



Systemic AA amyloidosis caused by inflammatory hepatocellular adenoma

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Systemic AA Amyloidosis Caused by Inflammatory Hepatocellular Adenoma

TO THE EDITOR: Amyloid A (AA) systemic amyloidosis is a complication of chronic inflammatory diseases that is caused by the deposition of insoluble aggregates of cleaved N-terminal fragments of serum amyloid A (SAA) protein in tissues and organs throughout the body.¹ Under physiologic conditions, SAA protein is produced by hepatocytes during the acute inflammatory phase in response to various cytokines such as interleukin-6. SAA is also overexpressed by neoplastic hepatocytes in inflammatory hepatocellular adenomas, a specific molecular subtype of benign liver tumors.^{2,3}

Here, we describe a 49-year-old female patient who presented with diarrhea, rectal bleeding, and leg edema and received a diagnosis of systemic AA amyloidosis. Clinical characteristics of the patient are provided in Table S1 in the Supplementary Appendix, available with the full text of this letter at NEJM.org. An inflammatory hepatocellular adenoma was identified and resected, resulting in the improvement of amyloidosis-related symptoms, with progressive normalization of the serum C-reactive protein (Fig. 1A) and SAA levels. Unfortunately, the patient's kidney function did not improve, and she died 18 months after surgery from fulminant septic shock.

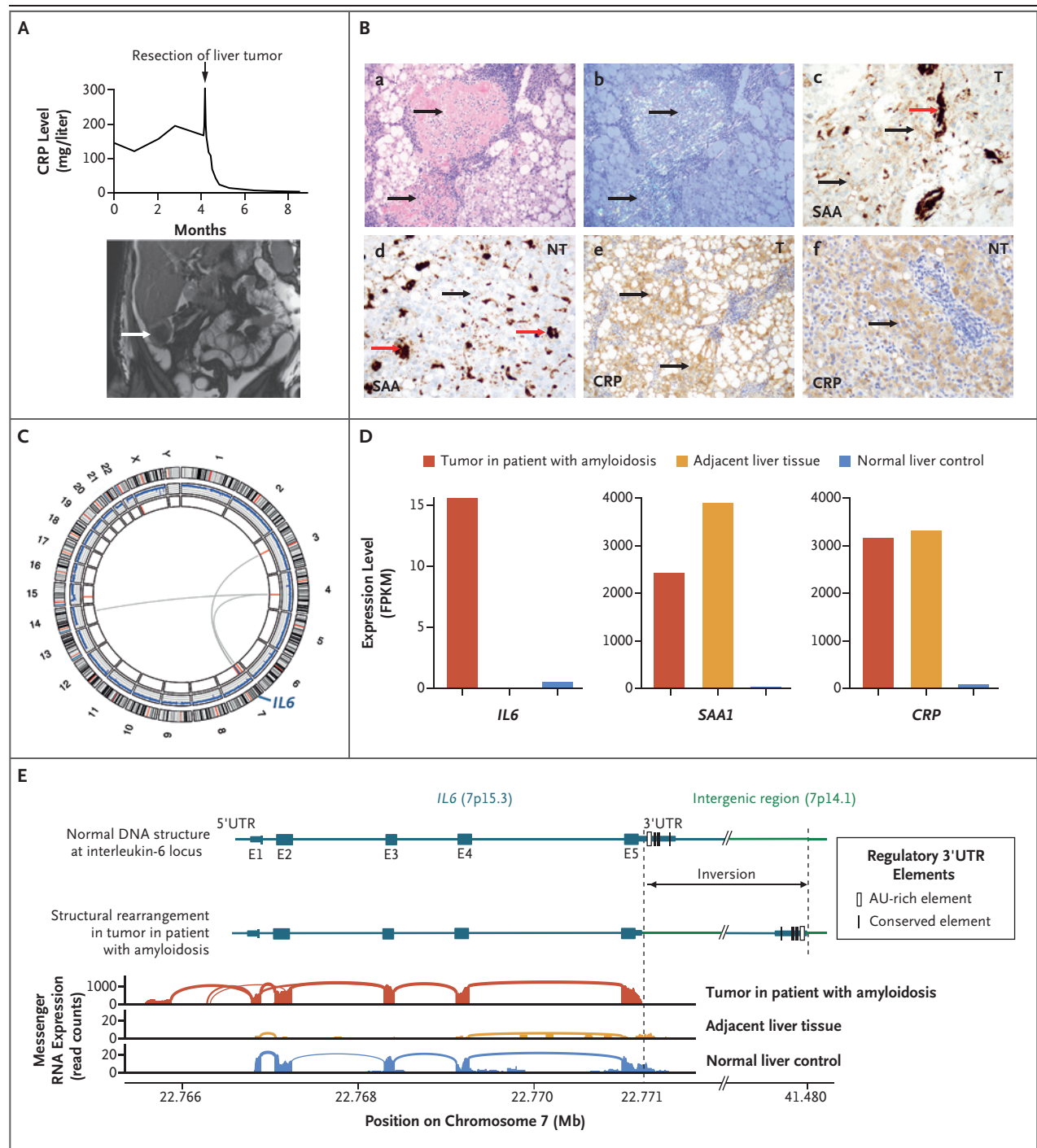
Whole-genome sequencing of the inflammatory hepatocellular adenoma revealed a complex structural rearrangement on chromosome 7 with an inversion leading to the truncation of the *IL6* 3' untranslated region (3'UTR), which comprises key sequences involved in the degradation of this unstable transcript (Fig. 1, and Tables S2 and S3

Figure 1 (facing page). Biologic, Imaging, Pathologic, and Molecular Findings in Our Patient.

As shown in Panel A, liver resection was followed by a clinically significant decrease in serum levels of C-reactive protein (CRP) (top), and magnetic resonance imaging showed a heterogeneous, 5-cm liver mass at the tip of segment VI on T₂-weighted sequences (bottom, arrow). As shown in Panel B, microscopic examination of the resected specimen revealed a well-differentiated tumor with massive amyloid deposits (arrows) (subpanel a, Congo red staining, low magnification) and typical yellow-green birefringence under polarized light (subpanel b, low magnification). Immunohistochemical tests showed positive staining for serum amyloid A (SAA) protein in neoplastic hepatocytes (black arrows) and amyloid deposits (red arrows) in the tumor (T) sample (subpanel c, high magnification); SAA expression was also observed in the adjacent nontumorous (NT) parenchyma (subpanel d, high magnification). CRP was expressed in the tumor (T) (subpanel e, low magnification) (neoplastic hepatocytes, black arrows) and the adjacent NT liver tissue (subpanel f, high magnification) (non-neoplastic hepatocytes, black arrows). As shown in Panel C, whole-genome sequencing of the tumor sample obtained from the patient revealed a cluster of somatic structural rearrangements at the *IL6* locus on chromosome 7. The blue line in the outer circle indicates copy number, and structural rearrangements are indicated in the inner circle. Red indicates deletion, blue inversion, gray interchromosomal translocation, and black classic chromatin staining of the cytobands. As shown in Panel D, although overexpression of *IL6* was specific to the tumor sample, the genes *SAA1* and *CRP* were also massively overexpressed in the adjacent liver tissue, as compared with normal liver tissue. FPKM denotes fragments per kilobase of exon per 1 million reads in the RNA sequencing experiment. As shown in Panel E, chromosomal inversion with one breakpoint located in the 3' untranslated region (UTR) of the gene *IL6* and the other breakpoint in an intergenic region at 7p14.1 led to the massive overexpression of an interleukin-6 transcript lacking regulatory 3'UTR elements in the patient's tumor (top, structural rearrangement identified by means of whole-genome sequencing; bottom, gene expression and transcript structure identified by means of RNA sequencing). AU denotes adenylate-uridylate.

in the Supplementary Appendix). RNA sequencing consistently showed a massive overexpression of an interleukin-6 transcript lacking all 3' regulatory elements; this overexpression was limited to the tumor area. However, in contrast to classic inflammatory hepatocellular adenoma, we observed overexpression of SAA in both the

tumor and nontumorous liver. Amyloid deposits were also identified in the adenoma and the adjacent parenchyma (Fig. 1). These results support an autonomous production of interleukin-6 by the inflammatory hepatocellular adenoma with activation of a paracrine Janus kinase–signal transducers and activators of transcription 3



(JAK–STAT3) pathway in the liver (Fig. S1 in the Supplementary Appendix).

In the literature, we identified eight additional patients with both AA amyloidosis and hepatocellular adenomas. In at least six of these patients, amyloid-related symptoms also improved after surgical resection of the liver tumor (Table S4 in the Supplementary Appendix). We reviewed the liver histologic findings of two patients from the literature and identified features that were similar to those of our patient; inflammatory hepatocellular adenoma with SAA expression by the tumor and the adjacent parenchyma was indeed identified in both cases from the literature (Fig. S2 in the Supplementary Appendix).^{4,5} Birefringent, SAA-positive amyloid deposits were also identified in the adenomas and adjacent liver tissue.

In conclusion, rare cases of inflammatory hepatocellular adenoma can induce massive SAA production by the tumor and nontumorous liver, leading to systemic AA amyloidosis. Molecular analyses identified a somatic rearrangement of the *IL6* 3'UTR leading to an autocrine interleukin-6 secretion by the tumor. We speculate that this subset of AA amyloidosis may be treatable by surgical resection of the liver tumor.

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Sodium Thiosulfate and Cisplatin-Induced Hearing Loss

TO THE EDITOR: Brock and colleagues (June 21 issue)¹ report the results of International Liver Tumor Strategy Group (SIOPEL) 6, a randomized, controlled trial of sodium thiosulfate for protec-

tion from cisplatin-induced hearing loss. In addition to confirming the efficacy of sodium thiosulfate, as previously reported in the Children's Oncology Group ACCL0431 trial,² SIOPEL 6 pro-