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1 ***Rhodobacter sphaeroides* methionine sulfoxide reductase P reduces R- and S-diastereomers of**
2 **methionine sulfoxide from a broad-spectrum of protein substrates**

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13

14 **Keywords:** Enzyme kinetics, methionine sulfoxide reductase, microbiology, molybdoenzyme, oxidative
15 stress, periplasm, protein oxidation, proteomics

16 **Abstract**

17 Methionine (Met) is prone to oxidation and can be converted to Met sulfoxide (MetO), which exists as
18 *R*- and *S*-diastereomers. MetO can be reduced back to Met by the ubiquitous methionine sulfoxide reductase
19 (Msr) enzymes. Canonical MsrA and MsrB were shown to be absolutely stereospecific for the reduction of
20 *S*- and *R*-diastereomer, respectively. Recently, a new enzymatic system, MsrQ/MsrP which is conserved in
21 all gram-negative bacteria, was identified as a key actor in the reduction of oxidized periplasmic proteins.
22 The haem-binding membrane protein MsrQ transmits reducing power from the electron transport chains to
23 the molybdoenzyme MsrP, which acts as a protein-MetO reductase. The MsrQ/MsrP function was well
24 established genetically, but the identity and biochemical properties of MsrP substrates remain unknown. In
25 this work, using the purified MsrP enzyme from the photosynthetic bacteria *Rhodobacter sphaeroides* as a
26 model, we show that it can reduce a broad spectrum of protein substrates. The most efficiently reduced
27 MetO are found in clusters of amino acid sequences devoid of threonine and proline on the C-terminal side.
28 Moreover, *R. sphaeroides* MsrP lacks stereospecificity as it can reduce both *R*- and *S*- diastereomers of
29 MetO, similarly to its *Escherichia coli* homolog, and preferentially acts on unfolded oxidized proteins.
30 Overall, these results provide important insights into the function of a bacterial envelop protecting system,
31 which should help understand how bacteria cope in harmful environments.

32

33 **Abbreviations:** BV, benzyl viologen; DMSO, dimethylsulfoxide; DTT, dithiothreitol; ESI-MS,
34 Electrospray Ionization-Mass spectrometry; H₂O₂, hydrogen peroxide;
35 HEPES4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; MES, 2-(N-morpholino)ethanesulfonic acid;
36 MetO, methionine sulfoxide; Met-*R*-O, *R*-diastereomer of MetO; Met-*S*-O, *S*-diastereomer of MetO; MS,
37 mass spectrometry, Msr, MetO reductase, NADPH, Nicotinamide adenine dinucleotide phosphate; NaOCl,
38 sodium hypochlorite.

39 **Introduction**

40 Aerobic life exposes organisms to reactive oxygen species (ROS) derived from molecular oxygen, such
41 as hydrogen peroxide (H₂O₂) or singlet oxygen (¹O₂). Bioenergetic chains are important sources of these
42 intracellular ROS. H₂O₂ is principally produced during respiration [1] and ¹O₂ arises from photosynthesis
43 [2]. In most organisms, these oxidative molecules act as signaling messengers playing major roles in
44 numerous physiological and pathological states. Furthermore, their production and elimination are tightly
45 regulated [3]. However, numerous stresses can affect ROS homeostasis and increase their intracellular
46 concentration to excessive values leading to uncontrolled reactions with sensitive macromolecules [4]. For
47 instance, photosynthetic organisms, such as plants or the purple bacteria *Rhodobacter sphaeroides* can
48 experience photo-oxidative stress in which unbalance between incident photons and photosynthetic electron
49 transfer generate detrimental accumulation of ¹O₂ [5]. Moreover, production of ROS could be used
50 advantageously in a defensive strategy against potential pathogenic invaders. For instance, neutrophils
51 produce the strong oxidant hypochlorite (ClO⁻) from H₂O₂ and chlorine ions to eliminate bacteria and fungi
52 [3]. Because of their abundance in cells, proteins are the main targets of oxidation [6]. Methionine (Met) is
53 particularly prone to oxidation and the reaction of Met with an oxidant leads to the formation of Met
54 sulfoxide (MetO), which exists as two diastereomers *R* (Met-*R*-O) and *S* (Met-*S*-O). Further oxidation can
55 then lead to the formation of Met sulfone (MetO₂) [7,8]. As opposed to most oxidative modifications on
56 amino acids, the formation of MetO is reversible, and oxidized proteins can be repaired thanks to methionine
57 sulfoxide reductases (Msr) enzymes that principally exist in two types, MsrA and MsrB. These enzymes,
58 present in almost all organisms, did not evolve from a common ancestral gene and possess an absolute
59 stereospecificity towards their substrates. Indeed, MsrA can reduce only Met-*S*-O [7,9–12] whereas MsrB
60 acts only on Met-*R*-O [10–14]. This strict stereospecificity was enzymatically demonstrated using
61 chemically prepared Met-*R*-O and Met-*S*-O from racemic mixtures of free MetO or by using HPLC methods
62 allowing discrimination of both diastereomers. A structural explanation was also provided by deciphering
63 the mirror images of their active sites, in which only one MetO diastereomer can be accommodated [10].

64 While MsrA can reduce Met-S-O, whether as a free amino acid or included in proteins, MsrB is specialized
65 in the reduction of protein-bound Met-R-O, and both are more efficient on unfolded oxidized proteins
66 [15,16]. Eukaryotic Msrs are important actors in oxidative stress protection, aging and neurodegenerative
67 diseases in animals [17], during environmental stresses and seed longevity in plants [18,19]. In bacteria,
68 MsrA and MsrB are generally located in the cytoplasm [3], except in *Neisseria* or *Streptococcus* species,
69 for which MsrA and MsrB enzymes can be addressed to the envelope [20,21]. They play a role in protecting
70 against oxidative stress and as virulence factors [3].

71 Beside these stereotypical Msrs found in all kinds of organisms, several other enzymes can catalyze
72 MetO reduction, principally in bacteria. For instance, numerous bacteria as well as unicellular eukaryotes,
73 such as *Saccharomyces cerevisiae*, possess another type of absolutely stereospecific Msr, called free-R-Msr
74 (fRMsr) or MsrC, which is specialized in the reduction of the free form of Met-R-O [22,23]. As MsrA and
75 MsrB, the fRMsr uses thiol-based chemistry and the reducing power coming from NADPH to reduce its
76 substrate [22,23].

77 In bacteria, several molybdenum cofactor-containing enzymes were also shown to be able to reduce
78 oxidized Met. Particularly, the biotin sulfoxide reductase BisC, or its homolog TorZ/BisZ, specifically
79 reduce the free form of Met-S-O, in the *Escherichia coli* cytoplasm and the *Haemophilus influenza*
80 periplasm, respectively [24,25]. Moreover, *E. coli* DMSO reductase reduces a broad spectrum of substrates,
81 including MetO [26], while the *R. sphaeroides* homolog was shown to be absolutely stereospecific towards
82 S-enantiomer of several alkyl aryl sulfoxides [27]. Finally, another molybdoenzyme, MsrP (formerly known
83 as YedY), was recently identified as a key player of MetO reduction in the periplasm [28,29]. MsrP was
84 shown to be induced by exposure to the strong oxidant hypochlorite (ClO⁻) and to reduce MetO on several
85 abundantly present periplasmic proteins in *E. coli* [28] or on a Met-rich protein in *Azospira suillum* [29]. A
86 most striking feature of *E. coli* MsrP (EcMsrP) is that, contrary to all known methionine sulfoxide
87 reductases, it seems capable of reducing both Met-R-O and Met-S-O [28]. The cistron, *msrP*, belongs to an
88 operon together with the cistron encoding the transmembrane protein MsrQ, which is responsible for the

89 electron transfer to MsrP from the respiratory chain. Of note, the cytosolic flavin reductase Fre was proposed
90 as a potential alternative electron-carrier to MsrQ [30]. The operon is conserved in the genome of most
91 gram-negative bacteria suggesting that the MsrP/Q system is very likely a key player for general protection
92 in the bacterial envelop against deleterious protein oxidation [28,29]. *R. sphaeroides* MsrP (RsMsrP) shares
93 50% of identical amino acid residues with EcMsrP and transcriptomic analyses evidenced that *RsmsrP* is
94 strongly induced under high-light conditions, suggesting a putative role in protecting the periplasm against
95 $^1\text{O}_2$ [31].

96 In this paper, we describe the biochemical characterization of RsMsrP regarding its substrate specificity.
97 Using kinetic activity experiments and mass spectrometry analysis, we show that RsMsrP is a very efficient
98 protein-bound MetO reductase, which lacks stereospecificity and preferentially acts on unfolded oxidized
99 proteins. Proteomic analysis indicates that it can reduce a broad spectrum of proteins in the *R. sphaeroides*
100 periplasm, and that Met sensitive to oxidation and efficiently reduced by RsMsrP are found in clusters and
101 in specific amino acids sequences.

102

103 **Material and methods**

104 **Production and purification of recombinant proteins**

105 Recombinant MsrP was produced similarly to the previously described protocol [32]. Briefly, *R.*
106 *sphaeroides* f sp. *denitrificans* IL106 *dmsA*⁻ strain carrying the pSM189 plasmid for production of a
107 periplasmic MsrP with a 6-His N-terminal tag was grown in 6-liter culture under semi-aerobic conditions
108 in Hutner medium until late exponential phase. The periplasmic fraction was extracted and loaded on a
109 HisTrap column (GE Healthcare), MsrP was then eluted by an imidazole step gradient. MsrP solution was
110 concentrated using 15-ml Amicon[®] Ultra concentrators with 10-kDa cutoff (Millipore), desalted with
111 Sephadex G-25 in PD-10 Desalting Columns (GE Healthcare). The protein concentration was adjusted to 1
112 mg.ml⁻¹ in 30 mM Tris-HCl pH 7.5, 500 mM NaCl, the Tobacco Etch Virus (TEV) protease was added
113 (1:80 TEV:RsMsrP mass ratio) and the solution incubated overnight at room temperature to remove the
114 polyhistidine tag. Untagged RsMsrP was purified on a second HisTrap column, then concentrated and
115 desalted in 50 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) pH 8.0. Protein solution
116 was then loaded on a Superdex[™] 200 10/30 gel filtration column equilibrated with 30 mM Tris-HCl pH
117 7.5. Main fractions were pooled and applied to a MonoQ[™] 4.6/100 PE (GE Healthcare). RsMsrP was then
118 eluted using a linear NaCl gradient (0 to 500 mM). Fractions were analyzed on SDS-PAGE using
119 NuPAGE[™], 10 % Bis-Tris gels with MES-SDS buffer (ThermoFisher). Recombinant MsrA, MsrB,
120 Thioredoxin Reductase (TR) 1, Thioredoxin 1 (Trx1) from *Saccharomyces cerevisiae* with polyhistidine
121 tags, as well as the glutathione-S-transferase (GST) from *Schistosoma japonicum*, were produced and
122 purified as previously described [15]. Protein concentrations were determined spectrophotometrically using
123 specific molar extinction coefficients at 280 nm: 6-His-RsMSRP, 56,380 M⁻¹.cm⁻¹; untagged RsMsrP,
124 54,890 M⁻¹.cm⁻¹; MsrA, 34,630 M⁻¹.cm⁻¹; MsrB, 24,325 M⁻¹.cm⁻¹; TR1, 24,410 M⁻¹.cm⁻¹; Trx1, 9,970
125 M⁻¹.cm⁻¹; GST, 42,860 M⁻¹.cm⁻¹, bovine β-casein (Sigma-Aldrich), 11,460 M⁻¹.cm⁻¹ and chicken lysozyme
126 (Sigma-Aldrich), 32,300 M⁻¹.cm⁻¹. Protein solutions were stored at -20°C until further use.

127

128 **Peptides**

129 Ser-Met(O)-Ser, QWGAGM(O)QAEED and TTPGYM(O)EEWNK peptides were obtained from
130 GenScript® (Hong-Kong).

131 **Preparation of oxidized bovine β -casein and its Met-R-O and Met-S-O containing counterparts**

132 For oxidation, bovine β -casein was prepared in Phosphate Buffered Saline (PBS) at 1 mg.ml⁻¹ in the
133 presence of 200 mM H₂O₂ and incubated overnight at room temperature. H₂O₂ was removed by desalting
134 using a PD-10 column and the protein solution was concentrated with 10-kDa cutoff Amicon® Ultra
135 concentrator. Oxidized GST was similarly prepared using 100 mM H₂O₂. To prepare Met-R-O containing
136 β -casein, a solution of oxidized β -casein was incubated in 30 mM Tris-HCl pH 8 at a final concentration of
137 6.5 mg.ml⁻¹ (260 μ M) in the presence of 25 mM dithiothreitol (DTT) with 10 μ M MsrA and incubated
138 overnight at room temperature. The solution was diluted 10-fold in 30 mM Tris-HCl pH 8 and passed over
139 a HisTrap column to remove the his-tagged MsrA. After concentration, the DTT was removed by desalting
140 using a PD-10 column. Met-S-O containing β -casein was prepared similarly replacing the MsrA by the
141 MsrB (14 μ M). The protein solutions were concentrated with a 10-kDa cutoff Amicon® Ultra concentrator
142 and the final concentration was determined spectrophotometrically. Protein solutions were stored at -20°C
143 until further use.

144 **Enzymatic activity and apparent stoichiometry measurements**

145 RsMsrP reductase activity was measured as described in ref. [32] with a few modifications. Benzyl
146 viologen (BV) was used as an electron donor and its consumption was followed at 600 nm using an UVmc1®
147 spectrophotometer (SAFAS Monaco) equipped with optic fibers in a glovebox workstation (MBRAUN
148 Labstar) flushed with nitrogen. We determined the specific molar extinction coefficient of benzyl viologen
149 at 8,700 M⁻¹.cm⁻¹ in 50 mM 2-(*N*-morpholino)ethanesulfonic acid (MES), pH 6.0 buffer. Each reaction
150 mixture (1 ml or 0.5 ml) contained 0.2 mM BV reduced with sodium dithionite, and variable concentrations
151 of substrates in 50 mM MES, pH 6.0 buffer.

152 Reactions were started by the addition of RsMsrP enzyme (10 to 46 nM). Reduction of MetO rates were
153 calculated from the $\Delta A_{600\text{ nm}}$ slopes respecting a stoichiometry of 2 (2 moles of BV are oxidized for 1 mole
154 of MetO reduced). Thus, the activity values presented as k_{cat} or k_{obs} (s^{-1}) represent the number of moles of
155 MetO reduced per mole of enzyme per second.

156 The apparent stoichiometry was determined similarly, using subsaturating concentrations of substrates:
157 1–10 μM oxidized β -casein, 1–10 μM Met-*R*-O containing β -casein and 1.5–15 μM Met-*S*-O containing
158 β -casein. The amount of oxidized BV was determined 1 hour after the addition of the RsMsrP (46 nM) by
159 subtracting the final $A_{600\text{ nm}}$ value from the initial one. Controls were done without the RsMsrP enzyme, or
160 without the MetO-containing substrate. Quantities of MetO reduced were plotted as a function of substrate
161 quantities and the apparent stoichiometry was obtained from the linear regression slope.

162 MsrA and MsrB activities were measured following the spectrophotometrical consumption of NADPH
163 at 340 nm using the thioredoxin system similarly to the previously described protocol [15]. A 500- μl reaction
164 cuvette contained 200 μM NADPH, 2 μM TR1, 25 μM Trx1 and 5 μM MsrA or MsrB and 100 μM oxidized
165 β -casein. Production of Met was calculated respecting a stoichiometry of 1 (1 mole of NADPH is oxidized
166 for 1 mole of Met produced).

167 Analysis and kinetics parameters determination were done using GraphPad® Prism 4.0 software (La
168 Jolla, CA, USA).

169

170 **Electrospray ionization/Mass spectrometry analysis of purified proteins**

171 For oxidation, bovine β -casein (5 $\text{mg}\cdot\text{ml}^{-1}$) in 50 mM HEPES, pH 7.0, was incubated overnight at room
172 temperature with H_2O_2 (50 mM). H_2O_2 was removed by desalting using a PD-10 column and the protein
173 solution was concentrated with a 10-kDa cutoff Amicon® Ultra concentrator. Oxidized β -casein (100 μM)
174 was reduced by addition of 44 nM MsrP in a reaction mixture containing 50 mM HEPES pH 7.0, 0.8 mM
175 BV and 0.2 mM sodium dithionite. After two hours of reaction in the glove-box, the repaired β -casein was
176 analyzed by electrospray ionization/mass spectrometry in comparison to non-oxidized and oxidized
177 β -casein. Analyses were performed on a MicroTOF-Q Bruker (Wissembourg, France) with an electrospray

178 ionization source. Samples were desalted in ammonium acetate buffer (20 mM) and concentrated with a
179 30-kDa cutoff Amicon® Ultra concentrator prior to analyses. Samples were diluted in CH₃CN/H₂O
180 (1/1-v/v), 0.2% Formic Acid (Sigma). Samples were continuously infused at a flow rate of 3 μL.min⁻¹. Mass
181 spectra were recorded in the 50-7000 mass-to-charge (*m/z*) range. MS experiments were carried out with a
182 capillary voltage set at 4.5 kV and an end plate off set voltage at 500 V. The gas nebulizer (N₂) pressure
183 was set at 0.4 bars and the dry gas flow (N₂) at 4 L.min⁻¹ at a temperature of 190 °C. Data were acquired in
184 the positive mode and calibration was performed using a calibrating solution of ESI Tune Mix (Agilent) in
185 CH₃CN/H₂O (95/5-v/v). The system was controlled with the software package MicrOTOF Control 2.2 and
186 data were processed with DataAnalysis 3.4.

187

188 **Generation of *R. sphaeroides* 2.4.1 *msrP* mutant**

189 The *msrPQ* operon was amplified from *R. sphaeroides* 2.4.1 genomic DNA with the primers
190 5'-AGATCGACACGCCATTCACC-3' and 5'-TCGGTGAGGCGCTATCTAGG-3'. The 2.2 kb PCR
191 product was cloned into pGEMT Easy (Promega). An omega cartridge encoding resistance to streptomycin
192 and spectinomycin [33] was then cloned into the *Bam*HI site of *msrP*. The resulting plasmid was digested
193 with *Sac*I and the fragment containing the disrupted *msrP* gene was cloned into pJQ200mp18 [34]. The
194 obtained plasmid, unable to replicate in *R. sphaeroides*, was transferred from *E. coli* by conjugation. The
195 occurrence of a double-crossing over event was confirmed by PCR and absence of the protein from the
196 SDS-PAGE profile.

197 **Preparation of periplasmic samples for proteomics analysis**

198 *R. sphaeroides* 2.4.1 *msrP* mutant was grown under semi-aerobic conditions. Periplasmic extract was
199 prepared as previously described [32] by cells incubation in 50 mM HEPES pH 8.0, 0.45 M sucrose, 1.3
200 mM Ethylenediaminetetraacetic acid (EDTA) and 1 mg.ml⁻¹ chicken lysozyme. For Met oxidation, the
201 periplasmic extract (0.7 mg.ml⁻¹) was incubated with 20 mM *N*-Ethylmaleimide (NEM) and 2 mM NaOCl
202 (Sigma-Aldrich) in 50 mM HEPES pH 8.0, 50 mM NaCl for 10 min at room temperature. NaOCl was

203 removed by desalting using a PD-10 column and buffer was changed to 50 mM MES pH 6.0. The protein
204 solution was concentrated with a 3-kDa cutoff Amicon® Ultra concentrator. Three reaction mixtures were
205 prepared in the glove box containing 35 µl of periplasmic extract, 1 mM benzyl viologen, 2 mM dithionite
206 in 50 mM MES pH 6.0. The protein concentration in each reaction was 2.5 mg.ml⁻¹. The first reaction
207 contained non-oxidized periplasmic extract, the second and third ones contained oxidized periplasmic
208 extract. For the third reaction (repaired periplasm) 10 µM RsMsrP was added. The reactions were incubated
209 for three hours at room temperature.

210 **Trypsin proteolysis and tandem mass spectrometry**

211 Protein extracts were immediately subjected to denaturing PAGE electrophoresis for 5 min onto a 4–
212 12% Bis-Tris gradient 10-well NuPAGE™ gel (Thermofisher). The proteins were stained with Coomassie
213 Blue Safe solution (Invitrogen). Polyacrylamide bands corresponding to the whole proteomes were sliced
214 and treated with iodoacetamide followed by trypsin as previously recommended [35]. Briefly, each band
215 was destained with ultra-pure water, reduced with DTT, treated with iodoacetamide, and then proteolyzed
216 with Gold Mass Spectrometry Grade Trypsin (Promega) in the presence of 0.01% ProteaseMAX surfactant
217 (Promega). Peptides were immediately subjected to tandem mass spectrometry as previously recommended
218 to avoid methionine oxidation [36]. The resulting peptide mixtures were analyzed in a data-dependent mode
219 with a Q-Exactive HF tandem mass spectrometer (Thermo) coupled on line to an Ultimate 3000
220 chromatography system chromatography (Thermo) essentially as previously described [37]. A volume of
221 10 µL of each peptide sample was injected, first desalted with a reverse-phase Acclaim PepMap 100 C18
222 (5 µm, 100 Å, 5 mm x 300 µm i.d., Thermo) precolumn and then separated at a flow rate of 0.2 µL.min⁻¹
223 with a nanoscale Acclaim PepMap 100 C18 (3 µm, 100 Å, 500 mm x 300 µm i.d., Thermo) column using
224 a 150 min gradient from 2.5 % to 25 % of CH₃CN, 0.1% formic acid, followed by a 30 min gradient from
225 25% to 40% of CH₃CN, 0.1% formic acid. Mass determination of peptides was done at a resolution of
226 60,000. Peptides were then selected for fragmentation according to a Top20 method with a dynamic

227 exclusion of 10 sec. MS/MS mass spectra were acquired with an AGC target set at $1.7 \cdot 10^5$ on peptides with
228 2 or 3 positive charges, an isolation window set at 1.6 m/z , and a resolution of 15,000.

229 **MS/MS spectrum assignment, peptide validation and protein identification**

230 Peak lists were automatically generated from raw datasets with Proteome Discoverer 1.4.1 (Thermo) and
231 an in-house script with the following options: minimum mass (400), maximum mass (5,000), grouping
232 tolerance (0), intermediate scans (0) and threshold (1,000). The resulting .mgf files were queried with the
233 Mascot software version 2.5.1 (Matrix Science) against the *R. sphaeroides* 241 annotated genome database
234 with the following parameters: full-trypsin specificity, up to 2 missed cleavages allowed, static modification
235 of carbamidomethylated cysteine, variable oxidation of methionine, variable deamidation of asparagine and
236 glutamine, mass tolerance of 5 ppm on parent ions and mass tolerance on MS/MS of 0.02 Da. The decoy
237 search option of Mascot was activated for estimating the false discovery rate (FDR) that was below 1%.
238 Peptide matches with a MASCOT peptide score below a P value of 0.05 were considered. Proteins were
239 validated when at least two different peptides were detected. The FDR for proteins was below 1% as
240 estimated with the MASCOT reverse database decoy search option.

241 **Ice logo analysis**

242 Ice logo analysis were performed using the IceLogo server
243 (<http://iomics.ugent.be/icelogoserver/index.html>) [38].

244

245

246 **Results**

247 **The *R. sphaeroides* MsrP is an efficient protein-MetO reductase**

248 The results showing that the EcMsrP is a protein-bound MetO reductase, able to reduce both *R*- and
249 *S*-diastereomer of MetO [28] prompted us to evaluate whether these properties are conserved for RsMsrP.
250 As the EcMsrP was determined to be 5-fold less efficient in reducing the Met-*S*-O than Met-*R*-O, and
251 knowing that all previously identified MetO reductases were absolutely stereospecific towards one
252 enantiomer, we thought that it cannot be excluded that a protein contamination might explain the apparent
253 ability of the EcMsrP to reduce the Met-*S*-O [28]. Such potential Met-*S*-O reductase contaminant should be
254 able to use benzyl viologen (BV) as electron provider in activity assays and a good candidate is the
255 periplasmic DMSO reductase [26,27]. Thus, we prepared the recombinant RsMsrP from a *R. sphaeroides*
256 strain devoid of the *dorA* gene encoding the catalytic subunit of the DMSO reductase [32]. After purification
257 on Ni-affinity column and removal of the polyhistidine tag, the mature enzyme was purified by gel filtration,
258 followed by strong anion exchange, yielding a highly pure enzyme (Supplementary Figure S1).

259 After optimal pH determination showing that RsMsrP acts efficiently between pH 5.5 and 8.0
260 (Supplementary Figure S2), we determined the kinetic parameters of RsMsrP using BV as an electron
261 provider and several model substrates: the free amino acid MetO, a synthetic tripeptide Ser-MetO-Ser and
262 the oxidized bovine β -casein (Table 1). The β -casein contains 6 Met, it is intrinsically disordered, and was
263 shown as an efficient substrate for the yeast MsrA and MsrB, after oxidation [15] (see also Supplementary
264 Figure S3). Commercial β -casein contains a mixture of genetic variants, appearing as multiple peaks on
265 mass spectrometry (MS) spectra (Figure 1A). After oxidation with H₂O₂, MS analysis confirmed an increase
266 in mass of 96 Da for each peak, very likely corresponding to the addition of 6 oxygen atoms on the Met
267 residues (Figure 1B). Using the free MetO, we determined a k_{cat} of $\sim 122 \text{ s}^{-1}$ and a K_{m} of $\sim 115,000 \mu\text{M}$,
268 yielding a catalytic efficiency ($k_{\text{cat}}/K_{\text{m}}$) of $\sim 1,000 \text{ M}^{-1} \cdot \text{s}^{-1}$ (Table 1). With the Ser-MetO-Ser peptide, the k_{cat}
269 and the K_{m} values were $\sim 108 \text{ s}^{-1}$ and $\sim 13,000 \mu\text{M}$, and thus the $k_{\text{cat}}/K_{\text{m}}$ was $\sim 8,300 \text{ M}^{-1} \cdot \text{s}^{-1}$. Compared to
270 free MetO, the ~ 8 -fold increase in catalytic efficiency is due to the lower K_{M} , and thus this indicates that

271 the involvement of the MetO in peptide bonds increases its ability to be reduced by RsMsP. With the
272 oxidized β -casein, the k_{cat} and the K_m were $\sim 100 \text{ s}^{-1}$ and $\sim 90 \text{ }\mu\text{M}$, respectively. The k_{cat}/K_m was thus \sim
273 $1,000,000 \text{ M}^{-1}\cdot\text{s}^{-1}$. This value, 4 orders of magnitude higher than the one determined with free MetO,
274 indicates that the oxidized protein is a far better substrate for RsMsP. Moreover, even assuming that all
275 MetO in the oxidized β -casein were equal substrates for the RsMsP and thus multiplying the K_M by 6, the
276 catalytic efficiency obtained ($\sim 175,000 \text{ M}^{-1}\cdot\text{s}^{-1}$) remained ~ 175 -fold higher for the oxidized protein than
277 for the free amino acid. These results indicate that the RsMsP acts effectively as a protein-MetO reductase.

278 **RsMsP reduces both Met-*R*-O and Met-*S*-O of an oxidized model protein**

279 To determine whether the RsMsP can reduce both MetO diastereomers, we chose the oxidized bovine
280 β -casein as a model substrate because it was efficiently reduced by the yeast MsrA and MsrB indicating the
281 presence of both *R* and *S* diastereomers of MetO [15]. After oxidation with H_2O_2 , we treated the protein
282 with MsrA and MsrB, taking advantage of their stereospecificity, to obtain protein samples containing only
283 the Met-*R*-O (“ *β -casein-R-O*”) or the Met-*S*-O (“ *β -casein-S-O*”), respectively. The absence of one or the
284 other diastereomer of MetO was validated by the absence of remaining Msr activity (Supplementary Figure
285 3). These three forms, containing either two or only one diastereomer of MetO, were tested as substrate for
286 RsMsP (Figure 2). We measured a k_{cat} of $\sim 45 \text{ s}^{-1}$ with the oxidized β -casein, which decreased to ~ 30 and
287 to 5 s^{-1} for the β -casein containing the *R* or the *S* sulfoxide, respectively. This result shows that RsMsP can
288 reduce both diastereomers of MetO, but appears 6-fold less efficient to reduce Met-*S*-O than Met-*R*-O.

289 From this result, we postulated that RsMsP should be able to reduce all MetO in the oxidized β -casein,
290 as this protein was intrinsically disordered and thus all MetO were very likely accessible. We evaluated this
291 hypothesis by mass spectrometry analysis. When incubated with RsMsP, the mass of the oxidized protein
292 decreased by 96 Da, showing that all MetO were reduced (Figure 1C). Altogether, these results clearly
293 showed that RsMsP was able to reduce both *R*- and *S*-diastereomers of MetO contained in the oxidized
294 β -casein, and thus lacked stereospecificity.

295

296 **The RsMsrP preferentially reduces Met-*R*-O but acts effectively on Met-*S*-O too**

297 To gain insight into the substrate preference of RsMsrP toward one of the diastereomers of MetO, we
298 performed kinetics analysis using the oxidized β -casein containing the *R* or *S* diastereomers of MetO (Table
299 1; Supplementary Figure S4). With the protein containing only the *R*-diastereomer of MetO
300 (“ β -casein-*R*-O”), we determined a k_{cat} of $\sim 50 \text{ s}^{-1}$, a K_m of $\sim 50 \mu\text{M}$ and thus a catalytic efficiency of \sim
301 $950,000 \text{ M}^{-1} \cdot \text{s}^{-1}$. In the case of the protein containing only the Met-*S*-O (“ β -casein-*S*-O”), the k_{cat} and K_m
302 were $\sim 8 \text{ s}^{-1}$ and of $\sim 50 \mu\text{M}$, respectively. This yielded a catalytic efficiency of $142,000 \text{ M}^{-1} \cdot \text{s}^{-1}$. This value,
303 ~ 7 fold lower than the one obtained with the β -casein-*R*-O, was due to the lower k_{cat} as the K_m was not
304 changed. These values seem to indicate that the RsMsrP preferentially reduced the *R* than the *S* diastereomer
305 of MetO in the oxidized β -casein. However, as we could not exclude that the proportion of Met-*R*-O was
306 higher than the proportion of Met-*S*-O in the protein, we developed an assay to estimate the number of MetO
307 reduced by RsMsrP in the three forms of oxidized β -casein. We measured the total moles of BV consumed
308 for the reduction of all MetO using subsaturating concentrations of the oxidized protein. Practically, the
309 absorbance at 600 nm was measured before and 90 min after substrate addition. As two moles of BV are
310 consumed per mole of MetO reduced, we obtained the apparent stoichiometry of RsMsrP toward the
311 oxidized protein by performing a linear regression on the straight part of the line and taking the slope, which
312 defines the amount of MetO reduced as a function of substrate concentration (Figure S5). The values
313 determined were ~ 4.6 , ~ 3.2 and ~ 1.8 for the oxidized β -casein, the β -casein-*R*-O and the β -casein-*S*-O,
314 respectively. In the case of the oxidized β -casein, we expected a value of 6 based on the data obtained by
315 mass spectrometry (Figure 1). This may have been due to the heterogeneity of the oxidized β -casein (all
316 Met were not initially fully oxidized) and/or to a too short time of incubation (all MetO were not fully
317 reduced, as indicated by the presence of a peak corresponding to a portion of β -casein not fully reduced in
318 Figure 1C). To compare the catalytic parameters, the data was normalized by multiplying the K_m by the
319 apparent stoichiometries, yielding values per reduced MetO, thereby allowing the removal of variation due
320 to the different numbers of reduced Met-*R*-O or Met-*S*-O. The catalytic efficiencies were thus 230,000,
321 300,000 and $80,000 \text{ M}^{-1} \cdot \text{s}^{-1}$ for the oxidized β -casein, the β -casein-*R*-O and the β -casein-*S*-O, respectively

322 (Table 1). The highest value was that obtained for the β -casein containing only the *R* form of MetO,
323 indicating that this diastereomer was the preferred substrate for RsMsP. However, the value obtained with
324 the β -casein-*S*-O was only less than 4-fold lower, showing that RsMsP can also act effectively on the
325 Met-*S*-O.

326 **RsMsP can reduce a broad spectrum of periplasmic proteins**

327 To identify potential periplasmic substrates of RsMsP and gain insight into its substrate specificity, we
328 applied a high-throughput shotgun proteomic strategy. Periplasmic proteins from the *mrsP* *R. sphaeroides*
329 mutant were extracted, oxidized with NaOCl and then reduced *in vitro* with recombinant RsMsP. Untreated
330 periplasmic proteins, oxidized periplasmic proteins and RsMsP-treated oxidized periplasmic proteins were
331 analyzed by semi-quantitative nanoLC–MS/MS. All experiments were done systematically for 3 biological
332 replicates and resulted in the identification of 362,700 peptide-to-spectrum matches. From all 11,320
333 individual peptide sequences, we identified 2,553 unique Met belonging to 720 proteins. The overall
334 percentages of Met oxidation were ~ 35%, ~ 71% and ~ 40% for proteins from the periplasm extract, the
335 oxidized periplasm extract and the RsMsP-repaired proteins, respectively (Supplementary Table 1). This
336 first result indicates that RsMsP is very likely able to reduce MetO from numerous proteins and to restore
337 an oxidation rate similar to the one of the periplasmic extract that has not undergone any oxidation.

338 The identification of preferential RsMsP substrates requires the precise comparison of the oxidation
339 state of Met residues from periplasmic proteins before and after the action of the enzyme. After tryptic
340 digestion, since most of the Met/MetO-containing peptides were found in low abundance (*i.e.* with very low
341 spectral counts), we focused on the proteins robustly detected in all samples. We selected the Met-containing
342 peptides for which at least 10 spectral counts were detected in two replicates for each condition (*i.e.*
343 untreated periplasm, oxidized periplasm and repaired oxidized periplasm) and at least 7 spectral counts were
344 found in the third replicate. This restricted the dataset to 202 unique Met belonging to 70 proteins
345 (Supplementary Table 2). Overall percentage of Met oxidation (calculated as the number of spectral counts
346 for a MetO-containing peptide vs. the total number of spectral count for this peptide) varied from 2% to

347 87%, from 9% to 100% and from 4% to 91% in the periplasm, oxidized periplasm and repaired oxidized
348 periplasm, respectively. Comparison of Met-O containing peptides between oxidized and RsMsrP treated
349 samples indicates that the percentage of reduction varied from 100 % to no reduction at all. Eleven MetO
350 were not reduced and 22 were reduced at more than 75 % (only 2 at 100 %). The percentage of reduction
351 for the remaining majority of MetO was almost uniformly distributed between inefficient (less than 25 %)
352 to efficient (75% or more) reduction (Figure 3A).

353 No clear evidence of sequence or structure characteristic arose from these 70 identified proteins, neither
354 in term of size or in Met content (Supplementary Table 2). The periplasmic chaperone SurA, the
355 peptidyl-prolyl cis-trans isomerase PpiA, the thiol-disulfide interchange protein DsbA, the
356 spermidine/putrescine-binding periplasmic protein PotD and the ProX protein were previously proposed as
357 potential substrates of EcMsrP [28]. All these proteins contain at least one MetO amongst the most
358 efficiently reduced by RsMsrP (Supplementary Table 2), indicating that they are potential conserved
359 substrates of the MsrP enzymes in *E. coli* and *R. sphaeroides*, and very likely in numerous other
360 gram-negative bacteria.

361 The sensitivity to oxidation of the Met belonging to these 70 proteins, and their efficiency of reduction
362 by RsMsrP show a wide range of variation, from Met highly sensitive to oxidation and efficiently reduced
363 to Met barely sensitive to NaOCl treatment and not reduced by RsMsrP (Supplementary Table 2). Moreover,
364 this diversity could be visible within a single protein, in which all Met may not be uniformly oxidized and
365 reduced. For instance, the ABC transporter DdpA, along with another putative ABC transporter (Figure
366 3B,C), contained one of the two only MetO found to be fully reduced in the dataset (Met-230 and Met-353,
367 respectively), although DdpA also contained the Met-243 that was neither efficiently oxidized nor reduced.
368 This is also illustrated by the case of the peptidyl-prolyl cis-trans isomerase, which possessed the Met found
369 to have the higher decrease in oxidation in the entire dataset (Met-172) but also a Met almost not reduced
370 by RsMsrP (Met-190) (Figure 3D). The Met-539 of the PQQ dehydrogenase XoxF illustrates the case in
371 which a Met was highly sensitive to NaOCl-oxidation and very efficiently reduced (Figure 3E). Twenty-one
372 Met were oxidized at 50 % or more and reduced by 50 % or more by RsMsrP (Supplementary Table S2).

373 Altogether, these results show that RsMsrP can reduce a broad spectrum of apparently unrelated proteins
374 (only 11 Met among 202 were not reduced). However, since all MetO were not reduced with similar
375 efficiency, some structural or sequence determinants could drive the ability of MetO to be reduced by
376 RsMsrP.

377 **The nature of the amino acids surrounding a MetO influences the RsMsrP efficiency**

378 Having in hand a relatively large dataset of oxidized and reduced Met prompted us to search for
379 consensus sequences that could favor or impair the oxidation of a Met or the reduction of a MetO by RsMsrP.
380 For all identified Met, we extracted, the surrounding 5 amino acids on the N- and C-terminal sides to obtain
381 an 11-amino acid sequence with the considered Met centered at the 6th position. As shown for bacterial
382 MsrB [39], this length might be sufficient to encompass the amino acids in physical contact with RsMsrP
383 during reduction. We then performed an IceLogo analysis aiming to identify whether some residues were
384 enriched or depleted around the target Met. The principle is to compare a ‘positive’ dataset of peptides, to a
385 ‘negative’ one [38]. To find potential consensus sequences of oxidation, we first compared all unique
386 MetO-containing peptides from both the untreated and the NaOCl-oxidized periplasmic extracts (our
387 positive dataset) to the theoretical *R. sphaeroides* proteome (our negative dataset). The IceLogo presented
388 in Figure 4A shows that MetO-containing sequences were mainly depleted of His and aromatic or
389 hydrophobic residues (Trp, Phe, Tyr, Leu, Ile) and were mainly enriched in polar or charged amino acids
390 (Asn, Gln, Asp, Glu and Lys). This suggests that Met in a polar environment, as commonly found at the
391 surface of proteins, are very likely more susceptible to oxidation than those located in hydrophobic
392 environments such as those in the protein core. We then compared all these unique MetO-containing
393 peptides to all the Met-containing peptides from the same samples (Figure 4B), and we observed that Trp,
394 along with His, Tyr and Cys, were principally depleted around the potentially oxidized Met. Strikingly, the
395 only amino acid significantly more abundant around an oxidized Met was another Met in position -2 and
396 +2. These results indicate that oxidation sensitive Met might be found as clusters.

397 To identify potential consensus sequence favorable to MetO reduction by RsMsP, we performed a
398 precise comparison of the oxidation percentage before and after the action of the enzyme. We thus defined
399 two criteria to characterize the reduction state of each Met: i) the percentage of reduction calculated using
400 the formula described in Supplementary Table S2 and based on the comparison of the oxidation percentages
401 in oxidized versus repaired oxidized periplasm. For instance, a Met found oxidized at 25 % in the oxidized
402 periplasm and at 5 % in the repaired oxidized periplasm was considered enzymatically reduced at 80 %. ii)
403 the decrease in percentage of oxidation by comparison of the 2 samples. For instance, the same Met found
404 oxidized at 25 % in the oxidized periplasm and 5 % in the repaired extract had a decrease in the oxidation
405 percentage of 20 %. This second criterion was used to avoid bias in which very little oxidized Met were
406 considered as efficient substrates (*i.e.* a Met oxidized at 5 % in the oxidized periplasm extract and at 1 % in
407 the repaired oxidized periplasm was reduced at 80 %, similarly to one passing from 100 % to 20 %, which
408 intuitively appears as a better substrate than the previous one). We selected as efficiently and inefficiently
409 reduced MetO those for which both criteria were higher than 50 % and lower than 10 %, respectively.
410 Comparison of the sequences surrounding the efficiently reduced MetO to the theoretical proteome of *R.*
411 *sphaeroides* showed no depletion of amino acid, but mainly enrichment of polar amino acids (Gln, Lys, and
412 Glu) around the oxidized Met (Figure 5A). Similar analysis with the inefficiently reduced MetO indicated
413 the enrichment of Thr and Ser in the far N-terminal positions (-5 and -4) and of a Tyr in position -2 (Figure
414 5B). The C-terminal positions (+ 1 to +5) were mainly enriched in charged amino acids (Gln, Lys, and Glu),
415 similarly to efficiently reduced MetO. This apparent contradiction may indicate that the amino acids in the
416 C-terminal position of the considered MetO did not really influence the efficiency of RsMsP but were
417 observed simply because of the inherent composition of the overall identified peptides. We then compared
418 the variation of amino acids composition of the MetO-containing peptides between both datasets, using the
419 inefficiently reduced MetO as a negative dataset (Figure 5C). The results were similar to those obtained by
420 comparison with the entire theoretical proteome of the bacterium, *i.e.* most enriched amino acids were polar
421 (Glu, Gln, Asp and Lys) at most extreme positions (-5, -4 and + 2 to + 5). Of note, the conserved presence
422 of a Gly in position -1, and the presence of several other Met around the central Met. This potential

423 enrichment of Met around an oxidation site is consistent with the result found for the sensibility of oxidation
424 (Figure 4B), and indicates that potential clusters of MetO could be preferred substrates for RsMsrP. We
425 found 16 peptides containing 2 or 3 MetO, reduced at more than 25 % by RsMsrP (Supplementary Table
426 S2). This was illustrated, for example, by the cell division coordinator CpoB which possesses two close Met
427 residues (66 and 69) highly reduced by the RsMsrP, or by the uncharacterized protein (YP_353998.1) having
428 4 clusters of MetO reduced by the RsMsrP (Supplementary Table S2).

429 From this analysis, the only depleted amino acids appeared to be Thr and Pro in positions -4 and -3
430 (Figure 4C). To validate these results, we designed two peptides, QWGAGM(O)QAEED and
431 TTPGYM(O)EEWNK, as representative of most efficiently and most inefficiently RsMsrP-reduced
432 peptide-containing MetO, respectively. We used them as substrates to determine reduction kinetics
433 parameters for RsMsrP (Table 1; Supplementary Figure S6). The results showed that the peptide
434 QWGAGM(O)QAEED was efficiently reduced, with the highest k_{cat} value from all the substrates we tested
435 ($\sim 480 \text{ s}^{-1}$) and a K_m of $\sim 4,500 \mu\text{M}$. This yielded a k_{cat}/K_m of $\sim 100,000 \text{ M}^{-1} \cdot \text{s}^{-1}$, which is 2 orders of magnitude
436 higher than the one determined for the free MetO, and 10-fold lower than for the oxidized β -casein (Table
437 1). On the contrary, the peptide TTPGYM(O)EEWNK was not efficiently reduced by RsMsrP (Table 1;
438 Supplementary Figure S6). Indeed, we could not determine the kinetic parameters as the activity value curve
439 never reached an inflection point using concentrations as high as $5,000 \mu\text{M}$. The maximal k_{cat} value was
440 determined at $\sim 70 \text{ s}^{-1}$ at $5,000 \mu\text{M}$ of peptide, which is ~ 3.5 -fold less than the one determined with the
441 same concentration of the other peptide ($\sim 250 \text{ s}^{-1}$) (Supplementary Figure S6). These results are in full
442 agreement with the proteomics analysis and confirm that the nature of the amino acids surrounding a MetO
443 in a peptide or a protein strongly influences its ability to be reduced by RsMsrP.

444 **The RsMsrP preferentially reduces unfolded oxidized proteins**

445 To test whether structural determinants affect RsMsrP efficiency of MetO reduction, we compared its
446 activity using oxidized model proteins, either properly folded or unfolded. We started with chicken
447 lysozyme as it is a very well folded protein highly stabilized by four disulfide bonds [40]. We oxidized it

448 with H₂O₂ and checked its oxidation state by mass spectrometry (Supplementary Figure S7). Surprisingly,
449 using a protocol similar to the one allowing the complete oxidation of the 6 Met of β -casein, we observed
450 only a weak and incomplete oxidation of the protein. The major peak corresponded to the non-oxidized form
451 and a small fraction had an increase in mass of 16 Da, likely corresponding to the oxidation of one Met.
452 Nevertheless, we prepared from this oxidized sample, an unfolded oxidized lysozyme by reduction with
453 dithiothreitol in 4M urea followed by iodoacetamide alkylation of cysteines, and both samples (oxidized
454 and unfolded oxidized), were used as substrates for RsMsRP (Figure 6). We also used
455 glutathione-*S*-transferase (GST) which possesses 9 Met and is highly structured. After oxidation with H₂O₂,
456 GST was incubated with 4 M of the chaotropic agent urea, a concentration sufficient to induce complete
457 unfolding of the protein [15]. For both oxidized proteins, we observed a dramatic increase in activity after
458 unfolding. Indeed, the RsMsRP activity increased 7-fold with the unfolded oxidized lysozyme compared to
459 the folded one, and 6-fold in the case of the unfolded oxidized GST compared to the folded oxidized GST
460 (Figure 6). As the unfolded oxidized protein solutions of lysozyme or GST contained a substantial amount
461 of urea, we performed controls in which the urea was added extemporaneously in the cuvette during the
462 measurements, showing that urea did not influence the RsMsRP activity (Supplementary Figure S8).

463 Mass spectrometry analysis showed that the RsMsRP was able to completely reduce the oxidized
464 lysozyme in these conditions (Supplementary Figure S7), suggesting that observed differences of repair
465 between the folded- and unfolded-oxidized lysozyme were not due to the incapacity of RsMsRP to reduce
466 some MetO, but were due to kinetic parameters. We thus determined the kinetics of reduction of these
467 proteins by RsMsRP (Table 1, Supplementary Figure S8). For the oxidized lysozyme, the k_{cat} and the K_{m}
468 were $\sim 4 \text{ s}^{-1}$ and $\sim 900 \mu\text{M}$, respectively. Using the unfolded oxidized lysozyme, the k_{cat} increased to $\sim 7 \text{ s}^{-1}$
469 and the K_{m} decreased to $\sim 100 \mu\text{M}$. The catalytic efficiency determined with the unfolded oxidized lysozyme
470 was thus ~ 18 -fold higher than the one determined using the oxidized lysozyme before unfolding ($70,200$
471 vs. $4,000 \text{ M}^{-1} \cdot \text{s}^{-1}$). Similar results were obtained with GST. Indeed, with the oxidized GST, we recorded k_{cat}
472 and K_{m} values of $\sim 8 \text{ s}^{-1}$ and $\sim 640 \mu\text{M}$, respectively. Whereas for the unfolded oxidized GST, the k_{cat} was
473 slightly higher ($\sim 12 \text{ s}^{-1}$), and the K_{m} was ~ 6 -fold lower ($\sim 100 \mu\text{M}$). The catalytic efficiency was 10-fold

474 higher for the unfolded oxidized GST than for its folded counterpart (Table 1; Supplementary Figure S8).
475 Altogether, these results showed that RsMsrP is more efficient at reducing MetO in unfolded than in folded
476 oxidized proteins. Moreover, as evidenced with lysozyme that contained only one MetO in our conditions,
477 the increase in activity using unfolded substrate is not dependent on the number of MetO reduced.

478 Discussion

479 All organisms have to face harmful protein oxidation and almost all possess canonical Msrs that protect
480 proteins by reducing MetO. Bacteria also have molybdoenzymes able to reduce MetO, as a free amino acid
481 for the DMSO reductase [26] or the biotin sulfoxide reductase BisC/Z [24,25], but also included in proteins
482 in the case of MsrP [28,29]. Genetic studies and the conservation of MsrP in most gram-negative bacteria
483 indicate that it is very likely a key player in the protection of periplasmic proteins against oxidative stress
484 [28,29] However, an in-depth characterization of its protein substrate specificity is still lacking. In this work,
485 we chose the MsrP from the photosynthetic purple bacteria *R. sphaeroides* as a model enzyme to uncover
486 such specificity. Using purified oxidized proteins and peptides, we showed that RsMsrP is a very efficient
487 protein-containing MetO reductase, with apparent affinities (K_m) for oxidized proteins 10 to 100-fold lower
488 than for the tripeptide Ser-MetO-Ser or the free MetO (Table 1). As reported for canonical MsrA and MsrB
489 [15], we observed important variations in the reduction k_{cat} of different oxidized proteins, arguing for the
490 existence of sequence and structural determinants affecting the enzyme efficiency (Table 1).

491 To find potential physiological substrates of RsMsrP and uncover their properties, we used a proteomic
492 approach aiming at comparing the oxidation state of periplasmic proteins after treatment with the strong
493 oxidant NaOCl, followed by RsMsrP reduction of these proteins. We found 202 unique Met, belonging to
494 70 proteins, for which the sensitivity of oxidation and the ability to serve as an RsMsrP substrate varied
495 greatly (Figure 3, Supplementary Table S2). MetO efficiently reduced by RsMsrP belong to structurally and
496 functionally unrelated proteins, indicating that RsMsrP very likely does not possess specific substrates and
497 acts as a global protector of protein integrity in the periplasm. Interestingly, we observed from our IceLogo
498 analysis that Met sensitive to oxidation are generally presented in a polar amino acid environment and can
499 be found in clusters (Figure 4). These properties might be common to all Met in proteins as similar results
500 were found in human cells [41,42] and plants [43]. Moreover, oxidized Met efficiently reduced by the
501 RsMsrP were also found clustered in polar environments and our analysis shows that the presence of Thr
502 and Pro in N-terminal side of a MetO strongly decrease RsMsrP efficiency (Table 1, Figure 5 and

503 Supplementary Figure S6). To our knowledge, the presence of a Thr close to a MetO was not previously
504 shown to influence any Msr activity, but the presence of a Pro was shown to decrease or totally inhibit MetO
505 reduction by the human MsrA and MsrB3, depending on its position [41].

506 The presence of oxidation-sensitive Met efficiently reduced by the RsMsrP in clusters on polar parts of
507 proteins should facilitate the oxidation/reduction cycle aiming to scavenge ROS as previously proposed for
508 canonical Msrs [44]. This is also illustrated by the methionine-rich protein MrpX proposed as main substrate
509 of the *A. suillum* MsrP, which is almost only composed of Met, Lys, Glu and Asp [29]. The presence of
510 numerous MetO on a single molecule of protein substrate should increase the RsMsrP efficiency as one
511 molecule of the substrate allows several catalytic cycles, potentially without breaking physical contact
512 between the enzyme and its substrate.

513 Comparison of the RsMsrP activity using folded or unfolded protein substrates (lysozyme and GST)
514 showed that it is far more efficient to reduce unfolded oxidized proteins (Figure 6). Similar results were
515 found for canonical Msrs [15]. In the case of MsrB it was because more MetO were accessible for reduction
516 whereas for MsrA this increase was independent of the number of MetO reduced. Here, the use of lysozyme
517 containing only one MetO (Supplementary Figure S7) undoubtedly showed that the increase in activity is
518 not related to the unmasking of additional MetO upon protein denaturation (Table 1; Figure 6). This could
519 indicate that the RsMsrP has better access to the MetO in the protein or that the MetO is more easily
520 accommodated in the active site of the enzyme because of increased flexibility. This should provide a
521 physiological advantage to the bacteria during oxidative attacks, which could occur during other stresses
522 such as acid or heat, hence promoting simultaneous oxidation and unfolding of proteins. Particularly,
523 hypochlorous acid, which was shown to induce *msrP* expression in *E. coli* [28] and *A. suillum* [29], has
524 strong oxidative and unfolding effect on target proteins [45].

525 Finally, previous work indicated that the *E. coli* MsrP lacks stereospecificity and can reduce both *R*- and
526 *S*-diastereomers of MetO chemically isolated from a racemic mixture of free L-Met-*R,S*-O [28]. This
527 discovery is of fundamental importance as it breaks a paradigm in Met oxidation and reduction knowledge,

528 and very likely for all enzymology as non-stereospecific enzymes were very rarely described. Indeed, to our
529 knowledge, all previously characterized enzymes able to reduce Met sulfoxide or related substrates were
530 shown to be absolutely stereospecific. This was the case for the canonical MsrA and MsrB, which reduce
531 only the *S*-diastereomer and the *R*-diastereomer, respectively [7,9–14], as well as for the free Met-*R*-O
532 reductase [22,23] and for the DMSO reductase [26,27] and BisC/Z molybdoenzymes [24,25]. To evaluate
533 the potential lack of stereospecificity of the RsMsrP, we chose to use a different strategy than the one used
534 for *E. coli* MsrP [28] and prepared oxidized β -casein containing only one or the other MetO diastereomer
535 using yeast MsrA and MsrB to eliminate the *S*- and the *R*-diastereomers, respectively. Activity assays and
536 kinetic experiments using a highly purified RsMsrP demonstrated that it can efficiently reduce the β -casein
537 containing only the *R*- or the *S*-diastereomer (Table 1; Figure 2 and Supplementary Figure S4). Moreover,
538 this lack of stereospecificity was undoubtedly confirmed by the ability of the RsMsrP to reduce all 6 MetO
539 formed on the oxidized β -casein (Figure 1). These results, consistent with Gennaris and coworkers finding,
540 indicate that this lack of stereospecificity is very likely common to all MsrP homologs. Together with the
541 apparent ability of the enzyme to repair numerous unrelated oxidized proteins, the capacity to reduce both
542 diastereomers of MetO, argues for a role of MsrP in the general protection of envelope integrity in gram
543 negative bacteria. However, it raises questions regarding the structure of its active site as the enzyme should
544 be able to accommodate both diastereomers. From this, we wondered whether the RsMsrP could reduce the
545 Met sulfone, which can be imagined as a form of oxidized Met containing both *R*- and *S*-diastereomers, but
546 we did not detect any activity (Supplementary Figure S9). Although it could be because of an incompatibility
547 in redox potential, it may indicate that this form of oxidized Met cannot reach the catalytic atom. The
548 three-dimensional structure of the oxidized form of *E. coli* MsrP indicated that the molybdenum atom, which
549 is supposed to be the catalytic center of the enzyme, is buried 16 Å from the surface of the protein [46]. The
550 next challenge will be to understand the MsrP reaction mechanism and will require the determination of the
551 enzyme structure in its oxidized and reduced forms bound to its MetO-containing substrates.

552

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559
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565
566 **Author contribution:** LT, PA, DP and MS designed the study. LT, SG, MIS and MS purified RsMsrP. LT
567 and MS prepared all other proteins. LT, SG, MIS, MS performed biochemical characterization of RsMsrP.
568 LT, MS and DL performed β -casein and lysozyme mass spectrometry analysis and analyzed the data. SG
569 and MS prepared *R. sphaeroides* 2.4.1 *msrP* mutant and periplasmic proteins samples. BA, GM and JA
570 performed proteomics analysis of periplasmic proteins and LT, MS, GM and JA analyzed the data. LT wrote
571 the manuscript with contribution of MIS, DL, PA, DP, JA and MS. All authors approved the final
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711 **Table**

712 **Table 1.** Kinetics parameters of RsMsrP reductase activity towards DMSO and various MetO-containing
713 substrates.

Substrates	k_{cat} (s^{-1})	K_{m} (μM)	$k_{\text{cat}}/K_{\text{m}}$ ($\text{M}^{-1}.\text{s}^{-1}$)
DMSO ^a	28 ± 1	$61,000 \pm 7,000$	465
Free L-Met- <i>R,S</i> -O	122 ± 20	$115,000 \pm 27,000$	1,000
Ser-MetO-Ser	108 ± 17	$13,000 \pm 3,400$	8,300
QWGAGM(O)QAEED	479 ± 24	$4,530 \pm 370$	105,700
TTPGYM(O)EEWNK	> 70	> 5,000	N.D.
Oxidized β -casein	100 ± 5	93 ± 9	1,075,000
β -casein- <i>R</i> -O	49 ± 3	51 ± 6	950,000
β -casein- <i>S</i> -O	8 ± 1	53 ± 10	142,000
Oxidized lysozyme	4 ± 1	886 ± 349	4,000
Unfolded oxidized lysozyme	7 ± 1	105 ± 17	70,200
Oxidized GST	8 ± 2	643 ± 194	12,400
Unfolded oxidized GST	12 ± 3	99 ± 33	120,000

714 ^a From [32]. *N.D.*, not determined.

715

716 **Figures legends**

717 **Figure 1. Mass spectrometry spectrum of β -casein non-oxidized (A), oxidized with H_2O_2 (B) and**

718 **repaired by RsMsrP (C).** A) Commercial β -casein exists as a mixture of genetic variants (7 in our batch).

719 β -casein was analyzed by ESI-MS. Main peaks masses: 1, 23982.7 Da; 2, 24021.6 Da; 3, 24035.9 Da; 4,

720 24075.0 Da; 5, 24089.1 Da; 6, 24127.5 Da; 7, 24142.5 Da. B) β -casein was oxidized with 50 mM H_2O_2

721 before MS analysis. All major peaks underwent an increase of ~ 96 Da compared to the non-oxidized

722 sample. Main peaks masses: 1, 24079.1 Da; 2, 24118.5 Da; 3, 24131.8 Da; 4, 24172.2 Da; 5, 24184.3 Da;

723 6, 24224.4 Da; 7, 24238.2 Da. C) Oxidized β -casein was incubated with RsMsrP (25 nM) in the presence

724 of BV (0.8 mM) and sodium dithionite (2 mM) as electron donors. All major peaks had masses

725 corresponding of the non-oxidized β -casein, showing the ability to reduce all MetO in this protein. Note the

726 presence of a peak with an increase of 16 Da (*, mass of 23999.4 Da) compared to the main reduced peak,

727 indicating an incomplete reduction of the total protein pool. Main peaks masses: 1, 23983.0 Da; 2, 24022.4

728 Da; 3, 24037.1 Da; 4, 24075.0 Da; 5, 24091.0 Da; 6, 24128.2 Da; 7, 24143.7 Da.

729 **Figure 2. RsMsrP activity using oxidized β -casein, β -casein-*R*-O and β -casein-*S*-O as substrates.** The

730 oxidized β -casein (100 μ M) containing both diastereomers of MetO, only the *R* one (" *β -casein-*R*-O*"), or

731 only the *S* one (" *β -casein-*S*-O*") were assayed as substrates of RsMsrP. The RsMsrP activity was determined

732 using benzyl viologen (BV) (0.8 mM) as an electron provider in a glove box under nitrogen. BV was initially

733 reduced with sodium dithionite (2 mM) and oxidation was followed at 600 nm after addition of the enzyme

734 (30 nM). The reaction was done in 50 mM MES, pH6.0. The activity values presented as k_{cat} (s^{-1}) represent

735 the number of mole of MetO reduced per mole of enzyme per second as 2 moles of BV is oxidized per mole

736 of MetO reduced. Data presented are averages of 3 replicates. \pm S.D.

737 **Figure 3. Characteristics of MetO reduction sites and oxidation state of Met in representative**

738 **proteins.** A) Repartition of the number of MetO per percentage of reduction by RsMsrP. B) Percentage of

739 oxidation of Met 353 of putative ABC transporter from HAAT family in the 3 analyzed samples. C)

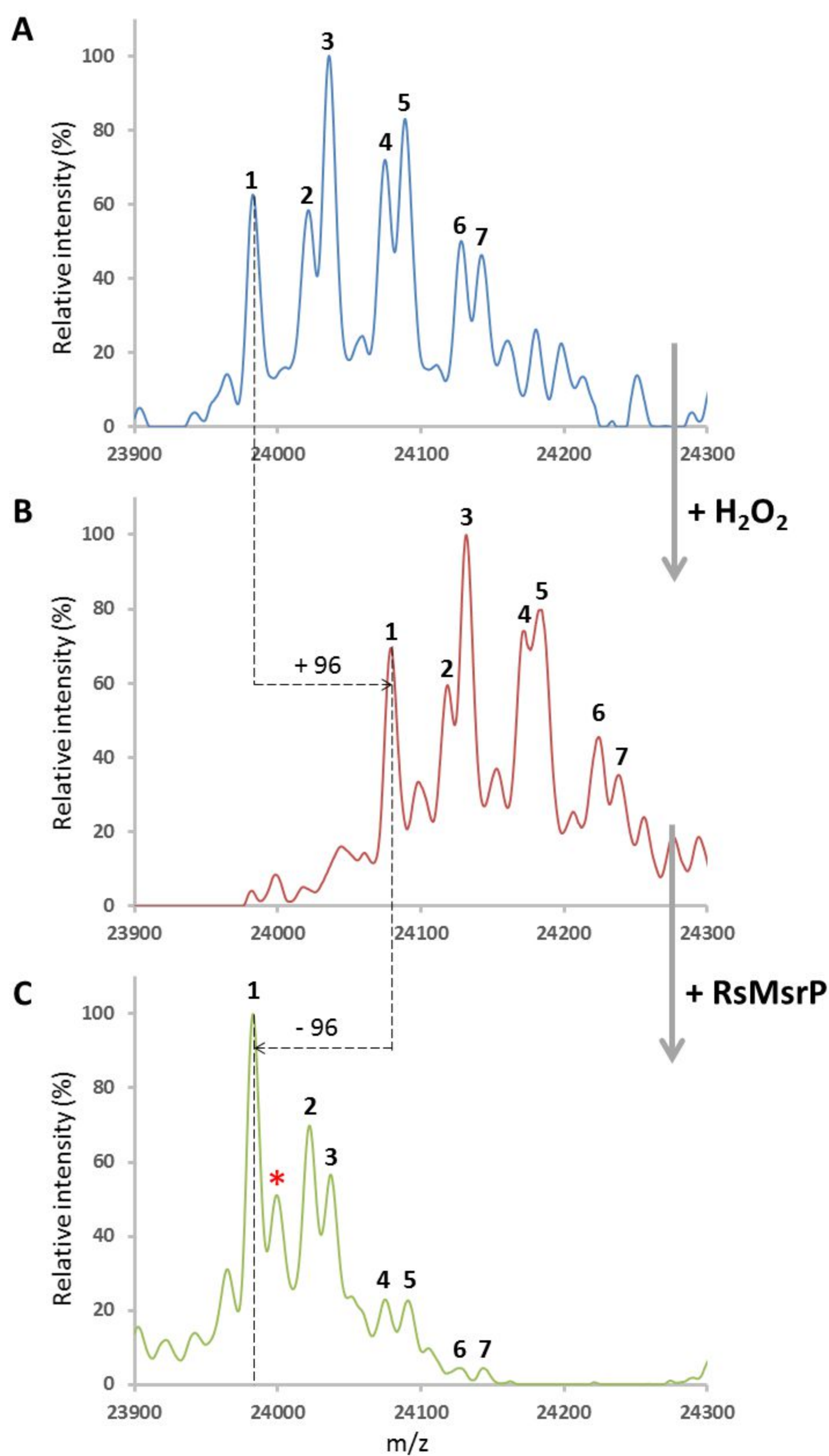
740 Percentage of oxidation of Met 230 and 243 of ABC transporter DdpA. D) Percentage of oxidation of Met

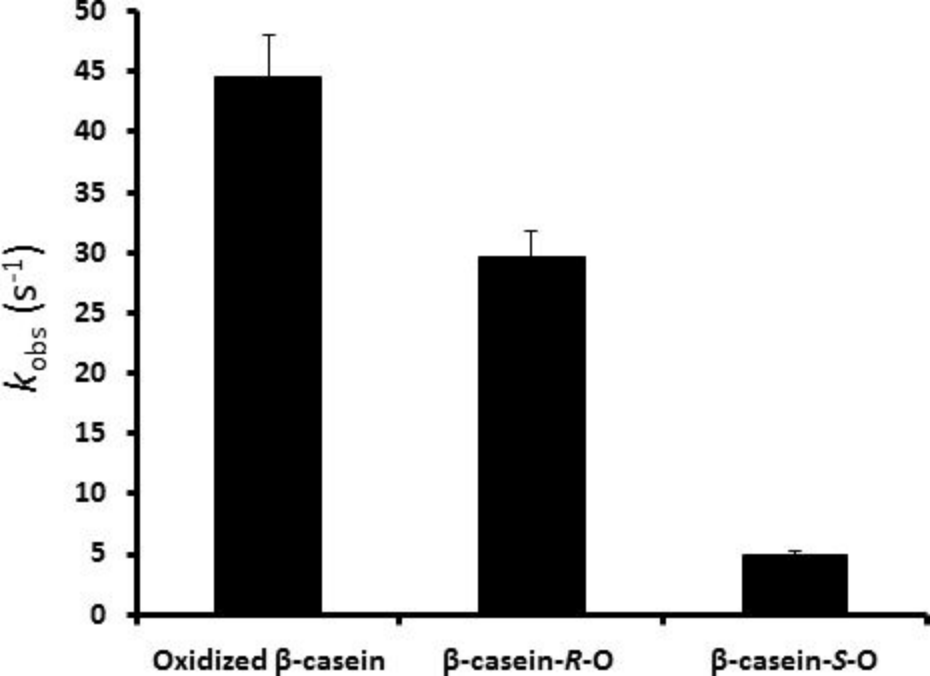
741 92, 172 and 190 in the peptidyl-prolyl cis-trans isomerase. E) Percentage of oxidation of Met 123, 438 and
742 539 of the pyrroloquinoline quinone (PQQ) dehydrogenase XoxF.

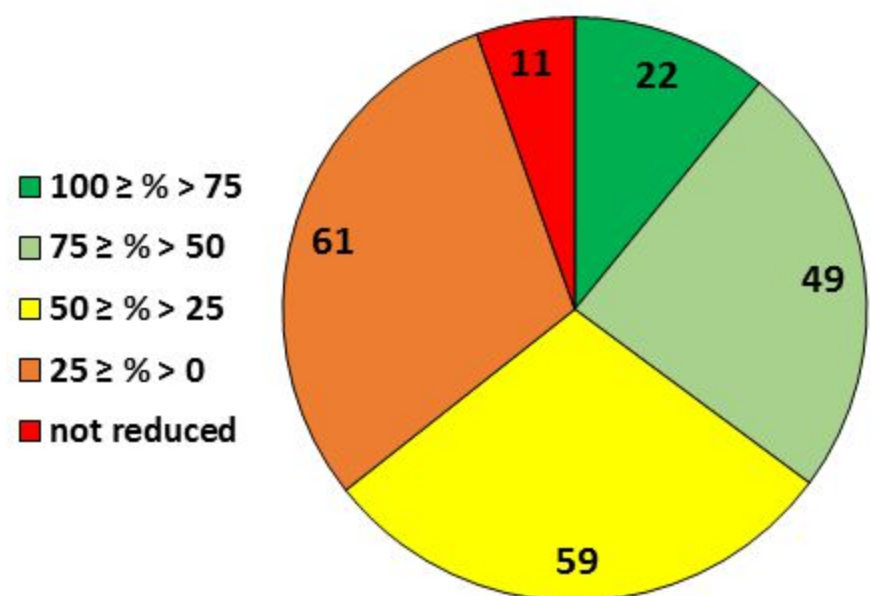
743 **Figure 4. IceLogo representation of enriched and depleted amino acids around the sites of Met**
744 **oxidation.** A) Enrichment and depletion of amino acids around the oxidized Met (M) found in periplasmic
745 extracts and oxidized periplasmic extracts by comparison with the theoretical proteome of *R. sphaeroides*.
746 B) The same oxidized peptides were analyzed using the peptides containing a non-oxidized Met from the
747 same samples (periplasm and oxidized periplasm extracts). Amino acids are colored according to their
748 physiochemical properties.

749 **Figure 5. IceLogo representation of enriched and depleted amino acids around the sites of MetO**
750 **reduction by RsMsrP.** A) Enrichment of amino acids in peptides centered on the MetO for which the
751 percentage of reduction and the decrease in the percentage of oxidation were both superior to 50 % by
752 comparison with the theoretical proteome of *R. sphaeroides*. B) Enrichment of amino acids from peptides
753 centered on the MetO for which the percentage of reduction and the decrease in percentage were inferior to
754 10 % by comparison with the theoretical proteome of *R. sphaeroides*. C) Enrichment and depletion of amino
755 acids from efficiently reduced MetO-containing peptides (dataset used in A)) by comparison with
756 inefficiently reduced MetO-containing peptides (dataset used in B)). Amino acids are colored according to
757 their physiochemical properties.

758 **Figure 6. Relative RsMsrP activity using unfolded oxidized proteins.** The RsMsrP activity was
759 determined as described in Figure 2. Oxidized and unfolded oxidized lysozyme were incubated at 100 μM
760 in 50 mM MES, pH 6.0. Initial turnover numbers were $0.65 \pm 0.12 \text{ s}^{-1}$ and $7.38 \pm 0.23 \text{ s}^{-1}$ with oxidized and
761 unfolded oxidized lysozyme, respectively. Activity with oxidized and unfolded oxidized GST (75 μM) was
762 determined similarly except that reaction buffer was 30 mM Tris-HCl, pH 8.0 because unfolded oxidized
763 GST precipitated in 50 mM MES, pH 6.0. Initial turnover numbers were $0.86 \pm 0.08 \text{ s}^{-1}$ and $5.31 \pm 0.39 \text{ s}^{-1}$
764 with oxidized and unfolded oxidized GST, respectively. Data presented are averages of three replicates. \pm
765 S.D.

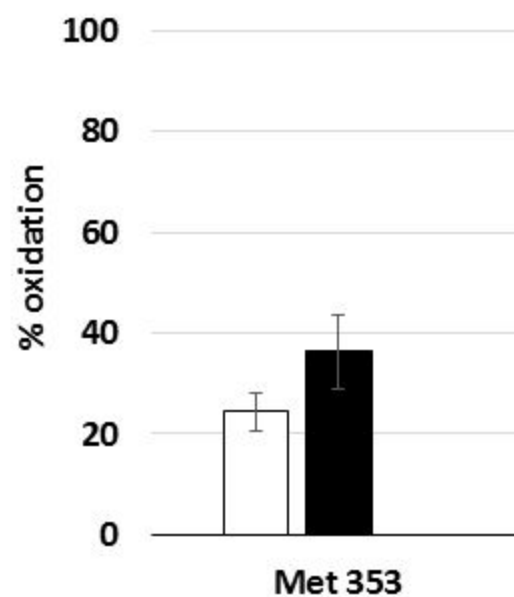




A**B**

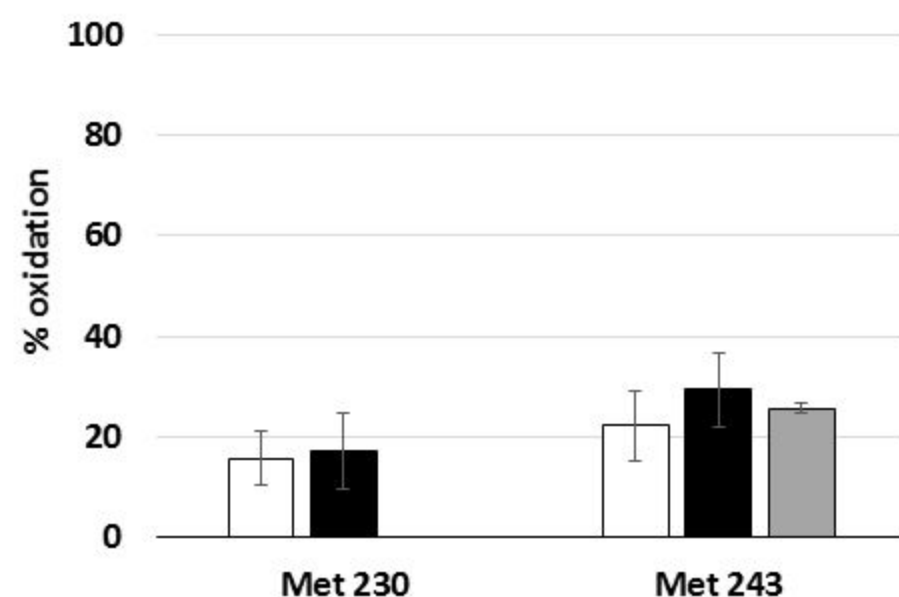
ABC trans. HAAT-fam.

□ Peri. ■ Ox. Peri. ■ Rep. ox. peri.

**C**

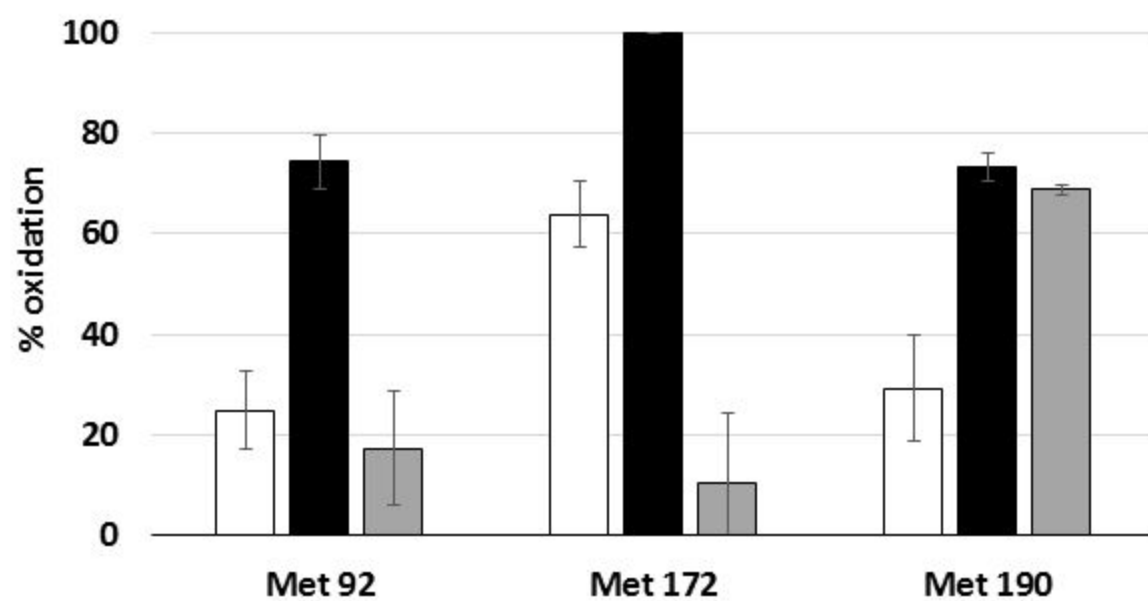
ABC transporter DdpA

□ Peri. ■ Ox. Peri. ■ Rep. ox. peri.

**D**

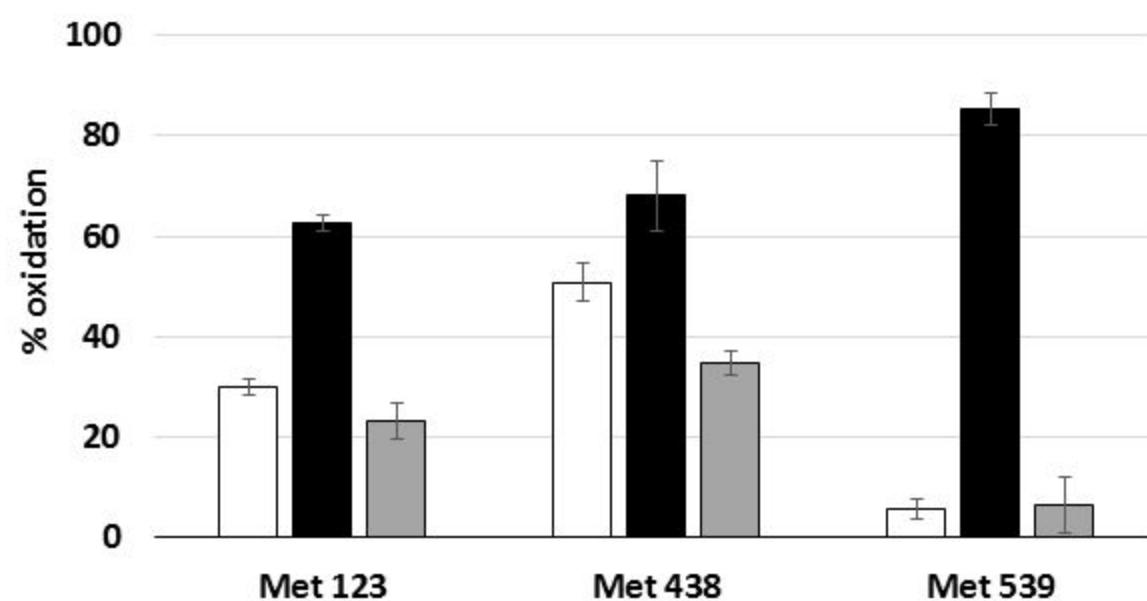
Peptidyl-prolyl cis-trans isomerase

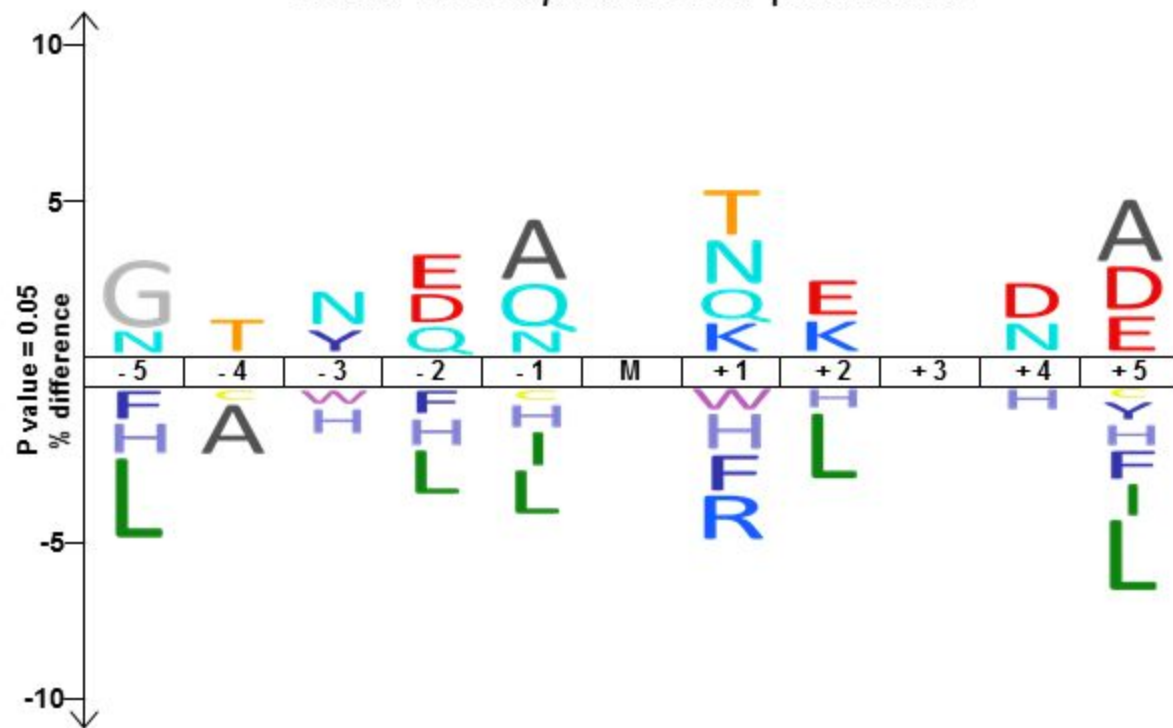
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**E**

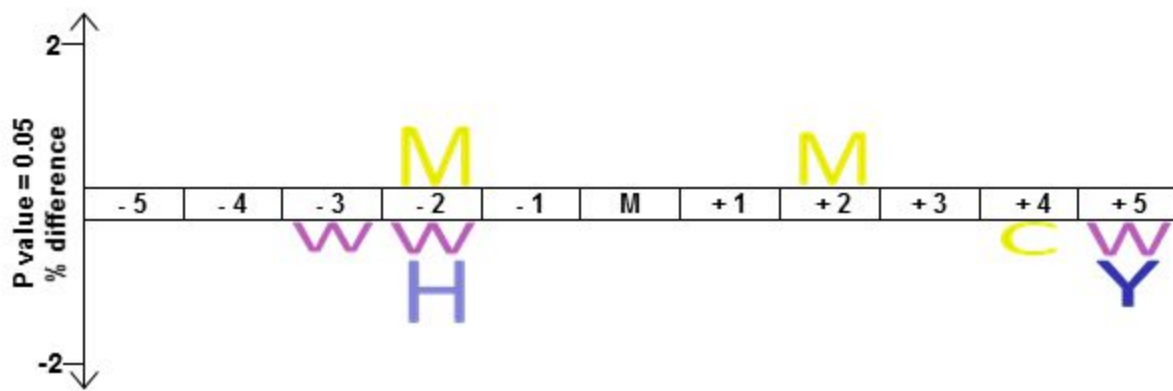
PQQ dehydrogenase XoxF

□ Peri. ■ Ox. Peri. ■ Rep. ox. peri.



AMetO vs. *R. sphaeroides* proteome**B**

MetO vs. Met



Relative RsMSRP activity

