, et al., Mass spectrometric sequencing of proteins from silver-stained polyacrylamide gels, *Anal. Chem.* 68 (1996) 850–858.
- [50] A.N. Shrivastava, V. Redeker, L. Pieri, et al., Clustering of Tau fibrils impairs the synaptic composition of α 3-Na⁺/K⁺ -ATPase and AMPA receptors, *EMBO J.* 38 (2019), e99871.
- [51] M. Götze, J. Pettelkau, S. Schaks, et al., StavroX-A software for analyzing crosslinked products in protein interaction studies, *J. Am. Soc. Mass Spectrom.* 23 (2012) 76–87.
- [52] I. Migneault, C. Dartiguenave, M.J. Bertrand, et al., Glutaraldehyde: behavior in aqueous solution, reaction with proteins, and application to enzyme cross-linking, *Biotechniques* 37 (2004) 790–802.
- [53] C. Hansen, E. Angot, A. Bergström, et al., α -Synuclein propagates from mouse brain to grafted dopaminergic neurons and seeds aggregation in cultured human cells, *J. Clin. Invest.* 121 (2011) 715–725.
- [54] E. Monsellier, V. Redeker, G. Ruiz-Arlandis, et al., Molecular interaction between the chaperone Hsc70 and the N-terminal flank of huntingtin exon 1 modulates aggregation, *J. Biol. Chem.* 290 (2015) 2560–2576.
- [55] V. Redeker, J. Bonnefoy, J.P. Le Caer, et al., A region within the C-terminal domain of Ure2p is shown to interact with the molecular chaperone Ssa1p by the use of cross-linkers and mass spectrometry, *FEBS J.* 277 (2010) 5112–5123.
- [56] R. Guerrero-Ferreira, N.M.I. Taylor, A.-A. Arteni, et al., Two new polymorphic structures of alpha-synuclein solved by cryo-electron microscopy, *Elife* 8 (2019) pii:e48907, <https://doi.org/10.7554/eLife.48907>.
- [57] H Dimant, D Ebrahimi-Fakhari, McLean PJ Molecular chaperones and co-chaperones in parkinson disease, *Neuroscientist* 18 (2012) 589–601.
- [58] I Lindberg, J Shorter, RL Wiseman, et al., Chaperones in Neurodegeneration, *J. Neurosci.* 35 (2015) 13853–13859.
- [59] Z Liu, C Wang, Y Li, et al., Mechanistic insights into the switch of α B-crystallin chaperone activity and self-multimerization, *J. Biol. Chem.* 293 (2018) 14880–14890.
- [60] GK a Hochberg, H. Ecroyd, C. Liu, et al., The structured core domain of α B-crystallin can prevent amyloid fibrillation and associated toxicity, *Proc. Natl. Acad. Sci. U. S. A.* 111 (2014) 200–203.
- [61] D. Cox, D.R. Whiten, J.W.P. Brown, et al., The small heat shock protein Hsp27 binds α -synuclein fibrils, preventing elongation and cytotoxicity, *J. Biol. Chem.* 293 (2018) 4486–4497.
- [62] L. Di, Strategic approaches to optimizing peptide ADME properties, *AAPS J.* 17 (2015) 134–143.