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Experimental and Theoretical Study of the Complexation of Cesium and Thallium Cations by a Water-soluble Cryptophane

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Abstract: Cs (cesium) and TI (thallium) are known to be very toxic for the environment and human health. Thus, the synthesis of molecular receptors aimed at extracting these two elements from the environment is strongly desired. In this Article, we report the synthesis of the two enantiomers of cryptophane- $223(OH)_7$ (1) and the study of their interaction with cesium and thallium cations in basic aqueous solutions. These two complexes have been studied by $^{133}\mathrm{Cs}$ and $^{205}\mathrm{TI}$ NMR spectroscopy to reveal the complexation of the two metallic cations and by chiroptical techniques (electronic and vibrational circular dichroism) to provide valuable information about the conformational changes occurring during the binding process. The thermodynamic parameters of complexation K, ΔH^0 and ΔS^0 obtained from titration experiments reveal a strong interaction between 1 and the two cations under a large range of experimental conditions. A decomposition of the total binding energy, performed by DFT calculations, allows us to characterize the nature of the interactions existing between the cage-molecule and these two cations. These calculations also reveal the importance of the spin-orbit coupling for predicting correctly the large frequency difference between the free TI⁺ and TI⁺@1 NMR signals and to understand its origin. In addition to the development of a methodology enabling detailed understanding of the host-guest interaction, this study indicates a very pronounced selectivity of this cage-molecule towards both Cs⁺ and Tl⁺ cations in various experimental conditions.

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Supporting information for this article is given at the end of the document.

Introduction Since the previous century, extraction of toxic ions from aqueous solutions by dedicated host molecules is a subject of intense research. During the last decade, we have reported that water-soluble cryptophanes possess remarkable binding properties for a large variety of guest molecules (halogenomethanes, oxiranes and cationic species).^[1] For instance, the efficient binding of cesium (Cs^+) and thallium (TI^+) cations with water-soluble cryptophanes bearing phenol groups has been evidenced in LiOH/H2O, NaOH/H2O and even in KOH/H₂O solutions. Thus, it has been shown that cryptophanes 2, 3, 4 and 5 (Scheme 1) are among the best-known molecular receptors for encapsulating cesium and thallium cations in aqueous solutions. The affinity of Cs^+ and TI^+ for these host molecules depends on external parameters (nature of the basic solution, concentration) but also the nature of the cryptophane structure. For instance, it has been clearly established that the presence of phenol groups and their number play a key role in the stabilization of these complexes. This suggests that coulombic interactions probably play an important role in their stabilization. Other types of interactions such as π -cation interaction probably exist but these different contributions are difficult to quantify without the use of quantum chemistry calculations.



Scheme 1. Chemical structures of water-soluble cryptophanes(OH)_x (x = 5, 6) **2-5** used previously for Cs⁺ and Tl⁺ encapsulation.

It has been also reported that cryptophane with larger inner cavities can bind efficiently Cs⁺ and Tl⁺ cations. For instance, compound **4**, which possesses two ethylenedioxy and one propylenedioxy bridges shows a strong affinity for these two cations. Interestingly, the structure of this cryptophane can be modified to introduce a reactive function on the propylenedioxy linker.^[2] This chemical group can be chosen in order to react orthogonally with the other groups grafted on the benzene rings.

Cryptophane 1 reported in this article (Scheme 2) belongs to this class of compounds. It contains a secondary alcohol function attached on the propylenedioxy linker and the synthesis of 1 in its racemic form has been recently described.^[2] Herein, we report the synthesis of the two enantiomers *MM*-1 and *PP*-1 and their characterization by ¹H, ¹³C NMR and high-resolution mass spectrometry. We show that compound 1 binds very efficiently Cs⁺ and Tl⁺ cations in LiOH/H₂O, NaOH/H₂O and KOH/H₂O. Electronic (ECD) and vibrational (VCD) circular dichroism techniques are used to investigate the conformational changes occurring during the binding process. Isothermal titration experiments (ITC) enable accurate determination of the parameters of complexation (*K*, ΔH^{0} , ΔS^{0}). In addition, ¹³³Cs and ²⁰⁵TI NMR spectroscopy give valuable information on the chemical environment of these two cations encapsulated into the cavity of the cage. Finally, we use DFT calculations and decomposition of the total binding energy to investigate and quantify the nature of the interactions that exist between these two cations and host **1**.



Scheme 2. Structure of the water-soluble cryptophane MM-1.

Results

Preparation of enantiopure cryptophane 1. Optically active cryptophane 1 has been prepared in a three-steps synthesis from cryptophane 6 whose synthesis has been reported recently in its racemic form.^[2] The two enantiomers of ${\bf 6}$ have been separated by semi-preparative chiral HPLC chromatography to give rise to enantiomers (-)-6 and (+)-6. The enantiomeric excess of (-)-6 and (+)-6 (ee > 99.5 %) has been checked by analytical HPLC chromatography (Figure S1 in the Supporting Information). Additional purification steps on silica gel and recrystallization in CH₂Cl₂/EtOH provide these two enantiomers with high chemical purity. The ECD spectra recorded in CH₂Cl₂ and CHCl₃ and polarimetric measurements (in CHCl₃) of the two enantiomers (-)-6 and (+)-6 are reported (Figure S2 in the Supporting Information). According to the ECD spectra and polarimetric values obtained for compounds 2-5, we found the following assignment (-)-MM-6 and (+)-PP-6 for the two enantiomers of 6.

Lithium diphenylphosphide (1 M in THF) has proved to be a very efficient reagent to deprotect in a single step both the six methyl groups and the allyl function attached on the propylenedioxy linker.^[2,3,4] Applied to compounds *MM*-6 and *PP*-6, this approach gives rise the two desired MM-1 and PP-1 derivatives in 97 and 99% yields (crude product), respectively. Since the two isolated compounds MM-1 and PP-1 contain impurities in small amount, an additional purification has been used. Thus, these two compounds have been allowed to react with anhydride acetic in pyridine to give cryptophanes MM-7 and PP-7 in 99 % and 78 % after purification, respectively. Then, MM-1 and PP-1 derivatives have been recovered in 98% and 92% yields, respectively by hydrolysis of the ester group under basic conditions (Scheme 3). The ^1H and ^{13}C NMR spectra of compounds MM-1 and PP-1 are identical to those previously reported for the racemic derivative (Figures S3-S6 in the Supporting Information). ¹H and ¹³C NMR spectra of *MM*-7 and PP-7 are also reported (Figures S7-S11 in the Supporting Information). It is noteworthy that compound 7 may exist under imploded form, which is ascertained by the presence of characteristic signals in the ¹H NMR spectrum.^[5,1c] As an example, the ¹H NMR spectrum of a sample containing mainly the imploded form is given (Figure S8 in the Supporting Information).



Scheme 3. Synthesis of the two enantiomers of cryptophanes 1 and 7 obtained from the two enantiomers of cryptophane 6. a) PPh_2Li (1 M), THF, 60°C, 48 hrs; (97 %, *MM*-1; 99 %, *PP*-1) b) Ac₂O, Pyridine, 0°C - 20°C, 5 hrs (99%, *MM*-7, 78%, *PP*-7); c) KOH (0.5 M), THF, 50°C, 16 h, (98 %, *MM*-1; 92%, *PP*-1).

Isothermal Titration Calorimetry (ITC). ITC is the method of choice to measure the association constants between cesium and thallium cations with host 1 in aqueous solutions. In the past, we have used this method to determine the binding constant Kand the thermodynamic parameters of complexation ΔH^0 , ΔS^0 and ΔG^0 of Cs⁺ and Tl⁺ cations with compounds 2-5.^[6] ITC experiments of (*rac*)-1 in the presence of Cs⁺, Tl⁺ and Rb⁺ have been performed in LiOH/H₂O, NaOH/H₂O and KOH/H₂O solutions at different concentrations (0.1 or 1.0 M) (Figures S12-S15 in the Supporting Information). Basic conditions are mandatory to ensure a good solubility of the cage. To interpret the data obtained from isothermal titration calorimetric (ITC) experiments, we will consider that, in our experimental conditions, only the encapsulation of the metallic cations by the molecular host takes place. Example of such an equation reaction is reported in Scheme 4 that describes the encapsulation of different metal cations ($M^+ = Rb^+$, Cs^+ , TI^+) by host **1** in NaOH/H₂O. The binding constants of $Rb^+@(rac)-1$, $Cs^{\dagger}@(rac)-1$ and $Tl^{\dagger}@(rac)-1$ complexes are reported in Table 1 whereas the thermodynamic parameters of complexation ΔH^0 . ΔS^{0} and ΔG^{0} are given in Supporting Information (Figure S16 in the Supporting Information). Even though Cs⁺ and Tl⁺ are the two main cations of interest in this study, the association constant between Rb⁺ and the cage has been measured in some cases to perform competition experiments for high values of K.



Scheme 4. One binding site model considered for the isothermal titration calorimetric experiments (ITC) in NaOH/H $_2$ O.

In LiOH/H₂O solution at 298 K, the ITC experiments show a very strong association between the Cs⁺ and Tl⁺ cations and the cage, even at high concentration. For instance, in LiOH/H₂O (0.1 M), an association constant $K = 3.6 \times 10^8 \text{ M}^{-1} (\Delta G^0 = -11.7 \text{ m}^{-1})$ kcal mol¹) has been measured for the $Cs^+@(rac)-1$ complex. At higher LiOH/H₂O concentration (1 M), this binding constant is decreased by 2 orders of magnitude ($K = 4.5 \times 10^{6} \text{ M}^{-1}$; $\Delta G^{0} = -$ 9.1 kcal mol⁻¹). It is noteworthy that, in the same experimental conditions, the TI⁺ cation shows higher affinity for the cage. For instance, a binding constant $K = 1.0 \times 10^9 \text{ M}^{-1}$ ($\Delta G^0 = -13.6 \text{ kcal}$ mol⁻¹) has been measured in LiOH/H₂O (0.1 M). As observed for the Cs⁺ cation, increasing the LiOH/H₂O concentration results in a decrease of the binding constant ($K = 3.1 \times 10^7 \text{ M}^{-1}$; $\Delta G^0 = -10.2$ kcal mol⁻¹). On the other hand, the $Rb^+@(rac)-1$ complex exhibits much lower association constants under the same experimental conditions. It must be mentioned that the K⁺ cation

shows also a moderate affinity for the cavity of **1**. For instance, the binding constant has been measured to be $K = 1000 \text{ M}^{-1}$ in LiOH/H₂O (0.1 M).

Table 1. Binding constants K (M⁻¹) measured at 298 K by ITC for the M^{*}@(*rac*)-1 complexes in LiOH (0.1 M in H₂O), NaOH (0.1 M in H₂O), KOH (0.1 M in H₂O) and LiOH (1 M in H₂O).

Cations M+	LiOH (0.1 M)	NaOH (0.1 M)	KOH (0.1 M)	LiOH (1 M)
Cs⁺	$3.6{\pm}~0.2{\times}10^{8[a]}$	$7.6\pm0.6{ imes}10^{8[a]}$	$1.0{\pm}0.04{\times}10^7$	$4.5\pm0.2{\times}10^6$
TI⁺	$1.0{\pm}~0.1{\times}10^{9[a]}$	$11.0\pm2{\times}10^{9[a]}$	$5.6\pm0.9{\times}10^7$	$3.1\pm0.2{\times}10^7$
Rb⁺	$1.3\pm0.3{\times}10^5$	$2.6\pm0.1{\times}10^5$	-	$1.8\pm0.2{\times}10^3$

[a] from competition experiments (competitor Rb⁺)

Replacing the LiOH solution by a NaOH solution leads to a slightly higher *K* value. For instance, in a 0.1 M NaOH solution, the Cs⁺@(*rac*)-1 complex shows a binding constant *K* = 7.6×10^8 M⁻¹ (ΔG^0 = -12.1 kcal mol⁻¹) slightly higher than that measured in the same conditions in LiOH solution. The same trend occurs for the Tl⁺@(*rac*)-1 complex. In contrast to what is observed in these solutions, the use of a 0.1 M KOH solution has a strong impact on the equilibrium constants of these two complexes. For instance, binding constants *K* = 1.0×10^7 M⁻¹ (ΔG^0 = - 9.6 kcal mol⁻¹) and *K* = 5.6×10^7 M⁻¹ (ΔG^0 = -10.6 kcal mol⁻¹) have been measured for the Cs⁺@(*rac*)-1 and Tl⁺@(*rac*)-1 complexes, respectively. Combined together these results show that both Cs⁺ and Tl⁺ cations are very well recognized under a large range of experimental conditions. A very high selectivity toward Li⁺, Na⁺ and even K⁺ cations is also observed.

¹³³Cs and ²⁰⁵TI NMR Spectroscopy. As both cesium and thallium nuclei have non-null spin isotopes, NMR spectroscopy can be used to characterize the Cs⁺@(*rac*)-1 and Tl⁺@(*rac*)-1 complexes. Thanks to the high polarizability of their electron clouds and the strong shielding effect induced by the six aromatic rings, high field NMR signals characteristic of the encapsulated form of these cations are observed.

The ¹³³Cs NMR spectrum of Cs⁺ in the presence of (*rac*)-1 shows two well-resolved signals at 298 K in NaOD/D₂O (0.1 M) solution. Cs⁺ free in solution gives an intense signal near 0 ppm whereas the Cs⁺@(*rac*)-1 complex gives rise to a characteristic ¹³³Cs NMR signal located at δ = -258 ppm. This signal is relatively broad at room temperature. Interestingly, a sharpening of the Cs⁺@(*rac*)-1 NMR signal is observed when the temperature of the solution increases (Figure S17 in the Supporting Information). In contrast, lowering the temperature results in a broadening of the Cs⁺@(*rac*)-1 NMR signal. This counter-intuitive effect has been previously observed with cryptophanes **2**, **3** and **4**, which also show a very high affinity for Cs⁺.

The ²⁰⁵TI NMR spectra are more difficult to record since this nucleus resonates at a frequency out of the scope of most of the commercial NMR probe heads. Consequently, a homemade system is necessary to facilitate the detection of this signal (see Experimental Section in the Supporting Information for more details). In addition, the detection of the TI⁺@1 signal is made difficult due to the large difference of chemical shift existing between the TI⁺ cation present within the cavity of the cryptophane host and TI⁺ dissolved in the bulk. In the present case, a high field shifted signal located at -680 ppm has been observed for the TI⁺@(*rac*)-1 complex. Another signal located at +340 ppm has been detected for the free TI⁺ cation. This leads to a chemical shift difference larger than 1000 ppm for this

system. It is noteworthy that in the case of the TI element, a slight modification in the chemical structure of the host can induce large chemical shift differences. This is evidenced in Figure 1 by comparing the ²⁰⁵TI NMR signals of two TI⁺@(*rac*)-1 and TI⁺@(*rac*)-4, which differ only by the presence of a secondary alcohol function attached on propylenedioxy linkers in host 1.



Figure 1.²⁰⁵TI NMR spectrum of the TI⁺@4 and TI⁺@1 complexes in NaOD/D₂O (0.1 M) at 348 K. The free TI⁺ signal is positioned at 340 ppm with respect to TINO₃ (not shown).^[7]

ECD Spectroscopy. Thanks to the preparation of enantiopure compounds *MM*-1 and *PP*-1, the encapsulation process of Cs⁺ and Tl⁺ cations can also be studied using chiroptical techniques such as ECD. This technique is known to be a valuable tool to study the conformational rearrangements that take place in solution upon the encapsulation process. Indeed, large spectral changes are sometimes observed. ECD spectra of the Cs⁺@*MM*-1 and Tl⁺@*MM*-1 complexes in LiOH/H₂O (0.1 M) in the presence of different amounts of Cs⁺ and Tl⁺ cations are presented in Figure 2.

Figure 2 shows that the addition of various quantities of a cesium hydroxide solution (from 0 to 1 equiv.) to the LiOH/H₂O (0.1 M) solution of free host **1** gives rise to a significant change of the overall ECD spectrum in the 220-380 nm region. The presence of three isosbestic points indicates the formation of a new species upon addition of Cs⁺. This new species can be attributed to the formation of the Cs⁺@*MM*-1 complex. Replacing the Cs⁺ cation by Tl⁺ cation leads to more marked modifications of the ECD spectra. These spectral changes mainly occur for the ECD bands of the ¹L_b transition (280 – 320 nm) for which addition of one equivalent of Tl⁺ results in a significant bathochromic shift. As observed with Cs⁺, the ECD spectra also reveal the presence of multiple isosbestic points indicating the formation of the Tl⁺@*MM*-1 complex.



Figure 2. ECD spectra of the Cs⁺@*MM*-1 (top spectra) and Tl⁺@*MM*-1 (bottom spectra) complexes recorded in LiOH/H₂O (0.1 M) at 293 K as a function of the amount of cesium hydroxide or thallium acetate added into the solution (0 eq, 0.25 eq, 0.5 eq, 0.75 eq and 1.0 eq). ECD spectra for higher Cs⁺ or Tl⁺ concentrations are identical to that recorded for 1 equivalent of Cs⁺ or Tl⁺.

A modification of the experimental conditions also produces important changes in the ECD spectra of the MM-1 derivative. For instance, the presence of a competitor (CHCl₃) present inside the cavity of MM-1 significantly affects the overall ECD spectrum of compound MM-1. The replacement of the chloroform molecule by the Cs⁺ or the Tl⁺ cations has a dramatic effect on the global shape of the ECD spectrum of MM-1, especially for the ECD bands of the allowed ¹B_b transition located at low wavelength (close to 220 nm). Indeed, a strong decrease in intensity is observed upon addition of Cs⁺ or Tl⁺ cations into the solution (Figure S18 in the Supporting Information). The presence of multiple isosbestic points indicates that both the Cs⁺@MM-1 and Tl⁺@MM-1 complexes are efficiently formed and that these two cations can successfully expel the CHCl3 molecule from the inner cavity of the host. It is noteworthy that a change of the concentration of the LiOH/H₂O solution produces similar effects (Figure S19 in the Supporting Information). For instance, higher LiOH/H₂O concentration (1 M) solution gives rise to ECD spectra similar to those recorded for the CHCl₃@MM-1 complex. The addition of CsOH/H₂O or TIOAc/H₂O into this solution strongly affects the ECD spectra of MM-1. In all cases, it is important to note that the ECD spectra remain unchanged when more than one equivalent of Cs⁺ or Tl⁺ are added into the solution. Finally, the replacement of LiOH/H₂O by a KOH/H₂O solution reveals two different behaviors in the ECD spectra of MM-1 upon addition of Cs⁺ or TI⁺ cations. In the presence of cesium hydroxide solution, very small modifications of the ECD spectra of MM-1 are observed, whereas the spectral changes are more pronounced when a thallium acetate solution was added (Figure S20 in the Supporting Information).

Vibrational Circular Dichroism Spectroscopy. The VCD spectrum of empty PP-1 is compared in Figure 3 with the VCD spectra of CDCI₃@PP-1 and Cs⁺@PP-1 complexes recorded in NaOD/D₂O (0.21 M) solution. Important spectral modifications occur upon complexation of CDCl₃ and Cs⁺, as already observed for compound 3.^[1f] Indeed, the intensity of the band at 1495 cm⁻¹ increases for the $CDCl_3@PP-1$ complex, whereas this band presents a bisignate pattern for the Cs⁺@PP-1 complex. On the other hand, the intensity of the band at 1600 cm⁻¹ strongly increases for the Cs⁺@PP-1 complex. These spectral changes, as well as those observed in the 1450-1250 cm⁻¹ region, which correspond to coupled modes involving wagging and twisting vibrations of the CH₂ groups (chains and caps), may be interpreted as a modification of the conformation of the linkers of host 1. Thus, the presence of CDCl₃ inside the cavity of 1 favors the trans conformation of the ethylenedioxy linkers whereas the presence of Cs⁺ cations favors the gauche conformations of the linkers in order to maximize the host-guest interactions.



Figure 3. VCD spectra of empty *PP*-1 as well as $CDCl_3@PP$ -1 and $Cs^*@PP$ -1 complexes recorded at 293 K in NaOD/D₂O solution (0.21 M). The concentration of host 1 was 0.030 M.

DFT Calculations. Quantum chemistry calculations have been performed to understand the nature of the various interactions that can exist between the host and the cations. These calculations are also expected to be useful to understand the origin of the large NMR chemical shift observed for the Cs⁺@1 and the TI⁺@1 complexes. Thus, we have carried out orbital, topological and energy decomposition analyses (EDA) along with the calculation of chemical shift for the Cs⁺@1 and the TI⁺@1 complexes with quantum chemistry methods.^[8,9] Computational details are provided in the Supporting Information. The optimized geometry of the complexes is rather similar for alkali metals (K^+ , Rb^+ , Cs^+) and for the heavier TI^+ cation, as shown in Figure 4. However, the position of the TI⁺ cation within the cryptophane cage differs from that of Cs⁺, revealing a modification of the interaction between the TI⁺ cation and the cade.

The nature of the bonding interactions between the metal and the host has been first investigated with an energy partitioning of the total interaction energy into components familiar to chemists, namely electrostatic, Pauli repulsion and orbital mixing.



Figure 4. Optimized geometry of the Cs $^*@1$ (blue) and Tl $^*@1$ (magenta) complexes.

It is important to note that all of the terms assignable to the bond (polarization, charge-transfer, etc.) are grouped into a single orbital interaction term. The results are reported in Figure 5.



Figure 5. DFT PBE/TZ2P bonding energy analysis with respect to the \mbox{M}^{*} and cryptophane fragments.

The $M^*@1$ total bonding energy is a negative value indicating stabilization during encapsulation of the cation, which originates mainly in attractive electrostatic (~ 32% of the total attractive interactions) and orbital interactions (~ 50% of the total attractive interactions). The dispersion forces are a non-negligible component of the interaction (~ 18% of the total attractive interactions). The spatial regions in which the non-covalent interactions (electrostatic, dispersion) are taking place are visualized in Figure 6.

The total bonding energy is much more negative for the TI^{+} complex than for the Cs⁺ complex. The Voronoi Deformation Density (VDD) analysis reveals if charge flows away or toward the space around a certain nucleus upon the formation of the complex from its fragments.^[10] Upon encapsulation of the alkali metals the VDD analysis indicates that no charge transfer occurs between the cage and the cation. In contrast, in the case of the thallium cation, 0.2 electrons (0.25 electrons from natural bond orbital (NBO) analysis) are transferred from the cage to the 6p orbital of the TI⁺ cation.^[11]

The 6d(TI) orbital mixes the 2p(C) orbital of the phenyl ring (symmetrized fragment orbital analysis).^[9] The calculation reveals that the TI⁺**@1** complex is also stabilized at lower energy (See Figure 1a in the computational details in the Supporting Information) by the mixing of the 6s(TI) orbital with the 2p(C) orbital of the aromatic ring. In the alkali metal complexes, the 5p(M) orbital mixes with the 2p(C) orbital of all cages (See Figure 1b in the computational details in the Supporting Information).



Figure 6. Spatial regions of non-covalent interaction. The non-covalent interaction regions appear in green and light brown for Cs^+ (a) and Tl^+ (b). Hydrogen atoms are removed for clarity. The cations are in blue color.

From this analysis, it is clear that the phenyl ring-Tl⁺ interaction is predominant inside the cryptophane cage. The Tl⁺@1 complex is stabilized by orbital interactions and by cation- π interactions. In the alkali metal series, cation- π interactions cannot be clearly identified.



Figure 7. Calculated chemical shifts using a secondary reference (See computational details in the Supporting Information).

These different interaction patterns, as revealed by the ECD spectra, have a consequence on the magnetic properties of the metal ions. The M⁺ isotropic shielding was calculated through the DFT/GIAO shielding tensor. The results are reported in Figure 7 for Cs⁺ and Tl⁺. As shown in Figure 7, it is critical to take into account explicitly all the relativistic contributions (scalar and spin-orbit) to the nuclear shielding for TI element. Upon complexation, the electronic effects (phenyl ring π electrons and cation-cryptophane orbital interaction) increase the electron density at the nucleus. For the TI⁺ complex, this effect is large, shifting the NMR signal to high field with a huge chemical shift variation with respect to that experienced by the alkali metal cations.

Discussion

Comparison of the binding properties of 1 with cryptophane congeners 2 - 5. ITC experiments performed on the $Cs^+@(rac)$ -1 and the $TI^+@(rac)$ -1 complexes reveal a very high affinity of host 1 for the cesium and thallium cations. This cage shows the highest affinity for the TI^+ cation. The affinity of 1 for Cs^+ is about ten times smaller than the affinity measured for TI^+ . In addition, the Rb⁺ and K⁺ cations are much less recognized by the host since the affinity for these two cations is about one thousand and one million times smaller, respectively, than the binding constant measured for the Cs⁺ cation. The same trend was

previously observed with cryptophanes **2** - **5**.^[1e,1f,6] Altogether, our results show that a clear relationship can be established between the magnitude of the binding constant and the polarizability of the M⁺ cation (TI⁺ > Cs⁺ > Rb⁺ > K⁺ >> Na⁺, Li⁺). The affinity of **1** for the Cs⁺ and TI⁺ cations lies in the same

The affinity of **1** for the Cs⁺ and Tl⁺ cations lies in the same range than the binding constant measured for compounds **2** and **4**.^[1e,6] These compounds also possess six phenol groups attached on the benzene rings but their cavity size is different. Compounds **1** and **4** have the closest structures since they differ only by the presence of a secondary alcohol function in **1**. Thus, upon complexation of Cs⁺ or Tl⁺ it is not surprising to observe comparable binding constants between these two compounds. The similarity in magnitude of the binding constants suggests that the alcohol group present in **1** does not play any crucial role in the stabilization of the cavity and cannot establish specific interactions with the metal cation present within the cavity. A minimized structure of the host obtained by DFT supports this assumption.

Comparison of the ECD spectra of 1 with compounds 2 and 3 reveals the same evolution of the ECD spectra in the presence of metal cations. ECD spectroscopy was found to be very sensitive to the conformational changes of water-soluble cryptophanes upon encapsulation of guest molecules or cations.^[1] The Cs⁺ or Tl⁺ encapsulation induces large spectral modifications, which are the consequence of the conformational rearrangements of the linkers. These conformational rearrangements allow the cage to change its cavity size in order to maximize its interactions with the metal cations. It is noteworthy that a higher concentration of Cs⁺ or Tl⁺ cations (more than 1 equivalent) does not induce further spectral changes, suggesting that host-guest interactions are solely responsible of the observed effect. Such spectral changes have already been observed with compounds ${\bf 2}$ and ${\bf 3}^{[1e,1f]}_{,}$ but it is interesting to notice that in the case of 1 these effects are not amplified, even though host 1 is potentially subject to larger conformational changes. Indeed, the propylenedioxy linker, present in the structure of 1, is certainly more favorable to conformational changes than ethylenedioxy linkers (compounds 2 and 3).

The ECD experiments performed in the presence of a competitor (CHCl₃) or in KOH/H₂O solution give new insight to interpret the behavior of 1 in the presence of metal cations. For instance, important similarities are observed between the ECD spectra of the CHCl₃@MM-1 complex (LiOH/H₂O, 0.1 M) and the spectrum recorded at higher LiOH/H₂O (1 M) concentration. This indicates that the cage adopts a similar conformation in solution. The chloroform molecule possesses a large molecular volume (V_{vdw} = 72 Å³) that forces the cage to enlarge its cavity (increasing the proportion of trans conformation of the linkers). Interestingly, increasing the concentration of the basic solution seems to produce a similar effect. Nevertheless, high concentrations of LiOH are not the most favorable conditions to observe efficient complexation of metal cations. However, the interactions between cage 1 and Cs⁺ or Tl⁺ seem to be strong enough to provoke a conformational change that reduces the cavity size (the complexation of Cs⁺ or Tl⁺ cations favors the proportion of gauche conformation). An opposite effect takes place when the experiments are performed in KOH solution. We have reported that K⁺ cations bind moderately to cage **1**. Thus, K⁺ acts as a competitor for Cs⁺ or Tl⁺, resulting in a decrease of the observed binding constants by two orders of magnitude. The encapsulation of the K⁺ cation also results in a pre-organization of the host molecule leading to moderate chiroptical changes upon Cs⁺ and Tl⁺ encapsulation. Indeed, these cations have similar volumes and the replacement of K⁺ (V_{vdw} = 11 Å³) by Cs⁺ $(V_{vdw} = 20 \text{ Å}^3)$ or TI⁺ $(V_{vdw} = 14 \text{ Å}^3)$ is not expected to modify significantly the cavity size of the host.

The ECD experiments also reveal a clear difference between the ECD spectra of the Cs⁺@*MM*-1 and Tl⁺@*MM*-1 complexes. Indeed, a bathochromic shift of the ECD bands of the ¹L_b transition (280 – 320 nm) is observed for the Tl⁺@*MM*-1 complex. This effect has been previously observed for compounds **2-5** in the presence of Tl⁺.^[1f,6] This seems to be an indication that Tl⁺ interact differently with the cages. DFT calculations support this assumption (see below).

Understanding the nature of the interaction between the metallic cations and the cage. For the first time, DFT calculations have been performed comparatively for the Cs⁺@1 and TI⁺@1 complexes. A theoretical approach that consists to decompose the total binding energy into different contributions (electrostatic, Pauli interaction, orbital interaction and dispersion energy) is very useful to understand the nature of the interactions that takes place between the cations and host 1. These DFT calculations confirm that the TI⁺ cation interacts more strongly with the cage than the Cs⁺ cation. In the case of the TI⁺ cation, the calculations reveal that the Pauli repulsion term is strongly positive for the TI⁺ and contributes significantly to the destabilization of the complex. However, this term is largely compensated by the strongly negative electrostatic and orbital interaction terms with a non-negligible charge transfer between TI⁺ and the cage. These terms dominate the host-guest interactions, and the summation over the different contributions leads to a large negative total binding energy ($\Delta E = -70$ kcal/mol), which is characteristic of the formation of a very stable complex. In the case of the Cs⁺ cation, the total binding energy is significantly lower (ΔE = -40 kcal/mol). Indeed, even though the positive contribution of the Pauli repulsion term is much less important than for TI⁺, the two electrostatic and orbital interaction terms are decreased by a factor 2 in magnitude leading to a weaker total binding energy.

These results are consistent with the ECD measurements that suggest a stronger interaction between the cage and the TI⁺ cation (bathochromic effect of the ¹L_b transition). In contrast, the calculations reveal a similar behavior for the interactions of the Cs⁺ and Rb⁺ cations with the cage. This is also consistent with the ECD measurements that show similar spectra for these two species (data not shown). Thus, it can be claimed that the ECD spectra are not only characteristic of the conformational changes of the host but they are also characteristic of the nature of the interaction that takes place between the metal cations and the two cyclotribenzylene units that constitutes the cryptophane skeleton.

A direct comparison between DFT binding energy (from EDA) and binding constants measured by titration experiments is not appropriate. Indeed, EDA is a very powerful method for a qualitative and quantitative analysis of the chemical bond. This method reports the instantaneous interaction energy between two fragments (the cage and the cation) in the molecular system. In this calculation, neither the full thermodynamics effects nor the solvation effects in reference to experiments are included. Hard cations such as K⁺ and Rb⁺ are probably more solvated that soft cation such as Cs⁺. Consequently, solvation of the cationic species is expected to decrease the electrostatic interaction term more rapidly in the case of the K⁺ and Rb⁺ cations than in the case of the Cs⁺ cation. The solvation of the cage must also be taken into account in our case. It is noteworthy that the presence of water molecules inside the cavity of the host is an important issue to understand the binding properties of the cryptophane derivatives in aqueous solution, especially when cationic species are present within the cavity. The presence of water molecules inside the cavity of cryptophane hosts has already been evidenced both experimentally (X-ray crystallography) and theoretically.^[12,13] The demonstration of the presence of a water molecule inside the

cavity and its role in the binding properties of charged species is very difficult to tackle from an experimental point of view. Experimental procedures based on ITC experiments have been recently proposed to address this problem.^[14] For instance, Eggers and co-workers have suggested treating water molecules as a co-reactant in order to have access to the global desolvation free energy term. This approach is interesting but also time demanding and it is out of the scope of this article. From a theoretical point of view, an implicit solvation model cannot take into account these phenomena. An explicit solvation model including in the case of the cryptophane cage inner water molecules and several outer solvation shells is mandatory, leading to time demanding calculations.

¹³³Cs and ²⁰⁵TI NMR spectra of the complexes. ¹³³Cs and ²⁰⁵TI nuclei are known to be very sensitive to their surrounding environment. In the past, we have reported that both nuclei give rise to very important chemical shift differences between the metallic cation dissolved in the bulk and the cation present within the cavity of the cryptophane hosts.^[1e,1f] These frequency differences were found more important for the TI⁺ cation than for the Cs⁺ cation. These findings are still observed with compound 1, and can be the consequence of several parameters. As mentioned above TI^{*} interacts more strongly with the cage and its polarizability is about twice that of the Cs⁺ cation. Consequently, a small change in the cryptophane structure can induce very large chemical shift differences for these two nuclei. For instance, a comparison of the ¹³³Cs and ²⁰⁵TI NMR spectra of compounds 1 and 4 reveals the high sensitivity of these two nuclei for their immediate environment. At room temperature, a chemical shift difference, $\Delta\delta$, of 10 - 15 ppm is observed between the two $Cs^+@(rac)-1$ and the $Cs^+@(rac)-4$ complexes. This difference is much larger in the case of the ²⁰⁵Tl nuclei, since $\Delta \delta$ = 110 ppm at 335 K (Figure 1).^[15]

It is noteworthy that the high sensitivity of the ²⁰⁵TI nucleus with its immediate environment also explains the large chemical shift differences between TI⁺ in the bulk and the TI⁺@(*rac*)-1 complex. This chemical shift reaches a value $\Delta \delta \sim 1030$ ppm at 335 K. DFT calculations are very helpful and informative to interpret such high chemical shift difference. For instance, in the case of ²⁰⁵TI, these calculations show that relativistic contributions such that spin-orbit coupling is a very important parameter, which has to be taken into account to predict correctly the chemical shift of the TI⁺@1 complex. In the case of the Cs⁺@1 complex, the contribution can be neglected.

The temperature effect observed in the case of the Cs⁺@1 complex is intriguing and counter-intuitive. It is noteworthy that a similar effect also occurs with cryptophanes 2, 3 and 4, which show high affinity for Cs⁺. In contrast, an opposite effect is observed with compound **5** that exhibits the lowest association constant with Cs^{+} .^[6] We have also noticed that the broadening of the Cs⁺ NMR signal observed at lower temperature seems related to the symmetry of the host. Thus, the higher the symmetry of the host the sharper the ¹³³Cs NMR signal at low temperature. For instance, at 278 K the Cs⁺ NMR spectrum of compound 1 (C_1 -symmetry) shows a signal larger than that of compound 2 (D₃-symmetry). DFT calculations reveal that coulombic interactions between Cs^{+} and the phenolate group are important. Considering that the Cs^{+} cation is small with respect to the volume of the inner cavity of the hosts, we can thus assume that Cs⁺ interacts with the different OH groups of the host. Consequently, it explores different regions of the cavity leading to a broadening of the signal at lower temperature. A decrease of the symmetry of the host results in a larger Cs⁺ NMR signal at low temperature. Increasing the temperature results in a mean value of this ¹³³Cs NMR signal. This assumption does not take into account important characteristics

of the cesium nucleus (¹³³Cs has a weak quadrupolar moment) and the possible conformation changes of the host that could take place with a change of the temperature. However, these considerations are not consistent with the fact that compound **5** shows an opposite behavior. In addition, it must be mentioned that a similar temperature effect seems to occur with the ²⁰⁵TI (no quadrupolar moment). Additional work is needed to fully address and understand this surprising effect.

Conclusions

We have described the synthesis of the two enantiomers MM-1 and PP-1 from enantiopure cryptophanes MM-6 and PP-6, respectively. The two enantiomers MM-6 and PP-6 have been isolated by semi-preparative HPLC on chiral stationary phase. Then, subsequent reactions afforded the desired MM-1 and PP-1 derivatives. We have also studied the binding properties of the water-soluble cryptophane 1 in basic aqueous solution in the presence of metallic cations ($M^+ = Rb^+$, Cs^+ , TI^+). The titration experiments (ITC) show a very high affinity of 1 for the Cs⁺ and TI⁺ cations under a large range of experimental conditions. For instance, a binding constant as high as $K = 1.0 \times 10^{10} \text{ M}^{-1}$ has been measured for TI^{\dagger} @1 complex in LiOH/H₂O (0.1 M). Chiroptical techniques (ECD and VCD) have been used to investigate the conformational changes that take place during the encapsulation process. The formation of the Cs⁺@(rac)-1 and TI⁺@(rac)-1 complexes has been also studied via ¹³³Cs and TI NMR spectroscopy, where the low-field signals with respect to the free cations in solution is used to unambiguously detect these two complexes even at high temperature (335 K).

On the other hand, DFT calculations have shown to be a valuable tool to understand the nature of the interaction existing between the cage and the different cations. These calculations suggest that the TI⁺ cation interacts differently with the cage than the other alkali cations studied in this article ($M^+ = K^+$, Rb^+ , Cs^+). For instance, thanks to an energy decomposition analysis, these calculations show the predominance of the electrostatic and orbital interactions in the stabilization of the TI⁺@1 complex. The theoretical investigation of the electronic structure and bonding revealed cation- π interactions in the TI⁺@1 complex. In the alkali metal series, π -cation interactions cannot be clearly identified. In addition, this theoretical work shows that the large chemical shift observed in the case of the TI⁺@1 complex ($\Delta \delta \sim$ 1000 ppm) can be only explain by including relativistic contribution such as spinorbit coupling interactions and a non-negligible charge transfer between TI⁺ and the cage.

The originality in the structure of host **1** lies in the presence of a hydroxyl function that can be used for subsequent reactions. Since the secondary alcohol does not directly participate to the stabilization of the M^+ @**1** complexes, it can be used at a later stage for grafting an organic spacer linked to a solid support. A synthetic work is under way to graft these molecules onto solid supports (superparamagnetic nanoparticles) with the objective of increasing the extraction yield of cesium and thallium cations from aqueous solutions.^[16]

Supporting Information Summary

Experimental section, HPLC and ECD spectra of *MM*-**6** and *PP*-**6**, ¹H and ¹³C NMR spectra of *MM*-**1** and *PP*-**1**, ¹H and ¹³C NMR spectra of *MM*-**7** and *PP*-**7**, isothermal titration calorimetric (ITC) experiments and thermodynamic parameters of the $M^+@1$ complexes ($M^+=Cs^+$ and TI^+), ¹³³Cs NMR spectra of the

¹³³Cs@1 complex at variable temperature, ECD spectra of the Cs⁺@1 and TI⁺@1 complexes, computational details.

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Conflict of Interest

The authors declare no conflict of interest.

Keywords: Calorimetry · Chirality · Circular Dichroism · DFT Calculations • Molecular Recognition

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Supporting Information

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Figure S1: Chromatograms (Chiralpak ID, 250×4.6 mm, EtOH/CH₂Cl₂: 20/80, 1 mL/min) of the collected anti-1 enantiomers after preparative separation on Chiralpak ID (250×10 mm, EtOH/CH₂Cl₂: 20/80, 5 mL/min) by 120 injections of 4.4 mg of racemate. Detection by UV-vis spectroscopy at 254 nm (black chromatograms) and CD spectroscopy at 254 nm (red chromatograms).

Figure S2: ECD spectra of compound *MM*-**6** and *PP*-**6** recorded in $CHCl_3$ (a: top spectrum; c = 1.12 10^{-4} M - 9.04 10^{-5} M) and in CH_2Cl_2 (b: bottom spectrum; c = 9.41 10^{-5} M - 8.88 10^{-5} M). Polarimetric measurements of compounds *MM*-**6** and *PP*-**6** recorded at 25°C in CHCl₃.

Figure S3: ¹H NMR spectrum (300 MHz) of the enantiomer *MM*-**1** recorded in DMSO-*d*₆ at 298 K.

Figure S4: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *MM*-**1** recorded in DMSO- d_6 at 298 K.

Figure S5: ¹H NMR spectrum (300 MHz) of the enantiomer *PP*-**1** recorded in DMSO- d_6 at 298 K.

Figure S6: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *PP*-**1** recorded in DMSO- d_6 at 298 K.

Figure S7: ¹H NMR spectrum (300 MHz) of the enantiomer *PP*-**7** (non-imploded form) recorded in CD₂Cl₂ at 298 K.

Figure S8: ¹H NMR spectrum (300 MHz) of the enantiomer *PP*-**7** (mainly imploded form) recorded in CD_2CI_2 at 298 K. in insert: comparison of the ¹H NMR spectra of the imploded and non-imploded form (aromatic region only).

Figure S9: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *PP*-**7** (non-imploded form) recorded in CD_2CI_2 at 298 K.

Figure S10: ¹H NMR spectrum (300 MHz) of the enantiomer *MM*-**7** recorded in CD₂Cl₂ at 298 K.

Figure S11: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *MM*-**7** (non-imploded form) recorded in CD_2CI_2 at 298 K.

Figure S12: Calorimetric titration of **1** in LiOH/H₂O (0.1 M) at 298 K in presence (or not) of RbCl (top left), CsOH (top right) or TIOAc (bottom). The host solution (top left, $c = 99.3 \mu$ M; top right, $c = 26.0 \mu$ M; bottom, $c = 30.2 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of RbCl (top left c = 1.029 mM), CsOH (top right, c = 0.404 mM) and TIOAc (bottom, c = 0.406 mM) were added at 3 min intervals. The concentration of RbCl was: top right, c = 0.302 mM; bottom, c = 0.598 mM (competition experiments).

Figure S13: Calorimetric titration of **1** in NaOH/H₂O (0.1 M) at 298 K in presence (or not) of RbCI (top left), CsOH (top right) or TIOAc (bottom). The host solution (top left, $c = 100 \mu$ M; top right, $c = 29.9 \mu$ M; bottom, $c = 30.2 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of RbCI (top left c = 1.0 mM), CsOH (top right, c = 0.398 mM) and TIOAc (bottom, c = 0.387 mM) were added at 3 min intervals. The concentration of RbCI was: top right, c = 0.500 mM; bottom, c = 0.911 mM (competition experiments).

Figure S14: Calorimetric titration of **1** in KOH/H₂O (0.1 M) at 298 K in presence CsOH (top) or TIOAc (bottom). The host solution (top, $c = 40.2 \mu$ M; bottom, $c = 75.4 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of CsOH (top, c = 0.400 mM) and TIOAc (bottom, c = 0.693 mM) were added at 3 min intervals.

Figure S15: Calorimetric titration of **1** in LiOH/H₂O (1 M) at 298 K in presence (or not) of RbCl (top left), CsOH (top right) or TIOAc (bottom). The host solution (top left, $c = 98.2 \mu$ M; top right, $c = 99.1 \mu$ M; bottom, $c = 29.8 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of RbCl (top left c = 1.029 mM), CsOH (top right, c = 0.986 mM) and TIOAc (bottom, c = 0.493 mM) were added at 3 min intervals.

Figure S16: Thermodynamic parameters ΔH^0 (kcal/mol), ΔS^0 (cal/mol/K) and ΔG^0 (kcal/mol) of the M⁺@1 complexes (M⁺ = Rb⁺, Cs⁺, Tl⁺) in LiOH/H₂O (0.1 M and 1M), NaOH/H₂O (0.1 M) and KOH/H₂O (0.1 M) solutions.

Figure S17: ¹³³Cs NMR spectra of the Cs⁺@(*rac*)-1 complex in NaOD/D₂O (0.1 M) at different temperatures: 278, 288, 298, 308, 318, 328, 338 and 348 K.

Figure S18: ECD spectra recorded at 293 K of *MM*-**1** in LiOH/H₂O (0.1 M) solution (saturated CHCl₃ solution) in presence of different amount of a) top spectra: cesium hydroxide. b) bottom spectra:

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thallium acetate solutions. [Host] was $1.3 \ 10^{-5}$ M for a) [Host] was $1.35 \ 10^{-5}$ M for b). ECD spectra for higher [Cs⁺] ([TI⁺]) are identical to that recorded for 1 eq. of Cs⁺ (TI⁺).

Figure S19: ECD spectra recorded at 293 K of *MM*-1 in LiOH/H₂O (1 M) in presence of different amount of a) top spectra: cesium hydroxide. b) bottom spectra: thallium acetate solutions. [Host] was 1.45 10^{-5} M for a) [Host] was 1.3 10^{-5} M for b). ECD spectra for higher [Cs⁺] ([TI⁺]) are identical to that recorded for 1 eq. of Cs⁺ (TI⁺).

Figure S20: ECD spectra recorded at 293 K of *MM*-1 in KOH/H₂O (0.1 M) solution in presence of different amount of a) top spectra: cesium hydroxide. b) bottom spectra: thallium acetate solutions. [Host] was 1.3 10^{-5} M for a) [Host] was 1.3 10^{-5} M for b) . ECD spectra for higher [Cs⁺] ([TI⁺]) are identical to that recorded for 1 eq. of Cs⁺ (TI⁺).

S25 – S28: DFT calculations (Geometry, Energy decomposition analysis, Non Covalent Interaction, Orbital mixing, Nuclear magnetic shielding).

Experimental Section

Separation of the two enantiomers of 6. The two enantiomers of 6 were separated on a semipreparative Chiralpak-ID ($250 \times 10 \text{ mm}$) chromatographic column. A mixture of Ethanol/CH₂Cl₂ (20:80) was used as a mobile phase (flow rate: 5 mL/min). UV detection was performed at 254 nm. 550 mg of racemic compound 6 have been dissolved in 15 mL of dichloromethane and 125 injections (120μ L every 3 minutes) have been necessary to complete the experiment. The first eluted enantiomer has been collected between 3.3 and 4 minutes and the second one between 4.4 and 5.6 minutes. 270 mg of the first eluted enantiomer ((-)-enantiomer) has been isolated with an enantiomeric excess ee > 99 %. Then, 270 mg, of the second eluted enantiomer ((+)-enantiomer) has been isolated with an enantiomeric excess ee > 99 %.

Synthesis of compound PP-7. Freshly prepared lithium diphenylphosphide (1 M; 6.2 mL) was added under argon to a stirred solution of cryptophane MM-6 (250 mg, 0.26 mmol) in THF (3 mL). The red mixture was stirred for 48 hours at 60°C. Then, it was poured in water. The aqueous phase was washed five times with CH₂Cl₂ (50 mL). The aqueous phase was then collected and cooled to 0°C. Acidification with conc. HCl solution gives rise to a white precipitate, which was collect on a frit. The white solid was washed several times with water and dried on air. Finally, the solid was washed with diethyl ether to give compound MM-1 as a white powder (0.21 g; 97 %). This compound contains trace of impurities and an additional purification step is necessary. Thus, compound MM-1 (0.21 g, 0.25 mmol) was added in a 25 mL round bottom flask with 7 mL of dry pyridine. The flask was cooled to 0°C. Then, anhydride acetic (1.4 mL) was added under an argon atmosphere. After addition, the flask was allowed to reach room temperature and the mixture was stirred for 5 hours at room temperature. Then, the solution was poured in a mixture of CH_2CI_2 and H_2O . The aqueous phase was extracted three times with CH₂Cl₂. The combined organic phase were washed once with water and then dried over Na₂SO₄. Evaporation of the solvent under reduced pressure gives rise to a solid residue, which was purified on silica gel (CH₂Cl₂/Acetone: 90/10). Different fractions were evaporated to give compound *PP-***7** as a white glassy solid (0.22 g; 78 %). ¹H NMR (300 MHz, CD₂Cl₂, 25°C) δ 6.84 (s, 2 H), 6.90 (s, 4H), 6.85 (s, 1 H), 6.84 (s, 1 H), 6.82 (s, 1 H), 6.81 (s, 1 H), 6.69 (s, 1 H), 6.66 (s, 1 H), 5.15 (m, 1 H), 4.625 (d, J = 13.8 Hz, 4 H), 4.61 (d, J = 13.8 Hz, 2 H), 4.30-3.85 (m, 12 H), 3.51 (d, J = 13.8 Hz, 2 H), 3.495 (d, J = 13.8 Hz, 2 H), 3.49 (d, J = 13.8 Hz, 2 H), 2.34 (s, 9 H; OCH₃), 2.32 (s, 3 H; OCH₃), 2.31 (s, 3 H; OCH₃), 2.30 (s, 3 H; OCH₃), 2.07 ppm (s, 3 H; OCH₃); ¹³C NMR (75.4 MHz,

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CH₂Cl₂, 25°C) δ 171.4 (2C), 170.1 (4C), 169.9 (2C), 150.4 (2C), 150.3, 150.2, 149.9, 149.6, 141.9 (2C), 141.8, 141.7, 140.6, 140.3, 139.7, 139.4, 139.3, 139.2, 138.7, 138.4, 135.0, 134.9 (2C), 134.6, 133.5, 133.2, 126.2, 126.1, 125.7, 125.6, 125.2, 124.6, 122.0, 121.6, 121.4, 121.3, 115.9, 115.3, 71.9, 70.8 (3C), 70.4, 66.2, 65.7, 37.8, 37.7, 37.4 (4C), 22.2 (6C), 22.0 ppm. The ¹H and ¹³C NMR spectra of compound *MM*-7 are identical to that of *PP*-7.

Synthesis of MM-1. A solution of KOH/H₂O (0.5 M; 7mL) was added in one portion to compound *PP*-**7** (0.22 g, 0.19 mmol) in THF (7 mL). The mixture was overnight stirred at 50°C under an argon atmosphere. Then, THF was removed under reduced pressure. 7 mL of water were added and the flask was cooled to 0°C. Acidification with few drops of conc. HCl gives rise to a white solid, which was collected on a frit and washed several times with water. The solid was then dried on the frit and washed with diethyl ether to give compound *MM*-**1** as a beige solid (0.16 g; 98 %). ¹H NMR (300 MHz, DMSO-*d*₆, 25°C) δ 8.57 (s, 1 H), 8.48 (s, 1 H), 8.43 (s, 1 H), 8.31 (s, 1 H), 8.14 (s, 1 H), 7.91 (s, 1 H), 6.77 (s, 1 H), 6.74 (s, 1 H), 6.69 – 6.62 (m, 8 H), 6.55 (s, 1 H), 6.54 (s, 1H), 5.31 (s, 1 H), 4.50 – 3.70 (m, 18 H), 3.22 (m, 6 H). ¹³C NMR (75.4 MHz, DMSO-*d*₆, 25°C) δ 146.4, 146.35, 145.7, 145.6, 145.5, 145.4, 145.3, 145.1, 144.5, 144.3 (2 C), 144.1, 133.9, 133.8, 133.6, 133.5, 133.3 (2 C), 131.1 (2 C), 130.5 (2 C), 129.5, 128.9, 120.2, 119.8, 119.0, 118.1 (2 C), 118.0, 117.6, 117.5, 117.4, 117.0, 116.9, 116.5. 71.5, 70.5, 69.2, 68.5 (3C), 68.2, 68.1, 35.1, 34.9, 34.8 HRMS m/z [M+H]⁺ calcd. for C₄₉H₄₅O₁₃ 841.2892 found. 841.2855.

Using the same synthetic approach derivative *PP*-**6** (0.24 g, 0.25 mmol) gives rise to *MM*-**7** (0.2 g; 71 %, 2 steps). In turn, hydrolysis of compound *MM*-**7** under basic solution gives compound *PP*-**1** (0.13 g; 88 %) as a beige solid. Spectroscopic data are identical to those reported for the *MM*-**1** enantiomer.

ECD measurements. ECD spectra were recorded at 293 K using 0.2 cm path length quartz cells. The concentration of the *MM*-1 derivative was in the range (5.0 $10^{-5} - 10^{-5}$ M) in basic LiOH/H₂O or KOH/H₂O solutions. Additional ECD spectra were recorded at higher LiOH/H₂O concentration (1 M) or in the presence of a competitor (saturated LiOH/H₂O solution with CHCl₃). Spectra were recorded in the 220 – 400 nm spectral range with a 0.5 nm increment and a 1 s integration time. Spectra were processed with standard spectrometer software and baseline corrected without smoothing. Spectral units are expressed in difference in molar extinction coefficients.

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VCD measurements. The IR and VCD spectra were recorded on a FTIR spectrometer equipped with a VCD optical bench.^[1] following the experimental procedure previously published. ^[2] Samples were held in a CaF₂ cell with a fixed path length of 45 μ m. IR and VCD spectra of the two enantiomers of **1** were measured in basic NaOD/D₂O (0.214 M) solution at a concentration of 0.030 M. Additional spectra were recorded in the presence of a very small amount of CDCl₃ and CsOH.

ITC experiments. Isothermal Titration Calorimetry (ITC) experiments were performed at 298 K. In a typical experiment, the host solution (~ 0.1 mM) in LiOH/H₂O (0.1 M) or KOH/H₂O (0.1 M) was placed in the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of guest solution (10 times more concentrated) were added via a computer-automated injector at 3 min intervals. Heat changes were recorded after each addition. Heats of dilution were measured by a blank experiment (in the absence of host) under the same conditions, and they were subtracted from the titration data prior to curve fitting. The first injection was discarded from each data set to remove the effect of guest diffusion across the syringe tip during the equilibration process. Titration curves were fitted with the one binding set model. In the case of very high binding for the Cs⁺@(*rac*)-1 and Tl⁺@(*rac*)-1 complexes, competition experiments were necessary. In our case, RbCl was a good competitor since it shows lower affinity with host 1. Thus, prior to measure enthalpogram for the Cs⁺@1 and the Tl⁺@1 complexes, the association constant K and the enthalpy of complexation ΔH^0 of the Rb⁺@1 complex were measured. Additional ITC experiments were performed at higher LiOH/H₂O (1 M) concentration under the same conditions. In this case, competition experiments were not necessary.

NMR experiments. ¹³³Cs NMR spectra were recorded on an 11.7 T Bruker Avance spectrometer using a 5 mm broadband probehead. A solution of cesium nitrate was used as a chemical shift reference (0 ppm). A delay time of 1s was applied between each pulse. At 298 K the delay time was 5 s. ¹³³Cs NMR spectra were also recorded at different temperature (278 - 348 K). The ²⁰⁵TI NMR experiments were performed on a Bruker DMX300 spectrometer (7 Tesla) equipped with a 5 mm BB Nalorac probehead recovered from a 11.7 Tesla spectrometer. First, a solution of thallium nitrate was used as a chemical shift reference (0 ppm). Then, the other spectra were recorded by part. The interscan delay was 0.9 s. For all the displayed ²⁰⁵TI NMR spectra the FIDs were processed by an exponential window of 200 Hz before Fourier transformation.

DFT Calculations. DFT calculations were performed using the ORCA and ADF program packages. ^{[3],[4]} To calculate NMR parameters, the two-component method of relativistic quantum chemistry was used in the framework of the zeroth-order regular approximation (ZORA) and all-electron basis sets.^[5] The non-covalent interactions were analyzed and visualized with MULTIWFN and VMD programs.^{[6],[7]} A comprehensive description of the used methods is provided in Supporting Information.

Analytical chiral HPLC separation for compound 6



Figure S1: Chromatograms (Chiralpak ID, 250×4.6 mm, EtOH/CH₂Cl₂: 20/80, 1 mL/min) of the collected anti-**6** enantiomers after preparative separation on Chiralpak ID (250×10 mm, EtOH/CH₂Cl₂: 20/80, 5 mL/min) by 120 injections of 4.4 mg of racemate. Detection by UV-vis spectroscopy at 254 nm (black chromatograms) and CD spectroscopy at 254 nm (red chromatograms).



Figure S2: ECD spectra of compound *MM*-**6** and *PP*-**6** recorded in $CHCl_3$ (a: top spectrum; c = 1.12 10^{-4} M - 9.04 10^{-5} M) and in CH_2Cl_2 (b: bottom spectrum; c = 9.41 10^{-5} M - 8.88 10^{-5} M). Polarimetric measurements of compounds *MM*-**6** and *PP*-**6** recorded at 25°C in CHCl₃.



Figure S3: ¹H NMR spectrum (300 MHz) of the enantiomer *MM*-**1** recorded in DMSO-*d*₆ at 298 K.



Figure S4: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *MM*-1 recorded in DMSO- d_6 at 298 K.



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Figure S5: ¹H NMR spectrum (300 MHz) of the enantiomer *PP*-**1** recorded in DMSO-*d*₆ at 298 K.



Figure S6: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *PP*-**1** recorded in DMSO- d_6 at 298 K.



Figure S7: ¹H NMR spectrum (300 MHz) of the enantiomer PP-7 (non-imploded form) recorded in CD_2CI_2 at 298 K.



Figure S8: ¹H NMR spectrum (300 MHz) of the enantiomer *PP*-**7** (mainly imploded form) recorded in CD_2CI_2 at 298 K. in insert: comparison of the ¹H NMR spectra of the imploded and non-imploded form (aromatic region only).



Figure S9: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *PP*-7 (non-imploded form) recorded in CD_2Cl_2 at 298 K.



Figure S10: ¹H NMR spectrum (300 MHz) of the enantiomer *MM*-7 recorded in CD₂Cl₂ at 298 K.

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Figure S11: ¹³C NMR spectrum (75.47 MHz) of the enantiomer *MM*-7 (non-imploded form) recorded in CD_2CI_2 at 298 K.



Figure S12: Calorimetric titration of **1** in LiOH/H₂O (0.1 M) at 298 K in presence (or not) of RbCl (top left), CsOH (top right) or TIOAc (bottom). The host solution (top left, $c = 99.3 \mu$ M; top right, $c = 26.0 \mu$ M; bottom, $c = 30.2 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of RbCl (top left c = 1.029 mM), CsOH (top right, c = 0.404 mM) and TIOAc (bottom, c = 0.406 mM) were added at 3 min intervals. The concentration of RbCl was: top right, c = 0.302 mM; bottom, c = 0.598 mM (competition experiments).



Figure S13: Calorimetric titration of **1** in NaOH/H₂O (0.1 M) at 298 K in presence (or not) of RbCI (top left), CsOH (top right) or TIOAc (bottom). The host solution (top left, $c = 100 \mu$ M; top right, $c = 29.9 \mu$ M; bottom, $c = 30.2 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of RbCI (top left c = 1.0 mM), CsOH (top right, c = 0.398 mM) and TIOAc (bottom, c = 0.387 mM) were added at 3 min intervals. The concentration of RbCI was: top right, c = 0.500 mM; bottom, c = 0.911 mM (competition experiments).



Figure S14: Calorimetric titration of **1** in KOH/H₂O (0.1 M) at 298 K in presence CsOH (top) or TIOAc (bottom). The host solution (top, $c = 40.2 \mu$ M; bottom, $c = 75.4 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of CsOH (top, c = 0.400 mM) and TIOAc (bottom, c = 0.693 mM) were added at 3 min intervals.



Figure S15: Calorimetric titration of **1** in LiOH/H₂O (1 M) at 298 K in presence (or not) of RbCl (top left), CsOH (top right) or TIOAc (bottom). The host solution (top left, $c = 98.2 \mu$ M; top right, $c = 99.1 \mu$ M; bottom, $c = 29.8 \mu$ M) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of RbCl (top left c = 1.029 mM), CsOH (top right, c = 0.986 mM) and TIOAc (bottom, c = 0.493 mM) were added at 3 min intervals.

M^+	LiOH/H ₂ O (0.1 M)	NaOH/H ₂ O (0.1 M)	KOH/H ₂ O (0.1 M)	LiOH/H ₂ O (1 M)
Rb ⁺	$\Delta H^0 = -5.1$ $\Delta S^0 = +6.3$ $\Delta G^0 = -7.0$	$\Delta H^0 = -4.4$ $\Delta S^0 = 10.0$ $\Delta G^0 = -7.4$	Not measured	$\Delta H^0 = -3.6$ $\Delta S^0 = +2.8$ $\Delta G^0 = -4.5$
Cs ⁺	$\Delta H^0 = -11.8$ $\Delta S^0 = -0.2$ $\Delta G^0 = -11.7$	$\Delta H^0 = -11.0$ $\Delta S^0 = + 3.6$ $\Delta G^0 = -12.1$	$\Delta H^0 = -6.6$ $\Delta S^0 = +9.8$ $\Delta G^0 = -9.6$	$\Delta H^0 = -11.5$ $\Delta S^0 = -8.2$ $\Delta G^0 = -9.1$
Tl ⁺	$\Delta H^0 = -10.4$ $\Delta S^0 = +10.8$ $\Delta G^0 = -13.6$	$\Delta H^0 = -10.2$ $\Delta S^0 = +11.7$ $\Delta G^0 = -13.7$	$\Delta H^0 = -5.9$ $\Delta S^0 = +15.8$ $\Delta G^0 = -10.6$	$\Delta H^0 = -10.9$ $\Delta S^0 = -2.2$ $\Delta G^0 = -10.2$

Figure S16: Thermodynamic parameters ΔH^0 (kcal mol⁻¹), ΔS^0 (cal mol⁻¹ K⁻¹) and ΔG^0 (kcal mol⁻¹) of the M⁺@1 complexes (M⁺ = Rb⁺, Cs⁺, Tl⁺) in LiOH/H₂O (0.1 M and 1M), NaOH/H₂O (0.1 M) and KOH/H₂O (0.1 M) solutions.



Figure S17: ¹³³Cs NMR spectra of the Cs⁺@(*rac*)-1 complex in NaOD/D₂O (0.1 M) at different temperatures: 278, 288, 298, 308, 318, 328, 338 and 348 K.





Figure S18: ECD spectra recorded at 293 K of *MM*-**1** in LiOH/H₂O (0.1 M) solution (saturated with CHCl₃) in presence of different amount of a) top spectra: cesium hydroxide. b) bottom spectra: thallium acetate solutions. [Host] was 1.3 10^{-5} M for a) [Host] was 1.35 10^{-5} M for b). ECD spectra for higher [Cs⁺] ([TI⁺]) are identical to that recorded for 1 eq. of Cs⁺ (TI⁺).



Figure S19: ECD spectra recorded at 293 K of *MM*-1 in LiOH/H₂O (1 M) in presence of different amount of a) top spectra: cesium hydroxide. b) bottom spectra: thallium acetate solutions. [Host] was 1.45 10^{-5} M for a) [Host] was 1.3 10^{-5} M for b). ECD spectra for higher [Cs⁺] ([TI⁺]) are identical to that recorded for 1 eq. of Cs⁺ (TI⁺).



Figure S20: ECD spectra recorded at 293 K of *MM*-1 in KOH/H₂O (0.1 M) solution in presence of different amount of a) top spectra: cesium hydroxide. b) bottom spectra: thallium acetate solutions. [Host] was 1.3 10^{-5} M for a) [Host] was 1.3 10^{-5} M for b). ECD spectra for higher [Cs⁺] ([Tl⁺]) are identical to that recorded for 1 eq. of Cs⁺ (Tl⁺).

Computational details

Geometry

Geometry optimizations of the M^+ (M=K, Rb, Cs, TI) encapsulated in the cryptophane cage were performed using density functional theory (DFT) with the ORCA 3.03 program package along with the Resolution of Identity approximation.[8,9] The near-field Coulomb interactions are evaluated employing the RI-J approximation. It speeds up calculation of the Coulomb term for large systems. The B97-D functional was considered corrected by the empirical dispersion term (D3) proposed by Grimme et al. with a finite damping as introduced by Becke and Johnson (BJ-damping) to account for the London dispersion energy. [10,11] The addition of the BJ-damping provides better results for nonbonded distances and more clear effects of intramolecular dispersion. The split valence plus polarization triple- ζ (def2-TZVP) basis set was used for H, Li, C, O. [12] For the alkali metal series and TI atom, we used the segmented all-electron relativistically contracted (SARC) basis sets in conjunction with a scalar relativistic Hamiltonian. [13] In the present work, we applied two-component relativistic DFT approach based on the zero-order regular approximation (ZORA). [14]

Energy decomposition analysis (EDA)

Analysis of the chemical interactions can be performed combining a fragment approach to the molecular structure of the complex with the decomposition of the total bonding energy into separate components familiar to chemists:

$$\Delta E_{int} = \Delta E_{el.} + \Delta E_{rep.} + \Delta E_{orb.}$$

The interaction energy $E_{int.}$ between two fragments A and B in a molecular system A-B is partitioned in three terms, namely, (1) the quasiclassical electrostatic interaction $E_{el.}$ between the fragments, (2) the repulsive exchange (Pauli) interaction $E_{rep.}$ between electrons of the two fragments having the same spin, and (3) the orbital (covalent) interaction $E_{orb.}$, which comes from the orbital relaxation and the orbital mixing between the fragments. [15]

The computations were performed with ADF 2016, ZORA-DFT/PBE-D3(BJ) and TZ2P all-electron relativistic basis sets at the ORCA geometries. [16] The fragment M^+ and the cryptophane cage were considered for the EDA.

Non Covalent Interaction (NCI)

Johnson and co-workers recently introduced a method for studying non-covalent interactions (NCIs) based on the electron density (ρ), the reduced gradient of the density ($s = |\nabla \rho|/(2(3\pi^2)^{1/3}\rho^{4/3})$, and the Laplacian of the density ($\nabla^2 \rho$).[17] The approach allows one to identify the interactions in real-space, thus enabling a graphical visualization of the regions where non-covalent interactions occur. To recover most of the interaction in the real space, explicitly correlated methods are required. In this work, we performed single-point DLPNO-CCSD(T) calculations with ORCA 3.03 program package and the relativistic Hamiltonian and basis set described previously. [18]

The non-covalent interaction plots were produced with MULTIWFN and VMD. [19,20]

Orbital mixing



Figure a: mixing of the 6p(TI) (left) and the 6s(TI) (right) orbitals with the cryptophane orbitals



Figure b: mixing of the 5p(Cs) orbital with the cryptophane orbitals

Nuclear magnetic shielding

The M⁺ isotropic shielding was calculated through the DFT/GIAO shielding tensor, the formal definition of its components being:[21]

$$\sigma_{\alpha\beta} = \frac{\partial^2 E}{\partial \mu_{\alpha} \delta B_{\beta}}$$

where *E* is the total electronic energy of the molecule; *B* is the external magnetic field and μ is the magnetic moment of the nucleus of interest.

The M⁺ nuclear magnetic shielding tensor was computed at the ORCA geometries with DFT and a PBE functional using Gauge-Including Atomic Obitals (GIAO) method as implemented in ADF 2016.[22] Slater-type orbitals (STOs) were employed as basis functions in SCF calculations. The allelectron valence triple zeta, polarized relativistic basis set TZ2P was used for all atoms. Scalar and spin-orbit coupling relativistic effects were considered using the zero-order regular approximation (ZORA). In practical applications calculations using GIAOs usually produce better results with small or medium basis sets that are mandatory for our large size systems.

To reduce systematic errors in calculating the NMR chemical shifts, a secondary reference was used. The NMR chemical shifts (in ppm) were obtained using the equation:

$$\delta_i = \sigma_{ref.} - \sigma_i + \delta_{ref.}$$

where $\sigma_{ref.}$ is the calculated NMR shielding of the corresponding nucleus in the reference compound (M⁺ in vacuum), σ is the calculated NMR shielding of the investigated nucleus (M⁺ in the complex), and $\delta_{ref.}$ is the experimental NMR chemical shift of the secondary reference relative to the primary standard. For TI⁺ and Cs⁺ the secondary references are TINO₃ and CsNO₃ in D₂O respectively. For TI⁺ $\delta_{ref.}$ =340 ppm and for Cs⁺ $\delta_{ref.}$ = 0.[23]

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